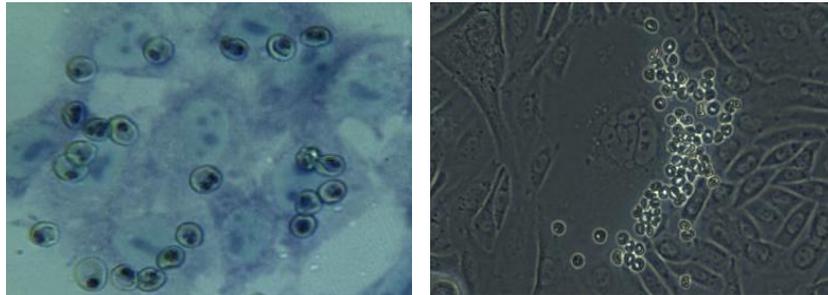


**Analyses of the transcriptome profiles of *Plasmodium falciparum* infected erythrocytes selected for binding to the human endothelial receptors (ICAM-1, P-selectin, E-selectin, CD9 and CD151)**



Dissertation with the aim of achieving a doctoral degree  
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I hereby declare, on oath, that I have written the present dissertation by my own and have not used other than the acknowledged resources and aids.

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signature

*Nahel Cebel*

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## Summary

*P. falciparum* is responsible for most of the morbidity and mortality accompanying malaria infections in humans. This is attributed to the ability of this parasite to evade the immune system and to sequester in small blood vessels of vital organs within its human host. This sequestration leads to serious life threatening complications, such as cerebral malaria, especially in children younger than five years of age. Cytoadhesion, that is required for sequestration, depends on the interaction between a human endothelial receptor and a matching parasite ligand. Until now, a panel of receptors (22 receptors) have been identified to interact with *P. falciparum* infected erythrocytes (IEs). The parasite's interactions to only four host receptors (CD36, ICAM-1, CSA and EPCR) were studied in detail, The main parasite ligand responsible for the cytoadhesion of mature trophozoites is the so-called *PfEMP1*, which is encoded by the *var* multigene family. *var* genes are mutually exclusively expressed and the parasite can switch expression between the about 60 copies present in its genome. The corresponding mutually expressed *var* gene is decisive for the parasite's tropism to the endothelial cells. The enormous diversity of *PfEMP1* domain structures makes it difficult to identify the binding domains for every receptor. This study aimed to analyse the transcriptome profile of the parasites that are able to bind to one out of these five different endothelial receptors (ICAM-1, P-selectin, E-selectin, CD9 and CD151) as a first step to identify their *PfEMP1*'s ligands. In order to achieve this aim, IEs were enriched by binding to the corresponding receptor for several rounds. The illumina NGS platform was used for mRNA sequencing and finally gene differential analysis was performed using a generalized lineal model via the DEseq Bioconductor package. The results confirmed the known ligands of ICAM-1. Two interesting group A *var* candidates were identified to be differentially expressed in the parasite population enriched by binding to P-selectin (IT4\_var02/07). Upon enrichment over CD9, eight *var* genes were differentially expressed; with four of them being group A *var* genes (IT4\_var02/64/09/07). No peremptory ligand was identified for both E-selectin and CD151. In conclusion, this study anticipated P-selectin and CD9 interactions with *PfEMP1*s' ligands as a new way to reveal more details about the exact mechanisms underlying cerebral malaria. These need more efforts to be investigated for further studies to exactly characterize the binding domain(s) and finally to develop an inhibitory mechanism.

## Abbreviations

<b>ATCC</b>	<b>American type cell culture</b>
<b>BNITM</b>	Bernhard Nocht Institute for tropical Medicine
<b>bp</b>	Base pair
<b>BSA</b>	Bovines serum albumin
<b>CD</b>	Cluster of Differentiation
<b>CHO</b>	Chinese Hamster Ovary
<b>CIDR</b>	Cysteine Rich Interdomain Region
<b>CM</b>	Cerebral malaria
<b>CO<sub>2</sub></b>	Carbon dioxide
<b>CSA</b>	Chondroitin Sulfat A
<b>CSP</b>	Circumspozoite protein
<b>CT</b>	C-Terminus
<b>DBL</b>	Duffy Binding Like
<b>DC</b>	Domain cassette
<b>DE</b>	Differential expression
<b>DMSO</b>	Dimethylsulfoxid
<b>DNA</b>	Deoxyribonucleic acid
<b>EGF</b>	Epidermal growth factor
<b>EPCR</b>	Endothelial protein C receptor
<b>EtOH</b>	Ethanol
<b>FACS</b>	Fluorescent activated cell sorting
<b>FDR</b>	False detection rate
<b>Fig</b>	Figure
<b>g</b>	Gramm
<b>GFP</b>	Green flourescent protein
<b>GLM</b>	Generalised lineal model
<b>hpi</b>	hours post invasion
<b>ICAM-1</b>	intercellular adhesion molecule 1
<b>IE<sub>(s)</sub></b>	Infected erythrocyte(s)
<b>IFA</b>	Immuno flouresemce analysis
<b>iFCs</b>	Inactivated fetal calf serum
<b>IL</b>	Interleukines
<b>KAHRP</b>	Knob associated histidine rich protein
<b>kDa</b>	Kilodalton
<b>L</b>	Liter
<b>MC</b>	Maurer's clefts
<b>MFS</b>	Malaria Freezing Solution
<b>ml</b>	Milliliter
<b>MSP</b>	Merozoite surface protein
<b>MTS</b>	Malaria Thawing Solution
<b>NGS</b>	Next generation sequencing
<b>nM</b>	Nanomolar
<b>NTS</b>	N-terminales Segment
<b>O<sub>2</sub></b>	oxygen
<b>OsO<sub>4</sub></b>	Osmium tetroxide

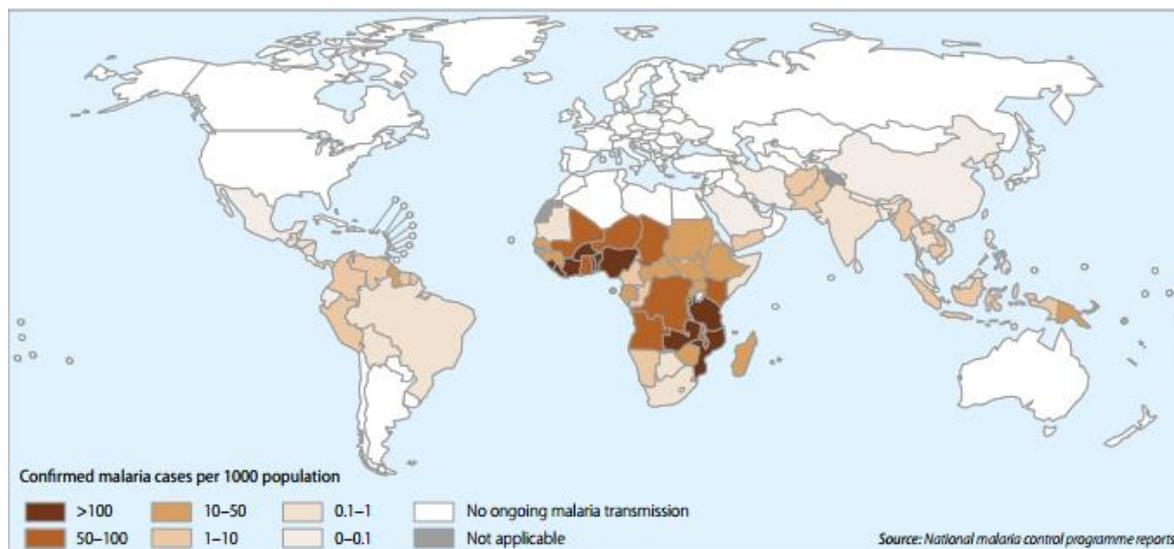
<i>P.</i>	<i>Plasmodium</i>
<b>Padj</b>	P-value adjustment
<b>PBS</b>	Phosphate Buffered Saline
<b>PECAM-1</b>	Platelet endothelial cell adhesion molecule
<b>PEXEL</b>	<i>Plasmodium</i> Export Element
<i>PfEMP1</i>	<i>Plasmodium falciparum</i> Erythrocyte Membrane Protein 1
<i>PfMC-2TM</i>	<i>Plasmodium falciparum</i> Maurer's clefts-2 Transmembrane
<i>PfMSP7</i>	<i>Plasmodium falciparum</i> merozoite surface protein
<b>PNEPs</b>	PEXEL- negative exported proteins
<b>PTEX</b>	<i>Plasmodium</i> translocon of exported proteins
<b>PV</b>	Parasitophorous vacuole
<b>PVM</b>	Parasitophorous Vacuole membrane
<b>QC</b>	Quality control
<b>REX1</b>	Ring exported protein 1
<i>rif</i>	Repetitive Interspersed Family
<b>RIN</b>	RNA integrity number
<b>RNA</b>	Ribonucleic acid
<b>RPMI</b>	Roswell Park Memorial Institute (cell culture medium)
<b>RSP</b>	Ring surface protein
<b>SBP1</b>	Skeleton binding protein 1
<i>stevor</i>	Subtelomeric Variable Open Reading Frame
<b>Supp.</b>	Supplementary
<i>surf genes</i>	Surface-associated interspersed genes (Coding for SURFIN)
<b>TNF<math>\alpha</math></b>	Tumor necrosis factor alpha
<b>UA</b>	Uranyl acetate
<b>UPS</b>	Upstream
<i>var</i>	variant gene
<b>VCAM-1</b>	Vascular cell adhesion molecule-1
<b>VSA</b>	Variant surface antigen
<b>VSG</b>	Variant surface glycoprotein
<b>WHO</b>	World Health Organisation
<b>x g</b>	gravity
<b><math>\alpha</math></b>	Anti
<b><math>\mu</math>g</b>	Microgram
<b><math>\mu</math>l</b>	Microliter

## 1. Introduction

### 1.1. Malaria Disease

The name of the disease is derived from Italian word “mala aria” or bad air due to ancient association of the disease with swampy and marshy areas. At that time, the causative organism was not known. The parasite which causes malaria was discovered in 1880 by Charles Louis Laveran in a patient sample. In 1898, the mosquito vector was discovered by Ronald Ross who won the Nobel prize for that discovery (Cox 2010).

Tropical and subtropical areas are affected with malaria (Fig 1.1). According to WHO, 214 million cases were reported globally in 2015. Malaria caused 438 000 deaths from which 306 000 deaths are children under 5 years of age, mainly in Sub-Saharan Africa. Malaria is caused by Apicomplexan parasites of genus *Plasmodium* and is transmitted by the bite of a female *Anopheles* mosquito. Humans are infected by four malaria species which are *P. falciparum*, *P. vivax*, *P. malariae* and *P. ovale*. Recently, zoonotic transmission of *P. knowlesi* (monkey malaria) was also recorded. *P. falciparum* and *P. vivax* cause the majority of cases. *P. falciparum* is the most pathogenic form which is considered the main cause of severe malaria (WHO 2015). It is difficult to enclose the exact manifestations of malaria; because it differs from one patient to another according to certain risk factors such as age (less than 5 years and greater than 65 years of age both are at risk of developing SM), pregnancy, immune status or any coexisting medical condition (Wassmer et al. 2015). Symptoms range from asymptomatic parasitemia to severe life threatening disease. In uncomplicated malaria (UM), patient suffers from influenza like symptoms such as fever, chills and headache which may be misdiagnosed (Lee et al. 2002). In case of severe malaria (SM), symptoms deteriorate within 3-7 days after onset of fever. The complications of SM are, cerebral malaria (CM), pulmonary edema, acute renal failure and severe anaemia (Philpott and Keystone 1987, Wassmer et al. 2015) in addition to metabolic complications, such as acidosis and hypoglycaemia (Trampuz et al. 2003). The severity of disease may be exaggerated by both higher parasitemia and IEs sequestration in specific tissues. In endemic areas, SM occurs mainly in young children (1 month to 5 years of age) whereas in industrialized countries, it occurs in nonimmune travellers returning from endemic areas (Trampuz et al. 2003, Suh et al. 2004).



**Fig. 1.1. Countries with continuous transmission of malaria (according to WHO report 2014).** approximately 3.3 billion people in 97 countries are at risk of being infected with malaria and 1.2 billion are at high risk.

### 1.1.1. Treatment of malaria

Treatment of malaria was considered a difficult mission in the last decades. The difficulty was to design a treatment plan that act to prevent the progression of the infection to severe life threatening syndrome and at the same time could prevent the spreading of the infection within the population. On the other hand, the complexity of the life cycle of malaria parasites made it difficult to target the different blood stages (sexual and asexual) as well as hypnozoites in case of *P. vivax*. For long time, the first line of treatment was quinine which can only kill the schizont stage. The development of quinine resistance especially by *P. falciparum*, brought the need for another drugs which were used in combination with quinine such as chloroquine and quinidine (Castelli et al. 2012, Labbe et al. 2001, Laloo and Bell 2010). A strong anti-parasite Chinese malarial remedy known as artemisinin was reported scientifically at the end of the 20<sup>th</sup> century, it acts on both asexual and sexual blood stages (Adjuik et al. 2004). Artemisinin was considered an ideal antimalarial drug until *P. falciparum* developed resistance to it. Resistance was also reported in *P. vivax* but in lower frequency and it is extremely rare in other species (Pelfrene et al. 2015).

The problem in SM patients is that, the antimalarial drugs take about 24 hours to kill the parasites which might be a long period during which most deaths occur (Rowe et al. 2009). Based on these facts, it becomes imperative to develop adjunctive therapies to be used in SM cases. For instance, targeting the cytoadhesion process which causes most of *P. falciparum* life-threatening complications.

### **1.1.2. Malaria vaccines**

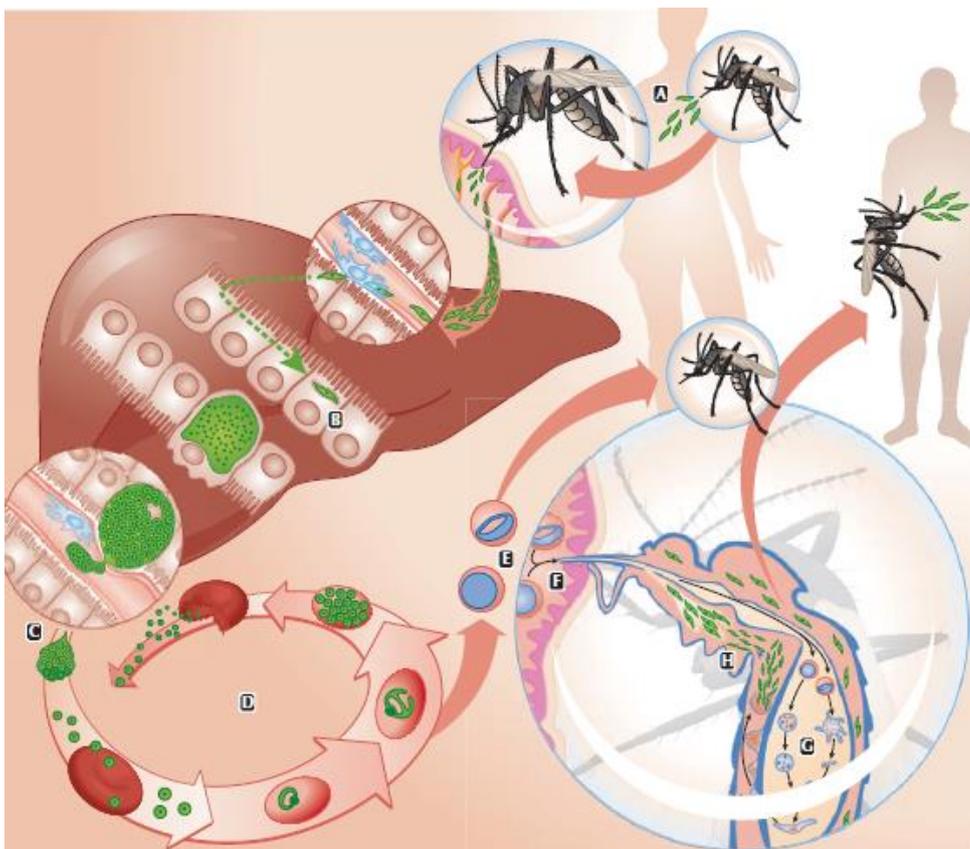
Researchers are hoping to develop a vaccine against malaria. The fact that malaria immunity can be acquired through natural infections is a strong evidence for the possibility to reach this aim (Hviid 2007). Until now, antimalarial vaccines are categorized into three main types (Arama and Troye-Blomberg 2014); First, transmission block vaccines, the idea was based on generating vaccines against gametes and zygotes that will be taken by the mosquito during the blood meal and inhibit the further development inside the mosquito's midgut. The problem is the feasibility of this idea because it requires the injection of the whole population with the vaccine to inhibit the transmission (Carter 2001, Carter et al. 2000, Saul 2008).

Second, preerythrocytic vaccine, it was proved that injection with irradiated sporozoites could induce a sterile immunity (Hoffman et al. 2002) in addition to some trials to develop vaccines against CSP (circumsporozoite protein) either recombinant or peptide vaccines (Long and Hoffman 2002, Hoffman et al. 1991, Roestenberg et al. 2012). Mosquirix or RTS,S/AS01 is a promising vaccine candidate engineered from hepatitis B surface antigen virus-like particles, incorporating a portion of the *P. falciparum* CSP protein and a liposome-based adjuvant (Gosling and von Seidlein 2016). Through this vaccine, the parasite will not be able to reach the liver due to the induction of humoral and cellular immunity with high antibody titers (Foquet et al. 2014). It is the first vaccine to be licensed against a parasite. In 2015, the results of a large pivotal trial were published, this trials started in 2009 and ended in 2011 with over 15000 infants and young children showing a promising results for this new vaccine (Rts 2015).

Third, blood stage vaccines, they target the merozoite surface antigens in order to block the erythrocyte invasion. The target antigens are AMA1 (apical membrane antigen 1), EBA-175 (erythrocyte binding antigen 175) and MSP1 (merozoite surface protein 1) which are still under investigations (Crompton et al. 2010, Wang et al. 2009, Goodman and Draper 2010, Holder et al. 1999). Recent studies addressed the possibility of developing a vaccine that targets the VSAs mainly the *PfEMP1s* family, knowing that these antigens play a role in the acquired immunity developed by patients in endemic areas although the sterile immunity is never achieved (Chan et al. 2014). Due to the fact that targeting the correct antigen is not an easy mission, this issue is still under investigations. A recent study was carried on to proof this concept, in which they injected *Aotus* monkeys with DBL1 $\alpha$  domain of MC *P. falciparum* strain which in turn generated antibodies that protected the monkeys from homologous parasites infections. As a result, *PfEMP1* vaccines drawn the attention of the scientific community, where efforts are directed towards the identification of *PfEMP1* domains that are associated with SM to prioritize them as vaccine candidates (Baruch et al. 2002, Chen 2007, Makobongo et al. 2006).

## **1.2. Life cycle of *P. falciparum***

This parasite exhibits a complex life cycle between two hosts and alternating between different developmental stages as well as the ability to reproduce in both sexual and asexual ways (Fig. 1.2). Life cycle starts when an infected female *Anopheles* mosquito takes a blood meal from a human host where it injects the sporozoite forms of the parasite into skin. The sporozoites enter the blood vessels and travel until they reach the liver (Cowman et al. 2012, Prudencio, et al 2006). In the liver, they travel through Kupffer cells and several hepatocytes until they reside in one hepatocyte and form the parasitophorous capsule. The parasite then replicates asexually to thousands of merozoites for 8-10 days. Then in vesicle like components called merozoites, the merozoites leave the liver through the liver sinusoids (Sturm et al. 2006). Upon reaching the blood stream, they rapidly infect erythrocytes. Inside the infected erythrocytes (IEs), the merozoites develop for 48 hours into ring, trophozoite, mature trophozoite and schizont respectively (Cowman et al. 2012).



**Fig. 1.2. Life cycle *P. falciparum*.** Starts with the injection of sporozoites into the skin of human host, then they rapidly move with the blood stream until they reach the liver where they travel through many cells and finally reside in one cell. The intrahepatic phase takes about 8-10 days and ends by the release of merozoites containing merozoites in the circulation. The merozoites rapidly invade the erythrocytes developing into ring, trophozoite and finally schizonts. Rupture of schizonts results in the release of numerous amount of merozoites in the blood stream where they repeat the cycle again. Some parasites then develop into the sexual forms (male and female gametocytes) which are taken with the feeding mosquito where they are fertilized and undergo further development until the sporozoit form reaches the mosquito's salivary, gland ready to infect another host (Portugal et al. 2011).

Then new merozoites are released in to the circulation invading new erythrocytes and repeating the cycle (Prudencio et al. 2006). Some of the merozoites differentiate into sexual forms which are male and female gametocytes. The sexual forms are then taken up by the mosquito during the blood meal where they develop to macro- (female) and micro- (male) gametocytes. In the intestine of the mosquito, fertilization occurs which results in the formation of a diploid zygote (ookinetes) which penetrates the mosquito's mitgut wall developing into oocyst which disrupts and release sporozoites into the mosquito body cavity. The sporozoites then enter the mosquito salivary glands, ready to infect another human host (Portugal et al. 2011, Petter and Duffy 2015, Rowe et al. 2009).

### **1.3. Developmental stages and host cell modifications.**

#### **1.3.1. Asexual stages**

##### **1.3.1.1. Merozoite stage**

A very important stage due to two facts; first, they remain for a while extracellular where they are exposed to the immunity. Second, they are well prepared to escape from the old IEs, attach to a new one and reinvade it thus propagating the disease and the survival of the parasite (Bannister et al. 2000). Merozoites are considered the smallest eukaryotic cell (1-2  $\mu\text{m}$ ) (Fig 1.3C). They are ovoid in shape with a flat apical prominence at one end (apical complex). The merozoite represents a typical apicomplexan which imposes an apical complex, mitochondrion, nucleus and a plastid. The apical complex encounters secretory organelles which are the micronemes, rophtries and dense granules (Fig. 1.3) (Langreth et al. 1978). A short description for multistep invasion process is as follow, the mature merozoites are egressed from the schizont, the merozoite then comes to contact with a circulating erythrocyte followed by a reorientation so that the merozoite apex faces the erythrocyte membrane (Gilson and Crabb 2009). Finally, the merozoite enters the erythrocyte. Shortly after invasion, echinocytosis takes place to seal the site of invasion then the erythrocyte restores its original shape within 10 minutes (Koch and Baum 2016).

The mechanism by which the merozoites attach to the surface of erythrocytes is mediated by a protein family called merozoite surface proteins (MSP) which covers the whole merozoite. The best characterized candidate is MSP1 which is able to interact with proteins on the erythrocyte called Band 3 or glycoprotein A. Studies also showed that it is not the only protein that acts during the attachment, others are MSP6/7 and MSPDBL1-2 (Cowman and Crabb 2006, Jaskiewicz et al. 2010). During invasion rophtries, micronemes discharge their contents to help in the formation of PV (parasitophorous vacuole).

The PV membrane (PVM) is considered as a parasite–host cell mediator. A complicated tubovesicular network was found to extrude from PVM to the erythrocyte cytoplasm. Its function is to help the nutrient uptake by the parasite (Elmendorf and Haldar 1993, Lauer et al. 1997). After entry into the erythrocyte, the dense granules discharge their contents within the PV. The most important content

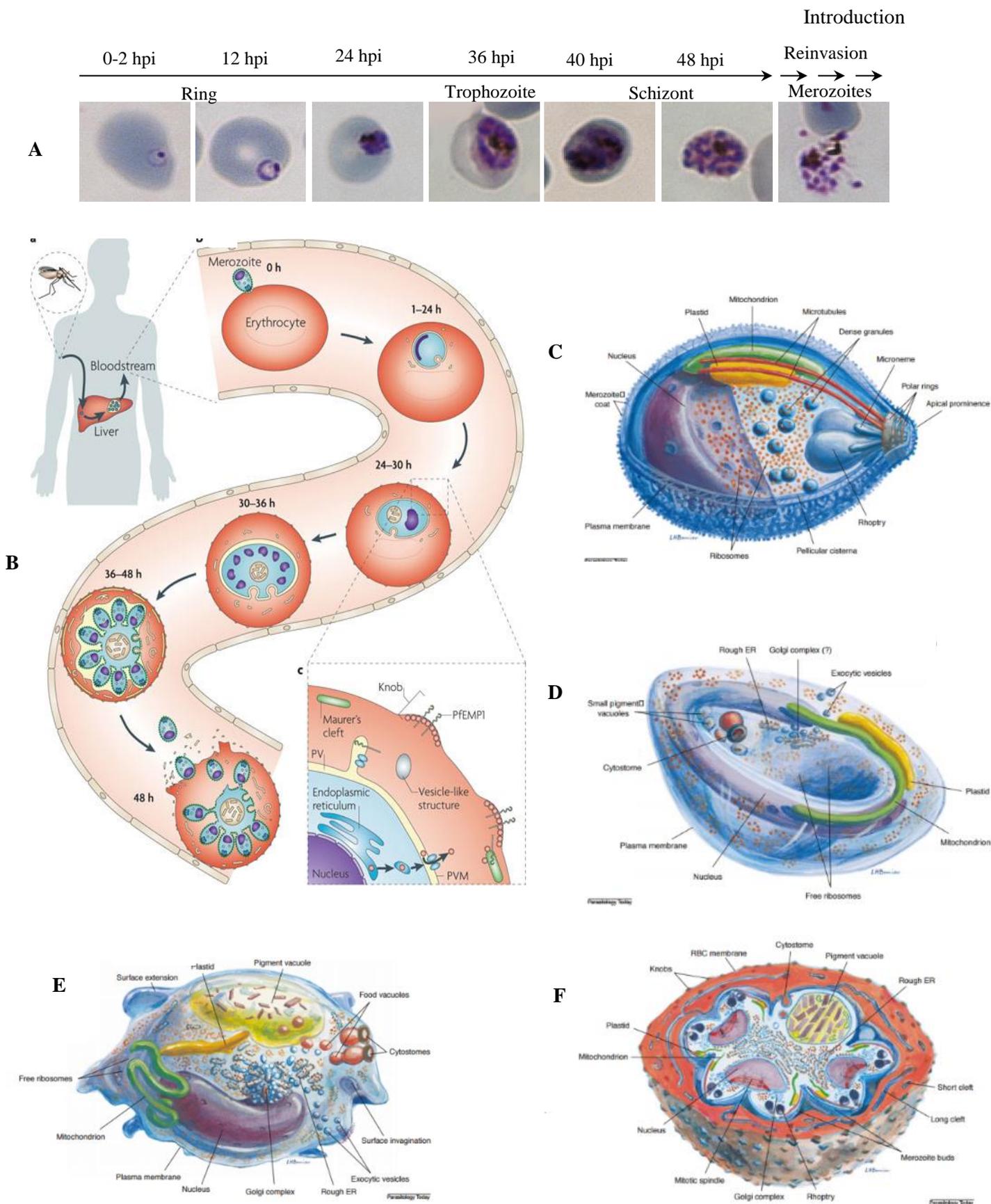
is RESA protein (ring infected erythrocyte surface protein) which is thought to interact with the spectrin of the membrane cytoskeleton causing cell membrane rigidity thus inhibiting further invasion (Cowman et al. 2012, Zuccala and Baum 2011).

### **1.3.1.2. Ring stage**

After invasion, the parasite is reformed into a flat disc shaped ring stage sometimes amoeboid form (Grüning et al. 2011). which has a thick cytoplasm enclose all the organelles while the centre of the disc is thin and contains less structure than the periphery (Fig. 1.3D). Due to this morphology and the position of the nucleus, upon staining with Giemsa, the parasite appears as a ring like structure, so came its name (Aikawa et al. 1967, Langreth et al. 1978). Ring stage last for about 24 hours and is considered a lag phase during which the parasite undergoes host cell modifications that are needed for export and surface expression of the surface antigens that are impinged in immune evasion and consequently the survival of the parasite within the host (Spielmann et al. 2006). The ring stage contains a cytosome through which it starts to feed from the IEs cytosol proteins. Haemoglobin catabolism results in the formation of the haem derivative which forms a brown hemozoin crystals that accumulate within a pigmented vacuole giving the trophozoite stage a unique character (Bannister et al. 2000). Ring stages are known to be found in the peripheral circulation of the host, that's why the first line of laboratory diagnosis is performing thin and thick blood smears from the patient's peripheral blood. Few studies reported the ability of ring stages to adhere to host cells using ring stage surface protein 1/2 (RSP1/2) but the significance and abundance of this adhesive phenotype is not yet well demonstrated (Craig and Scherf 2001, Douki et al. 2003, Pouvelle et al. 2000).

### **1.3.1.3. Trophozoite stage**

Is considered the most metabolically active stage (20-38 hours post invasion), with a great increase in the surface area of the trophozoite. Protein synthesis reaches to the max, this is demonstrated by the enlargement of the endoplasmic reticulum (ER) and the presence of large number of free ribosomes (Bannister et al. 2000). Proteins secreted from the parasites use a complex system of trafficking, these include Kinases, lipases, adhesions and proteases (Goldberg and Cowman 2010).



**Fig. 1.3. *P. falciparum* blood stages.** (A) Giemsa stain of *P. falciparum* asexual blood stages showing the time line of intraerythrocytic life cycle after merozoite invasion (hpi: hours post invasion). (B) Shows 48-hour-long developmental process starting with the ring stage (0–24 hours), then the trophozoite stage (24–36 hours) and the schizont stage (36–48 hours), during which merozoites are formed. (C) (D) (E) and (F) Show ultrastructure of merozoite, ring, trophozoite and schizont stages respectively (Bannister et al. 2000, Goldberg and Cowman 2010).

This stage is also accompanied by the appearance of electron dense knob like structure on the surface of IEs. The knobs thought to be the tangential point between the IEs and host endothelial cells during cytoadhesion (Leech et al. 1984a). These knobs were found to be formed mainly of one protein called KAHRP and a second major protein which is *PfEMP3* (Oh et al. 2000, Pei et al. 2005). Studies evidenced that KAHRP interacts with the IE's cytoskeleton components such as spectrin, actin and spectrin-actin band which in turn changes the physiological characters of the IEs. In addition, some regions of KAHRP can self-assemble and bind to the ATS of *PfEMP1*. Loss of KAHRP will result in knobless IEs which may affect its binding capacity (Pei et al. 2005, Rug et al. 2006, Crabb et al. 1997). Although, a recent study demonstrated that knobless IEs can also adhere (Tilly et al. 2015). The most characteristic event in this stage, is the expression of variant surface antigens (VSAs) on the surface of IEs trophozoite stages. VSAs modify the antigenic character of the IEs allowing the parasite to evade the host immune system and survive within the host. VSAs are *PfEMP1*, RIFIN, STEVOR and *PfMC-2TM* which will be described later in detail.

#### **1.3.1.4. Schizont stage**

It is the parasite stage where the parasite undergoes repetitive nuclear division, mitosis and DNA synthesis (starts during the trophozoite stage). A syncytial schizont is formed of 16-36 nuclei (Arnot and Gull 1998). Upon rupture of the IEs, new haploid merozoites are released in the circulation (Gerald et al. 2011). It was observed that the parasite continues to transport its proteins to the surface of IEs. The surface of the IEs becomes more irregular due to the cytoskeleton distortion caused by the parasite proteins. Atomic force scanning microscopy showed marked increase in the number of knobs on the surface of schizont stage IEs (Bannister et al. 2000).

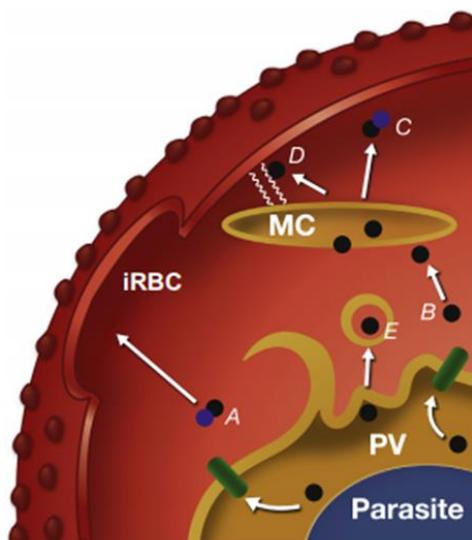
#### ***P. falciparum* protein transporting and trafficking systems**

Human erythrocytes are atypical eukaryotic cells, that they do not contain any organelles necessary for protein synthesis or components for protein trafficking. That's why the parasite has to establish its own mechanisms for protein transport and trafficking (Fig. 1.4.) (Haldar et al. 2006, Bhattacharjee et al. 2008). This begins directly as soon as the merozoite invades the erythrocyte, PV is formed as an

interface between the parasite and the cytoplasm of the host cell. Being inside the PVM, the parasite starts to develop a system to transport the needed nutrients from the erythrocyte cytosol and the extracellular environment owing to the fact that, the parasite needs other sources of amino acids than those of the haemoglobin (Charpian and Przyborski 2008, Lingelbach and Joiner 1998). It is difficult to find a single mechanism that explains the uptake of variety of substances by the parasite. Kirk and colleagues suggested that, the parasite creates new permeability pathways to alter the permeability of the erythrocyte membrane (Kirk and Saliba 2007). Another fact is, the presence of the tubovesicular membranes network (TVN) that extends from the PV to the erythrocyte periphery might help the parasite with nutrients transport (Lauer et al. 1997). However, the origin of the proteins mediating the transport across the erythrocyte membrane is debating. Some postulated that, these proteins are parasite derived owing to the lack of protein biosynthetic pathway in the erythrocytes (Baumeister et al. 2006). On the other hand, Huber and colleagues showed the activation of endogenous erythrocyte  $\text{Cl}^-$  channels by the parasite which plays a rule in the transportation process (Huber et al. 2002).

**The parasite's secretory pathway**, to be able to transport virulence proteins to the surface, the parasites develop a special trafficking system leading to the remodelling of the host erythrocyte surface, increasing its rigidity and forming knobs which in turn help in the cytoadhesion process of the IEs. Once the proteins reach the endoplasmic reticulum they enter a special secretory pathway, where the proteins to be exported are recognised by a signature sequence at the N-terminus known as PEXEL (*Plasmodium* export element) with a pentameric sequence (RxLxE/D/Q) (Marti et al. 2004, Hiller et al. 2004). After the cleavage of PEXEL sequence the proteins are trafficked into the PV then directed to a transporter known as PTEX (*Plasmodium* translocon of exported proteins) (de Koning-Ward et al. 2009). After that the proteins are unfolded then translocated across the PVM to the erythrocyte cytosol (Grüning et al. 2012) then to its final position. The final step is supposed to be mediated through different mechanisms which includes, transport in vesicles, chaperon mediated or in soluble complexes (Kulzer et al. 2010, McMillan et al. 2013). A secretory organelle known as Maurer's clefts (MC) that have a complex structural organisation and functions, was described for continuous secretion of parasites' proteins to the host erythrocyte surface (Tilley et al. 2008, Wickert and

Krohne 2007). Some studies proved the presence of transported PEXEL-independent proteins termed PEXEL negative exported proteins (PNEPs) which are present either permanent (such as SBP1 or REX1) in MC or transient (such as *PfEMP1* or SURFIN) (Dixon et al. 2011, Grüning et al. 2012, Haase et al. 2009 Spielmann et al. 2006). The exact trafficking of proteins to and from the MC is still completely unknown adding to this the presence of interconnections between the TVN and MCs is debating (Tamez et al. 2008, van Ooij and Haldar 2007, Wickert et al. 2004) .

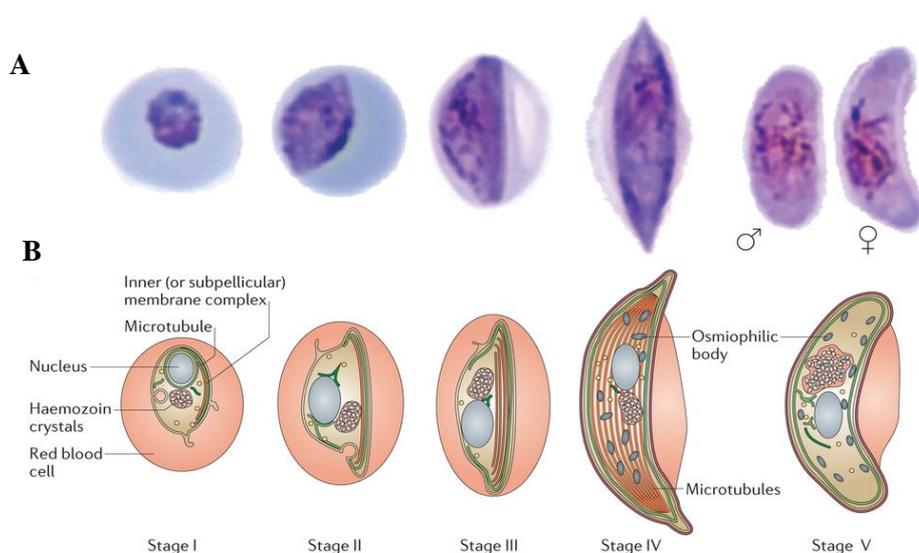


**Fig. 1.4. Schematic presentation of suggested protein export pathways in *P. falciparum* IEs.** The figure represents the parasite enclosed within the parasitophorous vacuole (PV). Black dots represent an exported protein, blue dots represent a chaperone and green cylinders are the PTEX translocon. Different pathways are adopted for protein transport either chaperone mediated (A) or trafficked in soluble complexes to MCs (B) then onto the IE's membrane which could be via a chaperone complex (C) or tethers or actin filaments (represented by the wavy lines) (D). Another suggested way is in the IEs cytosol in vesicles (E) (Proellocks et al. 2016).

### 1.3.2. Sexual stages (gametocytes)

During the asexual intraerythrocytic cycle, subpopulations of schizonts produce merozoites that are committed to differentiate into a single male or female gametocyte this takes about 10-12 days. The maturation time of gametocytes is five times longer than the asexual stages. During maturation gametocytes develop into five morphologically different stages (Fig. 1.5B) (Eksi et al. 2012). Stage I is difficult to be differentiate from the trophozoite stage due to the morphological similarity but with Giemsa staining the hemozoin crystal is smaller than in case of trophozoite. In stage II, the parasite elongates and resemble a D-shape disc. In this stage, the subpellicular microtubules start to be formed. In stage III, the parasite

continues to elongate and its ends become rounded. In stage IV, the gametocyte continues to elongate with a characteristic pointed ends. In stage V, they render their typical crescent shape (Talman et al. 2004). Female gametocytes are more elongated and curved than males (Fig. 1.5A) also using Giemsa stain females stain blue and males stain pink during stage V. The only stage that can be detected in the peripheral blood of patients is the mature stage V whereas other stages remain sequestered until maturation (Josling and Llinas 2015). The mechanism by which the gametocytes sequester is poorly understood. Some studies claimed the abundance of gametocytes in the spleen and bone marrow (Farfour et al. 2012), but the mechanism of their sequestration is still controversial. Sharp and colleagues identified the transcription of *var* genes in the gametocyte stages, which may give a clue for the mechanism of gametocyte sequestration (Sharp et al. 2006). Adding to this, Roger and colleagues postulated that mid and mature sexual stages can bind to HBM cell line (human bone marrow) and this binding is higher to that of sexual stages. They supposed that, the receptor which mediate this binding is ICAM-1 (Rogers et al. 1996). On the other hand, a recent study tested the binding capacity of 3D7 isolate gametocyte to different endothelial cell lines (HUVEC and HDMEC) and to HBM cell lines (HBMEC-60 and HBMEC33), to which the gametocytes showed minimal binding capacity compared to that of trophozoites, they concluded that gametocyte might use another mechanism to sequester other than that used by trophozoites (Silvestrini et al. 2012).



**Fig 1.5. Development and maturation of sexual stages (gametocytes) in *P. falciparum* IEs. (A) Giemsa stain of the five different gametocyte stages (B) Schematic presentation of the morphological changes that occur during the five gametocyte stages (Josling and Llinas 2015) .**

## **1.4. Pathogenesis of *P. falciparum* CM**

Several processes are impinged in the pathogenesis of CM. These mechanisms were recently reviewed by Cunnington and colleagues where they summarized the three main mechanisms postulated in the literature to be involved in CM pathogenesis. These three mechanisms are as follows, the sequestration of IEs, vascular endothelial dysfunction and finally the inflammation hypotheses. It is believed that a combination of these three mechanisms predisposes for bad prognosis and serious complications (Fig. 1.6) (Cunnington et al. 2013).

### **1.4.1. Sequestration of *P. falciparum* IEs**

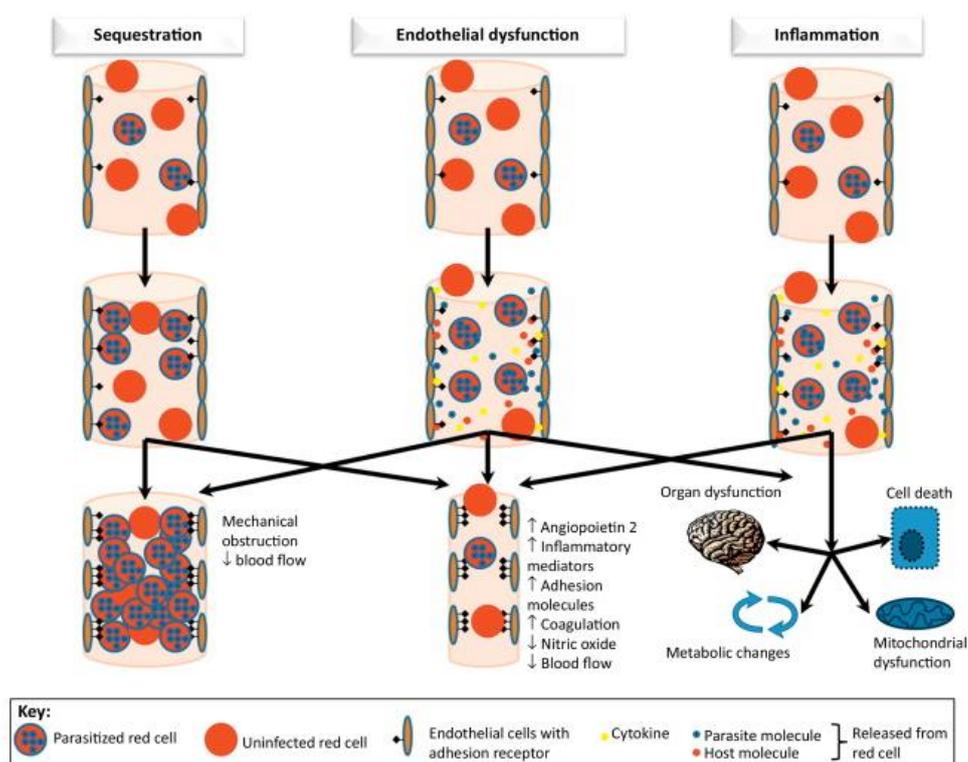
Three IEs adhesion phenotypes were described (Fig 1.7); cytoadhesion to endothelial cells, rosetting with non-infected erythrocytes and finally platelet-mediated clumping (Rowe et al. 2009). The cytoadhesion of IEs to endothelial cells in post capillary bed causes obstruction in blood flow which decreases both tissue perfusion and removal of waste product resulting in lactic acidosis and finally organ dysfunction (Idro et al. 2010). Cytoadherence occurs due to the binding of IEs to the vascular endothelium in a parasite host interaction mechanism. Different endothelial receptors were identified to be used by the IEs in this interaction such as CD36, ICAM-1, P-selectin, EPCR and others (discussed later). Parasite derived variant surface antigens (VSAs) are able to mediate these interactions. *PfEMP1* is the main player which thought to be responsible of most of the binding phenotypes caused by IEs others are RIFIN and STEVOR variants but their exact function in mediating cytoadhesion is still under investigations (Rowe et al. 2009).

### **1.4.2. Vascular endothelial dysfunction**

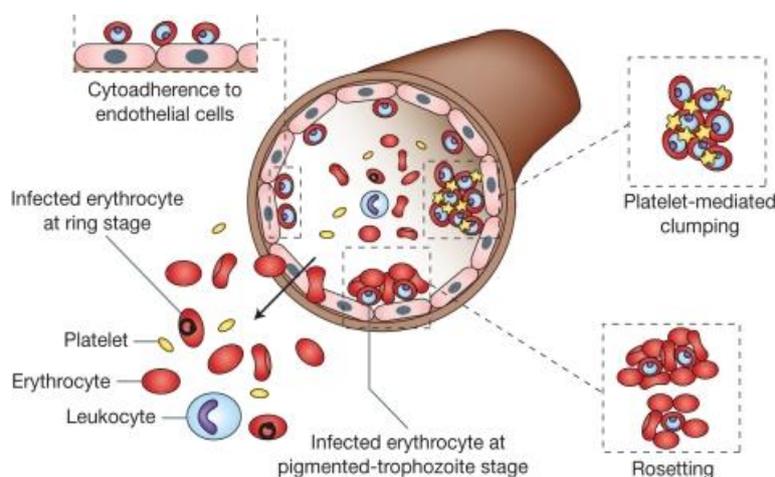
Cytoadhesion of IEs on the vascular endothelium results in endothelial activation due to the transcription of the genes involved in inflammation. This in turn leads to the release of endothelial microparticles (EMPs) and apoptosis of the host cell. Platelet mediated clumping -on the other hand- causes injury of the endothelium through a cytotoxic effect. In addition, the repair mechanism of the endothelium is impaired due to marked decrease in the circulating endothelial progenitor cells (Storm and Craig 2014).

### 1.4.3. Inflammation and inflammatory products

Several studies demonstrated an increased level of inflammatory mediators in SM compared to UM. This mechanism may explain the SM in case of *P. vivax* where it stimulates the production of more inflammatory products than in case of *P. falciparum* infection. TNF (tumor necrosis factor) is the most frequent cytokine released in CM. TNF causes upregulation of ICAM-1 in cerebral endothelium which in turn increases the cytoadhesion of IEs (Hunt and Grau 2003). A signalling molecule which might play a role in the pathogenesis, is NO (nitric oxide). Its overproduction in case of inflammation may be a cause for development of coma in CM due to the fact that it can cross the blood brain barrier, enter the brain tissue and alter the neurotransmission (Clark et al. 1992, Sharma et al. 2007).



**Fig. 1.6. The pathogenesis of severe malaria.** Sequestration of IEs on the vascular endothelium results in mechanical obstruction of the blood flow and activation of the vascular endothelium. Host cell releases cytokines and other inflammatory products, which result in endothelial activation and dysfunction, and therefore impaired blood flow. Inflammatory mediators are among the molecules, which can cause endothelial activation, but may also contribute directly to organ dysfunction, cell death, mitochondrial dysfunction, and metabolic derangements (Cunnington et al. 2013).



**Fig. 1.7. Different adhesion phenotype of *P. falciparum* IEs.** Three main binding phenotypes were described for *P. falciparum* IEs. First, cytoadhesion to endothelial cells mediated by interaction with a host cell receptor. Second, rosetting which is the binding of IEs to non-infected erythrocytes. Third, platelet mediated clumping (Rowe et al. 2009).

### 1.5. Antigenic variation and multigene families

In order to set up a chronic infection and survive within the host, meanwhile avoiding the antibody response of the immune system, some infectious organisms have the ability to change some of their surface antigens exposed to the immune system of their host. This process is called antigenic variation. This phenomenon exists in many pathogens including bacteria, fungi and protozoan parasites. These microorganisms use different methods for antigenic variation. The protozoan parasites control the antigenic variation system using multi-copy gene families where set of genes having the same function but every gene encodes for a different surface antigen (Dzikowski and Deitsch 2006). The best known example in protozoan parasites that can perform antigenic variation, is *Trypanosoma brucei*, harbouring VSG (variant surface glycoprotein) gene family which consists of about 1000 members (Deitsch et al. 2009). Similar examples but smaller in number exist in Plasmodium, *Giardia* and *Babesia* species (Deitsch et al. 2009, Dzikowski and Deitsch 2006). *Plasmodium* species have a similar but smaller family called variant surface antigens (VSAs). *Plasmodium* interspersed repeats, is the largest protein family presents in Plasmodium species (MacDonald et al. 2001). This family includes, *rif*, *vir*, *kir*, *cir*, *bir* and *yil* families in *P. falciparum*, *P. vivax*, *P. knowlesi*, *P. chabaudi*, *P. berghei* and *P. yoelli*, respectively (Singh et al. 2014).

In *P. falciparum*, four large multi-copy gene families encode the VSAs. These are the *var* gene family which consist of about 60 copies per haploid genome and encodes for PfEMP1s variants that play the main role in sequestration of IEs (will be discussed later in details). The other three families are, *rif* (repetitive interspersed family) about 185 copies, *stevor* (subtelomeric variable open read frame) about 66 copies and *Pfmc-2tm* (*P. falciparum* Maurer's clefts 2 transmembrane) about 13 copies, their gene products are RIFIN, STEVOR and PfMC-2TM proteins, respectively. The *rif* and *stevor* gene families attracted the researchers' attention in the last few years due to the possibility of their involvement in the pathogenesis of severe diseases. Both families are located in the subtelomeric regions of all 14 *P. falciparum* chromosomes and are clustered together with the *var* genes where they form repeats of several genes either on the same or the opposite strand (Gardner et al. 2002). Both *rif* and *stevor* have the same structure, where they are composed of two exons. The first predicted to code for the signal peptide and the second for the main part of the protein with semi conserved and highly variable parts (Bachmann et al. 2015, Gardner et al. 2002, Petter and Duffy 2015). As both proteins contain a signal peptide as well as PEXEL/HT motif, which labells them for export across the parasitophorous vacuole into the host cell (Hiller et al. 2004, Marti et al. 2004).

*rif* genes are small genes (about 1000 base pairs) they are divided into two groups (A and B) according to the existence of 25 amino acids in the semi-conserved domain (Gardner et al. 2002, Joannin et al. 2008, Petter et al. 2007). *rif* genes were predicted to have two transmembrane domains on both sides of the variable region (Cheng et al. 1998, Kyes et al. 1999, Sam-Yellowe et al. 2004); but a recent study predicted only one transmembrane domain for group-A *rif* genes (Bultrini et al. 2009). During the erythrocyte schizogony, *rif* genes are transcribed 12-27 hours post invasion (Bachmann et al. 2012), whereas during the mature stage V gametocytes, group-A *rif* was found to be transcribed (Petter et al. 2008). In addition, Wang and colleagues found high expression of *rif* genes in sporozoite stage (Wang et al. 2010). The function of RIFIN variants is until now not well described but several studies documented anti-RIFIN antibodies immune response with a stable response and rapid clearance of the parasite from the circulation (Schreiber et al. 2008).

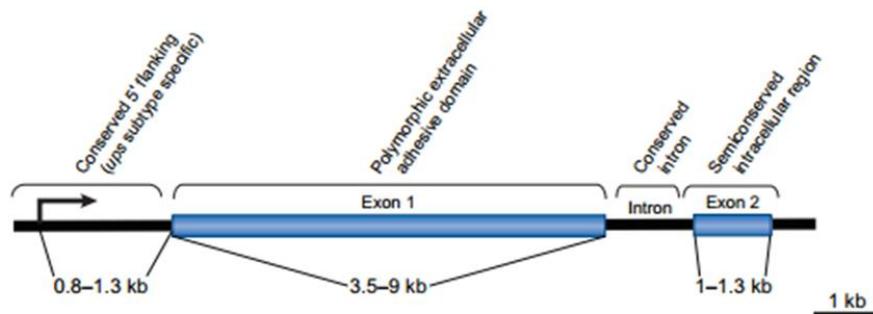
*stevor* genes are similar in structure to *rif* genes with debating old description of two transmembrane domain regions. Two studies recently described the presence of only one transmembrane domain (Bachmann et al. 2015, Niang et al. 2009). *stevor* were found to be expressed during different stages of the parasite development. During the erythrocyte schizogony, its transcription was found mainly 22-32 hours post invasion. Bachmann and colleagues reported also an early peak of transcription level in early ring stages in *ex-vivo* clinical isolates (Bachmann et al. 2012). Expression of *stevor* genes was also reported in merozoites, gametocytes and sporozoites suggesting a probable important functions of this gene family (Khattab et al. 2008, Khattab and Meri 2011, McRobert et al. 2004). Niang and colleagues found that anti-STEVEOR antibodies could inhibit the merozoite invasion mechanism. They also proved the role of STEVEOR in rosetting independently from *PfEMP1* through the interaction with glycoprotein C on the surface of erythrocytes (Niang et al. 2014).

Although the function of both RIFIN and STEVEOR variants in cytoadhesion is still controversial and under investigation a very strong evidence was found in a case study of a splenectomised patient with a malaria relapse it was found that the transcription of *var*, *rif* A and *stevor* was lost. This may give a clue of their involvement in the immune evasion mechanism (Bachmann et al. 2009).

## 1.6. *var* genes family

Decades of research on the *var* gene family were not enough to reveal all the secrets of the complexity of their genomic organisation. The history of their discovery started when Cox who investigated mice infected with *P. berghei* and concluded the possibility of presence of different parasite populations that survive safely within the immune host (Cox 1959). After that, Brown and Brown reported the antigenic variation between *P. knowlesi* IEs populations (Brown and Brown 1965). Few years later, the *PfEMP1* antigens were identified on the surface of trophozoite and schizont IEs (Leech et al. 1984b). Then it was reported that the genes encoding the *PfEMP1* -which are the *var* genes- are found only in *P. falciparum* and *P. reichenowi* (Jeffares et al. 2007). *var* genes family consists of approximately 60 copies per parasite genome (Kraemer and Smith 2006, Kyes et al. 2001, Su et al. 1995) each gene comprises two exons (Fig 1.8). Exon 1 encodes both the highly

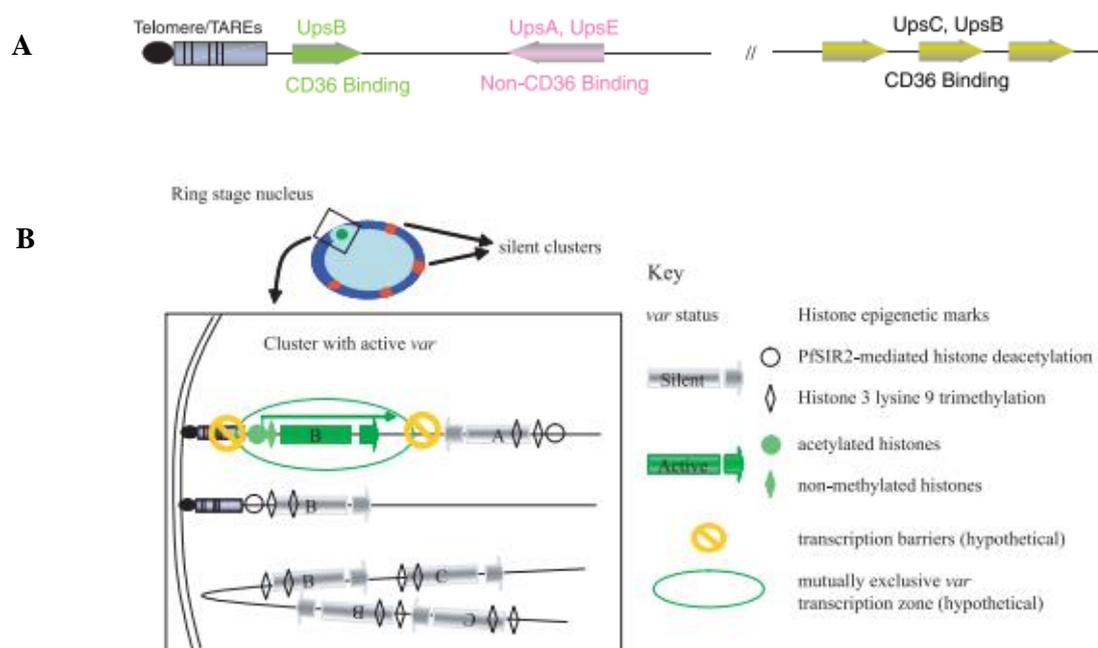
diverse extracellular region of the *Pf*EMP1 and transmembrane region. Exon 2 encodes the highly conserved ATS (acidic terminal segment) domain which is extended in the erythrocyte cytoplasm (Kyes et al. 2007, Scherf et al. 2008).



**Fig. 1.8. Characteristic features of *var* genes.** Exon 1 encodes both the highly diverse extracellular region of the *Pf*EMP1 and transmembrane region. Exon 2 encodes the highly conserved ATS domain (Scherf et al. 2008).

The position of the *var* genes in the 14 chromosomes is mainly subtelomeric. Whereas, one third of the *var* genes are located centrally in the chromosome. Each chromosome harbours from one to three *var* genes followed by *rif* or *stevor* genes. *var* genes are classified into four Ups (upstream sequence groups; A, B, C and E). This classification is based on the sequence similarity and 5' promoter regions. Ups C *var* genes are central in the chromosome while Ups B *var* genes are either central or subtelomeric. Both Ups C and Ups B are transcribed away from the telomere. On the other hand, Ups A and Ups E are subtelomeric and are transcribed towards the telomere (Fig 1.9A). (Kraemer et al. 2007, Kyes et al. 2007, Ralph and Scherf 2005). *P. falciparum* at a given time transcribes single gene of the entire *var* family, whereas all the others are silenced, in a process known as mutually exclusive expression (Chen et al. 1998, Horrocks et al. 2004, Kyes et al. 2003, Scherf et al. 1998). *var* gene switching results in the expression of only one *Pf*EMP1 on the surface of IEs, thus the parasite avoids recognition by the antibody response and can establish chronic infection within the host (Guizetti and Scherf 2013, Kraemer and Smith 2006). *var* gene expression is thought to be controlled by four main general steps; silencing, activation, poising and switching (Guizetti and Scherf 2013). Normally *var* genes are silenced, only a single *var* gene is activated in the early ring stage and *var* gene transcription is stopped in the late trophozoite stage (Scherf et al. 1998, Schieck et al. 2007). The poised *var* gene is epigenetically marked to be reactivated during the next cycle. The last step is that some cells perform *var* gene

silencing to switch to another member of the family (Scherf et al. 2008). Some facts about how one *var* gene is exclusively expressed can be summarized in three observations from previous studies (Fig 1.9B) (Kyes et al. 2007). 1. Nuclear repositioning, all *var* genes are located at the nuclear periphery most probably in telomeric clusters. Ralph and colleagues observed that the active *var* gene detaches from the silent cluster to a new position within the nuclear periphery (Ralph et al. 2005). 2. Histone modifications, it is well known that histone modifications can influence the gene expression. Two markers were observed associated with silent *var* genes. *PfSIR2* which causes histone deacetylation in the silent *var* gene. The second epigenetic marker reported by Chookajorn and colleagues, is the modified histone H3K9me3, where they observed that *var* gene silencing is associated with methylation of histone H3 at lysine K9 (Chookajorn et al. 2007). 3. Interaction between *var* 5'promotor and *var* intron promotor (Kyes et al. 2007). Frank and colleagues in their experiments showed, that loss of the intron results in *var* promoter activation. Also they proved that, each intron could only silence a single promoter, suggesting a one to one pairing requirement for silencing (Frank et al. 2006).



**Fig. 1.9. (A) Classification of *var* genes according to upstream promoter type.** Arrows indicate the direction of transcription (Kyes et al. 2007). **(B) Schematic presentation of the common observed evidence for *var* gene silencing and activation** (nuclear repositioning and chromatin modifications) (Kyes et al. 2007).

## 1.7. *PfEMP1*

*PfEMP1* is responsible for determination of the IEs adhesive properties (Baruch et al 1996). It is a large protein ranging in size between 200 and 350 kDa. The *PfEMP1* consists of N-terminal segment (NTS), Duffy binding like domains (DBL), cysteine rich interdomain regions, one transmembrane domain and the acidic terminal segment (ATS). There are seven major classes of DBL domains ( $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ ,  $\epsilon$ ,  $\zeta$  and  $\chi$ ). Whereas, CIDR domains were divided into three main classes ( $\alpha$ ,  $\beta$ ,  $\gamma$ ) (Rask et al. 2010). Most of *PfEMP1* (except Type 3 and VAR2CSA) have a head structure at their N-terminus composed of semi-conserved DBL $\alpha$  domain and a CIDR domain (Gardner et al., 2002). The head structure is followed by a second, more diverse, DBL-CIDR pairs and thus totally composed of 7-10 extracellular domains. Although there is a great similarity in the general architecture of the *PfEMP1* variants, there is a great sequence diversity among different parasite isolates when comparing their amino acid sequence. This high level of sequence diversity is believed to be created through mitotic recombination that generates new combinations of the proteins' structural domains. As a result, there are unlimited possibilities for antigenic variation within the *PfEMP1* repertoire (Pasternak and Dzikowski 2009). According to these facts and in order to facilitate *PfEMP1* comparisons, Rask and colleagues performed an alignment of domain subtype architecture of 399 known *PfEMP1*s which resulted in the identification of 23 conserved domain composition called domain cassettes (DC). They defined the DC as two or more consecutive domains present in three or more of the 7 genomes used in their study (Fig. 1.10) (Rask et al. 2010). The most conserved domains were found to be, DC1 which represents var1 *PfEMP1* variant and it is characterised by only one CIDR $\alpha$  domain in its architecture. The second is DC2 which represents the unique var2csa with its special *PfEMP1* domains that are able to bind to CSA (which is abundant within the placental intervillous space) and induce pregnancy associated malaria (PAM). Finally, DC3 which represents very short variants with only two DBL domains (Rask et al. 2010). In the previous studies it was observed that individual domains as well as domain sub classes show specific ligand binding interaction with specific endothelial cell receptors. The problem is that the variable sequence polymorphism in each subclass made it difficult to set a common rule to identify the binding specificity. However, some predictions based on several studies

are now clear. For instance, the *PfEMP1* head structure is nearly always comprised of a DBL-CIDR, followed by other DBL and CIDR tandems (Fig 1.8) (Smith et al. 2013). The *PfEMP1* head structure plays a key role in binding; most group B and C *PfEMP1* proteins appear bind to CD36 (with CIDR $\alpha$ 2-6 domain) (Baruch et al. 1996, Robinson et al. 2003, Smith et al 2013), whereas many group A variants bind to EPCR (with CIDR $\alpha$ 1 domain) (Turner et al. 2013). Single domain interactions were also identified for example DBL $\beta$ 5 domain binds to ICAM-1 (Bengtsson et al. 2013, Howell et al. 2008). In addition, DC4 was recently identified to bind to ICAM-1 (Bengtsson et al. 2013, Madkhali et al. 2014). The paradox of *P. falciparum* IEs ability to bind to variable endothelial receptors is still a big unsolved puzzle in the field of malaria research. Until now about 22 endothelial receptors were identified to interact with *P. falciparum* IEs. Only four receptors (CD36, ICAM-1, EPCR and CSA) were studied in detail, the other receptors are still under investigations (Baruch et al. 1996, Robinson et al. 2003, Smith et al 2013, Esser et al. 2014, Rowe et al. 2009, Turner et al. 2013, Fried and Duffy 2002).

Cassette #	Alias	UPS	PfEMP1 domain cassette structure										
			Component 1	Component 2	Component 3	Component 4							
2	VAR2CSA	E	DBLpam1	DBLpam2	CIDRpam	DBLpam3	DBLpam4	DBLpam5	DBL $\epsilon$ 10				
3	VAR3	A								DBL $\alpha$ 1.3	DBL $\epsilon$ 8		
1	VAR1	A2	DBL $\alpha$ 1.1/4	CIDR $\alpha$ 1.2/3	DBL $\beta$ 1.1/11	DBL $\gamma$ 1/15	DBL $\epsilon$ 1			DBL $\gamma$ 8	DBL $\zeta$ 1/2	DBL $\epsilon$ 5	
5		A					DBL $\gamma$ 12	DBL $\delta$ 5	CIDR $\beta$ 3/4	DBL $\beta$ 7/9			
16		A	DBL $\alpha$ 1.5/6	CIDR $\delta$									
13		A	DBL $\alpha$ 1.7	CIDR $\alpha$ 1.4									
15		A	DBL $\alpha$ 1.2	CIDR $\alpha$ 1.5									
11		A	DBL $\alpha$ 1.8	CIDR $\beta$ 2	DBL $\gamma$ 7						DBL $\epsilon$ 11	DBL $\zeta$ 2/3	DBL $\epsilon$ 6
6		B(A,C)									DBL $\gamma$ 14	DBL $\zeta$ 5	DBL $\epsilon$ 4
7		B(C)									DBL $\epsilon$ 2	DBL $\epsilon$ 7	DBL $\epsilon$ 3
9		B1									DBL $\gamma$ 3	DBL $\zeta$ 4	
10		B(A,C)									DBL $\zeta$ 6	DBL $\epsilon$ 9	
12		B(A)									DBL $\zeta$ 3	DBL $\epsilon$ 12	
8		B2	DBL $\alpha$ 2	CIDR $\alpha$ 1.1	DBL $\beta$ 12	DBL $\gamma$ 4/6							
14		B	DBL $\alpha$ 0.6	CIDR $\alpha$ 3.1	DBL $\beta$ 5								
17				CIDR $\alpha$ 5	DBL $\beta$ 5								
22		B,C	DBL $\alpha$ 0.4/18	CIDR $\alpha$ 6	DBL $\beta$ 5								
21		C(B)	DBL $\alpha$ 0.18/21	CIDR $\alpha$ 2.1	DBL $\beta$ 2								
18		B1	DBL $\alpha$ 0.14	CIDR $\alpha$ 4									
19		B1(C1)	DBL $\alpha$ 0.16	CIDR $\alpha$ 3.4									
20		B1(C1)	DBL $\alpha$ 0.9	CIDR $\alpha$ 2.7									

**Fig. 1.10. Schematic structure for the proposed domain cassettes.** Alignment of domain subtype architecture of 399 known *PfEMP1*s in seven parasite genomes. The comparison results in the identification of 23 conserved domain composition called domain cassettes (DC). DC are two or more consecutive domains present in three or more of the 7 genomes analysed (Rask et al. 2010).

## **1.8. Receptors of interest in this study**

### **1.8.1. ICAM-1**

Intra cellular adhesion molecule 1 (ICAM-1) is a transmembrane glycoprotein with five extracellular immunoglobulin-like domains (D1-D5). ICAM-1 induces cell to cell adhesion allowing intercellular communication, T cell-mediated defense mechanism, and inflammatory response (Egal et al. 2014). ICAM-1 mediates leucocyte adhesion and migration to sites of inflammation by binding to LFA-1 and Mac-1 (Lennartz et al. 2015). It is expressed at minimal levels on endothelial cells and during malaria-induced inflammation it is highly upregulated (Turner et al. 1994). There is a big debate about the role of ICAM-1 in CM. A co-localisation of IEs with ICAM-1 in patients who died of CM was reported (Turner et al. 1994). Silamut and colleageus observed that vessels with higher ICAM-1 levels have higher levels of sequestration (Silamut et al. 1999). Ochola and colleagues recently showed a correlation between ICAM-1 binding and cerebral malaria (Ochola et al. 2011). Another study showed higher binding capacity to ICAM-1 in patients isolates with clinical malaria compared with asymptomatic malaria (Newbold et al. 1997).

### **1.8.2. Selectins**

Selectins are family of cell adhesion molecules which shares a conserved domain structure. They contain a N-terminal lectin (Lec) domain, an epidermal growth factor (EGF)-like domain, variable numbers of short consensus repeat (SCR) domains, a single-span transmembrane region, and a C-terminal cytoplasmic domain (Erbe et al. 1992). Three selectin members are known P-selectin, E-selectin and L-selectin. During the leukocyte aggregation cascade, selectins mediate leukocyte rolling, which reduces the velocity of leukocyte movement on the endothelial cells to allow firm adhesion (Granger and Kubes 1994). P-selectin is expressed by both endothelial cells and platelets, while E-selectin is only expressed by endothelial cells (Ley et al. 2007). P-selectin is normally stored in endothelial cells granules known as Weibel–Palade bodies, from which it can be rapidly mobilized to the cell surface by histamine and leukotrienes (Bevilacqua and Nelson 1993, Mayadas et al. 1993).

Both P- and E-selectin were found to be highly expressed in the lung, small intestine, and heart after endotoxin challenge, while the brain and skeletal muscle exhibit the smallest responses (Panes and Granger 1998).

### **1.8.3. Tetraspanins**

Tetraspanins consists 33 protein families with a molecular mass of 20-50 kDa. They have four transmembrane domains, and two extracellular loops (one small and one large). Tetraspanins recently were found to play a role in different infectious diseases (van Spriel and Figdor 2010). In 1991, Power and colleagues described CD9 as a motility-related factor where they found that anti CD9 mAbs inhibits the migration of multiple cancerous cell lines (Powner et al. 2011). CD9 and CD81 tetraspanins thought to have the same structure, as they have a similar length, same number of cysteine residues and both lack post-translational modifications (Hulme et al. 2014). Interestingly, CD81 is known to be used by *P. falciparum* parasite to enter the hepatocyte (Chambrion and Le Naour 2010). CD151 tetraspanin was found to be expressed on the surface of endothelial cells and plays a role in the regulation of pathologic angiogenesis (Zhang et al. 2011). It exists in a variety of epithelial and mesenchymal cells. CD151 was found to be impinged in different biological processes and identified as a global regulator of endothelial cell-cell and cell-matrix adhesions (Sterk et al. 2000). Deletion of CD151 gene results in defects in the epithelial integrity of both kidney and the skin, adding to this impaired wound healing process and platelet aggregation (Zevian et al. 2011). Both CD9 and CD151 were first described as receptors for *P. falciparum* by Esser and colleagues where they tested the binding capacity of IEs to these receptors using a pool of patient isolates and the laboratory isolate IT4 (Esser et al. 2014).

## 2. Aim of this study

*P. falciparum* IEs sequestration within the capillary beds of vital organs leads to life threatening complications especially in children under five years of age. The mechanism of sequestration is accomplished due to an interaction between a host receptor and a parasite ligand. According to previous studies *P. falciparum* IEs are able to bind to numerous endothelial receptors. Until now, 22 receptors are known to be implicated. Investigating the molecular mechanisms behind the sequestration of IEs will be the first step to develop anti-adhesive therapies which may act as an adjuvant therapy in treatment of patients with severe malaria.

The aim of this study was to:

1. Identify the binding capacity of the two *P. falciparum* laboratory isolates (IT4 and 3D7) to the receptors of interests (ICAM-1, P-selectin, E-selectin, CD9 and CD151).
2. Select and enrich *P. falciparum* IEs that are able to bind to the receptors of interest.
3. Analyse the transcriptome profiles of:
  - Starting *in vitro* culture (which resembles the whole populations).
  - IEs population that can bind to wild-type CHO cells
  - Four different IEs populations selected for binding to the human endothelial receptors (ICAM-1, P-selectin, E-selectin, CD9 and CD151).
4. Identify the *P. falciparum* ligand mediating binding to the receptor of interest.

### 3. Materials and methods

#### 3.1. Materials

##### 3.1.1. Technical and mechanical devices

Type	Specification	Distributor
Bioanalyzer System	Agilent 2100	Agilent Technologies
Centrifuge	5415 D	Eppendorf
Centrifuge	5427 R	Eppendorf
Centrifuge	5810 R	Eppendorf
Counting chamber		Marienfeld
FACs sorter	BD FACSAria™	BD Biosciences
Fine scale		Sartorius
Flow cytometer	Accuri C6	BD Biosciences
Fluorescent microscope	EVOS FL	ThermoFisher
Freezing container	Nalgene Cryo 1°C	Nalgene
Incubator with CO <sub>2</sub>	Function Line	Heraeus Instruments
pH-Meter	CG 840	Schott
Pipettes	2 /20/ 200/1000µL	Gilson
Qubit®Fluorometer		ThermoFisher Scientific
Sequencing platform	HiSeq™ 4000	Illumina
Sequencing platform	MiSeq	Illumina
Spectrophotometer	NanoDrop 2000	Thermo Scientific
Sterile bench	Class II BSC	ESCO Labculture®
Thermocycler (PCR system)	ABI 9700 GeneAmp	PE Applied Biosystems
Thermomixer	5355R	Eppendorf
Transmission electron microscope	Tecnai Spirit	FEI
Transmitted inverted light microscope	EVOS XL	ThermoFisher
Transmitted light microscope	Axiostar plus	Zeiss
Transmitted light microscope	cx31	Olympus
Vortex mixer	VF2	JANKE & KUNKEL
Water bath	SW 20	Julabo

##### 3.1.2. Softwares

Software	Distributor
Agilent 2100 Expert Software	Agilent Technologies
DEseq version 1.24	Bioconductor
FACSDiva™ (version 6.1.3)	BD Biosciences
FastQC version 0.11.5	By S. Andrews
Illumina Experiment Manager	Illumina
ImageJ version 1.48V	National institute of health USA (Public domain)
MiSeq Control Software	Illumina
MiSeq Reporter (MSR)	Illumina
Prism® (version 6)	GraphPad

### 3.1.3. Labware and disposables

Labware	Manufacturer/ Distributer
24-well-tissue culture plate (flat bottom with lid) (83.3922)	Sarstedt
Glass coverslips	Engelbrecht
Hydrophobic Filters, 0.45µm, Sterile	Thermo Scientific
Petri dish 92x16mm with cams	Sarstedt
Safe-Lock Tube PCR clean 1.5ml	Eppendorf
Tissue Culture Coverslips 13 mm (Plastic)	Sarstedt
Tissue culture flask 250 ml (T75) (658175)	Greiner Bio-One
Tissue culture flask 50 ml (T25) (690175)	Greiner Bio-One
Sterile FACs tubes (PP) (5ml)	Falcon
Millipore Stericup™ Sterile Vacuum Filter	EMD
CellTrics® 30 µm (sterile pyrogen free cell filter)	Partec
Gas mixture CRYSTAL-mix (1% O <sub>2</sub> , 5% CO <sub>2</sub> , 94% N <sub>2</sub> )	Air liquid
Falcon™ 15mL Conical Centrifuge Tubes	Falcon
Falcon™ 50mL Conical Centrifuge Tubes	Falcon

### 3.1.4. Chemical and biological reagents

Reagents	Manufacturer/ Distributer
Accutase (100ml) (L11-007)	PAA
Biocoll separating solution	Biochrom
Dako Fluorescence Mounting Medium	Dako
DMSO - Dimethylsulfoxid 250ml	Roth
D-Sorbitol	Sigma
Ethanol	Merck
FailSafe Enzyme Mix only (100 units)	Biozym
G 418-BC (Neomycin) (50 ml) (658175)	Greiner Bio-One
Gelatin solution (100 ml)	G1393---016M6030
Gentamicin 40mg/ml	ratiopharm
Giemsa solution (500ml)	Merck
Glutaraldehyde (25%) (100 ml)	Merck
Glycerol	Merck
Ham's F12	PAN Biotech
Human Blood (0 <sup>+</sup> )	Universitätsklinikum Hamburg-Eppendorf
Human Serum (A <sup>+</sup> )	Interstate Blood Bank, Inc (Memphis, TN, USA)
Hypoxanthine (9636-5G)	Sigma
Immersion oil 518 N	Zeiss
Isopropanol	Merck
Leica CV Mount	Leica Biosystems
Lipofectamine 2000 reagent	Life Technologies
Methanol	Sigma
Penicillin-Streptomycin (10.000 U/ml)	Life Technologies
Percoll (17-0891-02)	GE Healthcare

<b>Reagents</b>	<b>Manufacturer/ Distributer</b>
RNase AWAY™ Spray Bottle	Roth
RPMI-1640 + L-Glutamin; + 25 mM HEPES; - NaHCO <sub>3</sub>	AppliChem
TRIzol reagent®	Life Technologies

### 3.1.5. Kits and standards

<b>Kit</b>	<b>Manufacturer/ Distributer</b>
AGENCOURT® RNAClean up kit	Beckman Coulter
Agilent RNA 6000 Pico Kit	Agilent Technologies
Agilent High Sensitivity DNA Kit	Agilent Technologies
AMPure XP Purification	Beckman Coulter
MinElute PCR Purification Kit	Qiagen
MiSeq Reagent Kit V3 (150 cycles)	Illumina
Mycoplasma Removal Agent (5 ml)	VWR
PureLink RNA Mini Kit	Ambion
Qubit® dsDNA HS Assay Kit	Thermofisher Scientific
Ribo-Zero™ Magnetic Gold Kit (Human /Mouse/Rat)	Epicentre
RNA-Seq Barcode Primers (Set1) (Set3)	Illumina
RNeasy MinElute Cleanup Kit (Cat. No. 74204, Qiagen)	Qiagen
ScriptSeq™ v2 RNA-Seq Library Preparation Kit	Illumina
TURBO DNAase Kit	Life Technologies

### 3.1.6. Antibodies

<b>Primary Antibody</b>	<b>Manufacturer/ Distributer</b>
Mouse anti human CD151 (LIA 1/1) (monoclonal IgG1) 5µg/ml (1:33)	Homemade (AG Horstmann)
Mouse anti human E-selectin /CD62E (monoclonal IgG1) 5µg/ml (1:33)	R&D Systems
Mouse anti human P-selectin /CD62P (monoclonal IgG1) (1:33)	Santa Cruz Biotechnology
Rat- Anti-mouse CD44 (monoclonal KM114) 5µg/ml (1:33)	Antikoerper-online.de
<b>Secondary Antibody</b>	
Alexa Fluor 594 αMaus IgG (H+L) 2 mg/ml (1:400)	Ziege Invitrogen
<b>Labelled Antibody</b>	
Mouse anti-human CD151-APC labelled	Biolegend
Rat Anti mouse/human CD44 (monoclonal IM7) PE/Cy labelled 0.2 mg/ml	Biolegend
Mouse anti-human E-selectin/CD62E-APC labelled	Biolegend

### 3.1.7. Stock solutions, buffers and mediums

#### 3.1.7.1. *P. falciparum* culture

##### **RPMI- complete culture medium (1L)**

RPMI 1640 (dissolve in 700 ml dist. water)	16.4g
Hypoxanthine (dissolve in 50 ml dist. water) (boil for 3min.)	0.05g
Human serum (A+) (Inactivated 2x 56°C for 30 min)	100ml
NaHCO <sub>3</sub> (7,5%)	30 ml
Gentamicin (40 mg/ml)	250 µl
Fill up with dist. water – sterile filtrate- keep at 4°C for 2 maximum weeks	

##### **Synchronization solution (5 % D-Sorbitol)**

D-Sorbitol	25.0 g
Dist. water	500 ml
Sterile filtrate- keep at 4°C	

##### **Freezing solution (MFS)**

D-Sorbitol	3 g
NaCl	0.65 g
(dissolve in 72 ml dist. water)	
Glycerol (autoclavated)	28 ml
Sterile filtrate- keep at 4°C	

##### **Thawing solution (MTS)**

NaCl	35g
(dissolve in 1 L dist. water) sterile filtrate- keep at 4°C	

##### **Blood (sterile, human erythrocyte concentrate, blood group 0<sup>+</sup>)**

##### **10 % Giemsa stain**

Giemsa solution	10 ml
Water	90 ml

##### **Weiser-buffer (2 L)**

Na <sub>2</sub> HPO <sub>4</sub>	2.18 g
KH <sub>2</sub> PO <sub>4</sub>	0.98 g
1.5 L dist. water – heat to dissolve-cool down– pH 7.0 fill up 2 L– autoclave.	

##### **Binding medium (1L)**

RPMI 1640	16.4 g/l
Glucose 2 %	20 g/l
Fill up with 1 L dist. water – PH 7.2 (NaOH) - sterile filtrate- keep at 4°C	

**Percoll gradient solutions****90% stock**

Percoll solution	45 ml
10x PBS	5ml

**80% solution**

90% Percoll stock	8.9 ml
D-Sorbitol	0.8 g
Culture medium	1.1ml

**60% solution**

90% Percoll stock	6.7 ml
D-Sorbitol	0.8 g
Culture medium	3.3 ml

**40% solution**

90% Percoll stock	4.4 ml
D-Sorbitol	0.8 g
Culture medium	5.6 ml
Sterile filtrate – keep at 4°C	

**3.1.7.2. CHO cell culture****Culture medium**

Ham's F12 mit L-Glutamin und 25 mM HEPES	500 ml
Foetal calf serum (inactivated at 56°C for 45 min)	50 ml
Penicillin/ Streptomycin Mix (100x)	5 ml
Keep at 4°C maximum 3 weeks.	

**CHO-PBS (10x)**

NaCl	80 g	1,37 M
KCl	2 g	27 mM
Na <sub>2</sub> HPO <sub>4</sub> (water free)	14.4 g	101 mM
KH <sub>2</sub> PO <sub>4</sub>	2.4 g	18 mM
Fill up with 1 L dist. water (pH 7.4, Autoclave)		

**FACS-buffer**

In activated foetal calf serum	2 ml
CHO-PBS (1x)	100 ml
Sterile filtrate – keep at 4°C	

**CHO freezing solution**

CHO-culture medium (Ham's F12 with iFCS and Pen/Strep)	45 ml
DMSO (10% – sterile)	5 ml
Vortex, Sterile filtrate – keep at 4°C	

### 3.1.8. *Plasmodium falciparum* isolates

#### IT4 *P. falciparum* isolate

On August 1976, 1 ml of patient blood was obtained from the clinic at MRCL, Fajara-Gambia, West Africa. This was diluted in 10 ml of RPMI 1640 and 10% FCS was shipped on wet ice to New York City, parasite was cultured directly from a human infection into continuous culture using the Petri dish-candle jar technique (Jensen und Trager, 1978), giving line FCR3 or IT4. It was suspected to be mixed with another Brazilian isolate but now only one genome is present. This isolate was friendly supplied from Mo Klinkert, BNITM, Hamburg.

#### 3D7 *P. falciparum* isolate

This isolate is a clone from NF54 isolate which is derived from a patient living near Schipol Airport, Amsterdam, who had never left the Netherlands (Ponnudurai *et al.*, 1981). This isolate was friendly supplied from Mo Klinkert, BNITM, Hamburg.

### 3.1.9. CHO cell line

Chinese hamster ovary cells were established in 1957 in the laboratory of Dr. Theodore T. Puck from 0.1 gram of ovary tissue of a Chinese hamster (Puck 1958). This cell line is xylosyltransferase I deficient pgsA-745 clone from ATCC (American Type Culture Collection). Human endothelial receptors were expressed on the surface of transgenic CHO cells as a protein fused to GFP which is used as a marker. These were friendly supplied from Birgit Föster – Molecular Medicine, BNITM, Hamburg.

Name	Vector	Receptor
CHO-745-AcGFP-clon3	pAcGFP-N1	mock
CHO-745-CD36-AcGFP-clon3	pAcGFP-N1	CD36
CHO-745-ICAM1-clon3-AcGFP	pAcGFP-N1	ICAM-1
CHO-745-P-Selektin-AcGFP-clon4	pAcGFP-N1	P-Selektin
CHO-745-E-Selektin AcGFP-clon8	pAcGFP-N1	E-Selektin
CHO-745-CD9-EGFP-clon8-3	pEGFP-N1	CD9
CHO-745-CD151-EGFP-clon14-10	pEGFP-N1	CD151

## 3.2. Methods

### 3.2.1. Cell biological methods

#### 3.2.1.1. *P. falciparum* culture methods

##### 3.2.1.1.1. *P. falciparum* culture

Both isolates were cultured according to standard culture methods (Trager and Jensen 1997). Parasites were cultivated in 10 ml Petri-dish with cams in 10% human serum (A<sup>+</sup>) at 5% haematocrit (0<sup>+</sup>) and incubated at 37°C in 5% CO<sub>2</sub>, 1% O<sub>2</sub> and 94% N<sub>2</sub> gas mixture. Medium was changed daily and parasitemia was adjusted according to the excrement's requirements.

##### 3.2.1.1.2. Giemsa stain

In order to estimate the percentage of parasitemia within the culture, 8 µl from the culture were put on a glass slide and smeared to obtain a cell monolayer. The slide was air dried and then fixed with methanol for 20 seconds. The slide was then stained in 10% Giemsa solution for 10 minutes. After air drying, the slide was examined under light microscope.

##### 3.2.1.1.3. *P. falciparum* synchronisation

In order to obtain highly synchronised ring stage parasite culture, parasites were grown until at least 5% ring stage parasitemia was reached. The culture was spun down for 5 minutes at 800 x g, resuspended in eight pellet volumes of 5% D-sorbitol solution and finally incubated at 37°C for 5 minutes. Again culture was spun down for 5 minutes at 800 x g, the supernatant was aspirated and then the pellet was washed one time with culture medium and centrifuged as mentioned above. Finally, the pellet was resuspended in 10 ml medium and cultured as usual.

##### 3.2.1.1.4. *P. falciparum* freezing

To store *P. falciparum* stablate for long time, the culture with at least 5% rings (the more the better) was spun down for 5 minutes at 800 x g. Afterwards, the supernatant was removed then the pellet was resuspended in four pellet volumes of malaria freezing solution. The cell suspension was transferred to cryotubes and stored in -80°C in Mr. Frosty<sup>tm</sup> freezing container until transferred to liquid nitrogen.

### **3.2.1.1.5. Thawing of *P. falciparum***

The frozen cryotube (containing IEs) was thawed in water bath at 37°C. Equal volume of pre-warmed malaria thawing solution (37°C) was added. After centrifugation at 300 x g for 2 minutes the supernatant was removed and the pellet was washed 2 more times with 1 ml malaria thawing solution. Then the pellet was resuspended in 10 ml medium and cultured as usual.

### **3.2.1.1.6. Trophozoite stage separation using percoll gradient**

This method was used for the separation of trophozoite stage of *P. falciparum* parasite that were analysed by electron microscopy. The culture was spun down for 5 minutes at 80 x g, the supernatant was removed then the pellet was resuspended in two pellet volumes of 1xPBS and centrifuged as mentioned above. The pellet was again resuspended in 500 µl 1xPBS. The percoll gradient consists of 1 ml 80%, 60% and 40% percoll solutions. 60 % and 40% were carefully piled over the 80 % layer in a 15 ml sterile falcon tube. The resuspended pellet was piled carefully over the 40% layer. Then the gradient was centrifuged for 5 minutes at maximum speed. After centrifugation the trophozoite stage was recovered from the middle two layers by aspirating these layers and resuspending them in 2x volume of 1xPBS. After that the cell suspension was centrifuged 5 minutes at 800 x g. For electron microscopy examination, the pellet was fixed with 2.5% glutaraldehyde and 2.5% paraformaldehyde in 50 mM cacodylate buffer pH 7.2.

### **3.2.1.2. CHO cells culture methods**

#### **2.2.1.2.1. CHO cell culture**

Either in T25 or T75 culture flasks, the adherent CHO cells were cultivated in 5 ml or 15 ml complete medium, respectively at 37°C and 5% CO<sub>2</sub>. Neomycin (G418; 50 mg / ml; final 0.7 mg / ml) was used for selection. The vitality of the CHO cells and the respective confluence were controlled by an inverted fluorescence microscope. The culture medium was changed every third day or the culture was splitted. To split the cells, first the culture medium was removed and cells were washed with pre-warmed (37°C) 1x CHO PBS. To detach the cells, 400 µl accutase (for T25 flask) (37°C) was added and incubated for five minutes (37°C). The activity of accutase was stopped by addition of complete CHO culture medium, the cells then were splitted as required and cultivated as mentioned above.

#### **2.2.1.2.2. CHO cells freezing**

After detaching the cells by accutase as mentioned above, 5 ml of complete CHO culture medium was added, the cell suspension was centrifuged at 300 x g for two minutes and the pellet was resuspended with CHO freezing solution (Three ml for T25 flask). The cell suspension was transferred to cryotubes and stored in -80 °C in Mr. Frosty<sup>tm</sup> freezing container for few days then stored normally at -80°C.

#### **2.2.1.2.3. CHO cells thawing**

The frozen cryotubes were incubated in water bath at 37°C. The thawed cells were transferred to a 15 ml centrifuge tube and resuspended with 10 ml prewarmed (37°C) CHO complete medium. The cell suspension was centrifuged at 300 g for 2 minutes. The pellet was then resuspended with culture medium and cultured as mentioned above.

#### **2.2.1.2.4. CHO cells transfection**

ICAM-1, P-selectin and E-selectin cDNAs were cloned into pAcGFP1-N1 (Clontech Laboratories), whereas CD9 and CD151 were cloned into pEGFP-N1 (Clontech Laboratories). CHO-745 cells were transfected using Lipofectamine 2000 (Invitrogen) complexed with pAcGFP1-ICAM-1, pAcGFP1-P-selectin, pAcGFP1-E-selectin, pEGFP1-CD9, pEGFP1-CD151, according to the protocol of the manufacturer. Two days before, the cells were seeded in 24 well culture plate at  $2 \times 10^4$  cells per well and resuspended in complete culture medium. On the day of transfection, the desired confluent is 70–90%. Four  $\mu\text{l}$  Lipofectamine<sup>®</sup> Reagent was diluted in 100  $\mu\text{l}$  Opti-MEM<sup>®</sup> Medium. DNA was diluted at 0.16  $\mu\text{l}$  in 100  $\mu\text{l}$  Opti-MEM<sup>®</sup> Medium and incubated for 5 minutes at room temperature. The diluted DNA was added to the diluted lipofectamine (1:1) and incubated for 20 minutes at room temperature. Afterwards, 100  $\mu\text{l}$  from the mixture were added to every well plate containing CHO cells. Respectively, the culture plate was incubated at 37°C and 5% CO<sub>2</sub>. Two days after transfection, 0.7 mg/ml G418 (Biochrom AG) was added to the cultures. G418-resistant cells were harvested and subjected to FACS sorting. (As a positive control an empty vector was used pAcGFP1-N1 (1.17  $\mu\text{g}/\mu\text{l}$  and the negative control was Herring Sperm DNA Solution 1.18  $\mu\text{g}/\mu\text{l}$ ).

#### **2.2.1.2.5. Fluorescence-activated cell sorting (FACS)**

In order to sort the transfected CHO cells that express the receptor of interest on its surface the following steps were done. The culture was detached with accutase as mentioned above then was resuspended in 5 ml CHO culture medium. Thereafter, the cell count was estimated using a Neubauer chamber at least  $1 \times 10^6$  CHO cells was required. The cell suspension was spun down by  $250 \times g$  for 2 minutes, the supernatant was removed, the cells were resuspended in 5 ml FACS buffer and finally transferred to a sterile FACS tube. The next centrifugation step was done at  $4^\circ\text{C}$  by  $250 \times g$  for 5 minutes, then the supernatant was removed by turning over the tube. Again the pellet was resuspended in 5 ml FACS buffer with vortexing and centrifuged as before. One  $\mu\text{l}$  of APC labelled antibody was added and then the cells were vortexed. This was followed by an incubation period of 15 minutes over ice in a dark chamber. Afterwards, washing with FACS buffer was repeated two times as mentioned above. Then finally, the cells were resuspended in 2 ml FACS buffer, filtered through FACS filter, and transferred to a new FACS tube. The tube was then kept on ice until sorting. For this a BD FACSAria cell sorter was used. First the cells showing GFP signal were identified. Then the cells showing both GFP and APC signal were sorted in a 15 ml tube containing CHO culture medium then rapidly transferred to a T25 culture flask and cultivated as usual. For selection G418 was used as mentioned above.

#### **2.2.1.2.6. Immunofluorescence assay (IFA)**

##### **Two days before the assay**

Using a sterile forceps, glass cover slips were placed in 24 well plate. Each cover slip was coated with  $500 \mu\text{l}$  1% gelatine solution (2% gelatine (sigma) with 1x PBS-sterile filtrated). Followed by an incubation step for 30 minutes at  $37^\circ\text{C}$ , then the gelatine was aspirated. CHO cells were seeded at  $2 \times 10^4$  per well and resuspended in complete culture medium with G418.

##### **On the day of assay**

Each well was washed three times with 1x PBS. Two fixation steps were performed to avoid cell damage first 2% PFA followed by 4% PFA ( $500 \mu\text{l}$  was added per well and the plate was incubated at  $37^\circ\text{C}$  for 15 minutes). Followed by washing three

times with 1x PBS. For permeabilisation of the cells, 500 µl 0.2% Triton X-100/PBS was added and the plates were incubated at room temperature for five minutes followed by three times washing in 1x PBS. The wells that were not permeabilised were incubated with 500 µl 1x PBS. For blocking, 500 µl 1% BSA/PBS was used for 30 minutes at 37°C. This was followed by 3 x wash with 0.1 % BSA/PBS. Next, 40 µl/cover slip of the first antibody solution (in 0.1% BSA/PBS) was added for dilution (see page 27). The cover slips were incubated upside down on a parafilm covering a 24 well culture plate in a humid chamber for 60 minutes at 37°C. Later the cover slips were washed 3x with 0.1 % BSA/PBS. Thereafter, the second antibody was added (40 µl of antibody solution in 0.1% BSA/PBS per coverslip) and coverslips were incubated as mentioned above for 60 minutes at 37°C this was followed by three washing steps in 1x PBS and one time with dist. water. Afterwards, the cover slips were then mounted with DAKO fluorescent mounting medium, stored in a dark box and then examined under immunofluorescent microscope. (As a control the CHO cells were stained with the second antibody only using the same steps and during the incubation with the first antibody the control cells were incubated with 0.1% BSA/PBS).

### **3.2.1.3. *P. falciparum* static binding assay**

#### **Two days before the assay (seeding of CHO cells)**

Using a sterile forceps, glass cover slips were placed in 24 well plate. Each cover slip was coated with 500 µl 1% gelatine solution (2% gelatine (sigma) with 1x PBS-sterile filtrated). Followed by an incubation step for 30 minutes at 37°C, then the gelatine was aspirated. CHO cells were seeded at  $3 \times 10^4$  per well and resuspended in complete culture medium with G418.

#### **On the day of assay**

Trophozoite stage parasite cultures were prepared and parasitemia was adjusted to 5% in 1% haematocrit suspended in binding medium. After 30 minutes of incubation in binding medium at 37°C, the CHO-745 transfectants were incubated with binding medium for five minutes at 37°C then the medium was aspirated. Afterwards, IEs were co-incubated with CHO-745 transfectants at 37°C for 1 hour

with gentle mixing every 15 minutes. Thereafter, unbound erythrocytes were gently removed by washing 2x in binding medium, and the cells were fixed with 1% glutaraldehyde in PBS at room temperature for 30 minutes. Cells on coverslips were stained with Giemsa diluted with Weiser buffer (1:10). The cover slips were then mounted with Leica mounting medium and stored until examined under light microscope. The numbers of adherent IEs per 300 CHO-745 cells were counted using a light microscope. Assays were performed in triplicate in three independent experiments.

#### **3.2.1.4. Selection and enrichment of *P. falciparum* over CHO-wild type cells and over receptor of interest**

##### **Before the assay**

The week before the assay, the parasite cultures were prepared so that the parasitemia on the day of assay is at least 10% highly synchronized trophozoite stage.

**For enrichment over CHO wild type cells**, CHO-745 cells were seeded on T75 culture flask(s) two days before so that the confluent on the day of the enrichment reaches at least 70%.

**For enrichment over the CHO-745 transfectant** (expressing the receptor of interest), two days before the assay, three types of T75 flasks were prepared as follows, for two consecutive preabsorbption steps, step 1 1x CHO\_Wild and step 2 1x CHO\_Mock were prepared. For the enrichment, 1x CHO\_Receptor was prepared.

##### **Selection and enrichment (Fig 3.1)**

Parasite cultures rich in trophozoite stage were washed one time with binding medium and again resuspended in binding medium to have 5% haematocrit. CHO cell lines were washed with 10 ml binding medium and incubated for five minutes at 37°C and 5% CO<sub>2</sub>. Afterwards, parasite cultures were co-incubated with the desired CHO cell line (according to the experiment's design) at 37°C and 5% CO<sub>2</sub> for 30 minutes (in case of preabsorbption) or 1 hour (in case of enrichment) with gentle shaking in circular movements in two directions every 15 minutes. Thereafter, the unbound erythrocytes were removed by setting the culture flask in upright position and aspiration of the cell suspension. Then the flask was gently

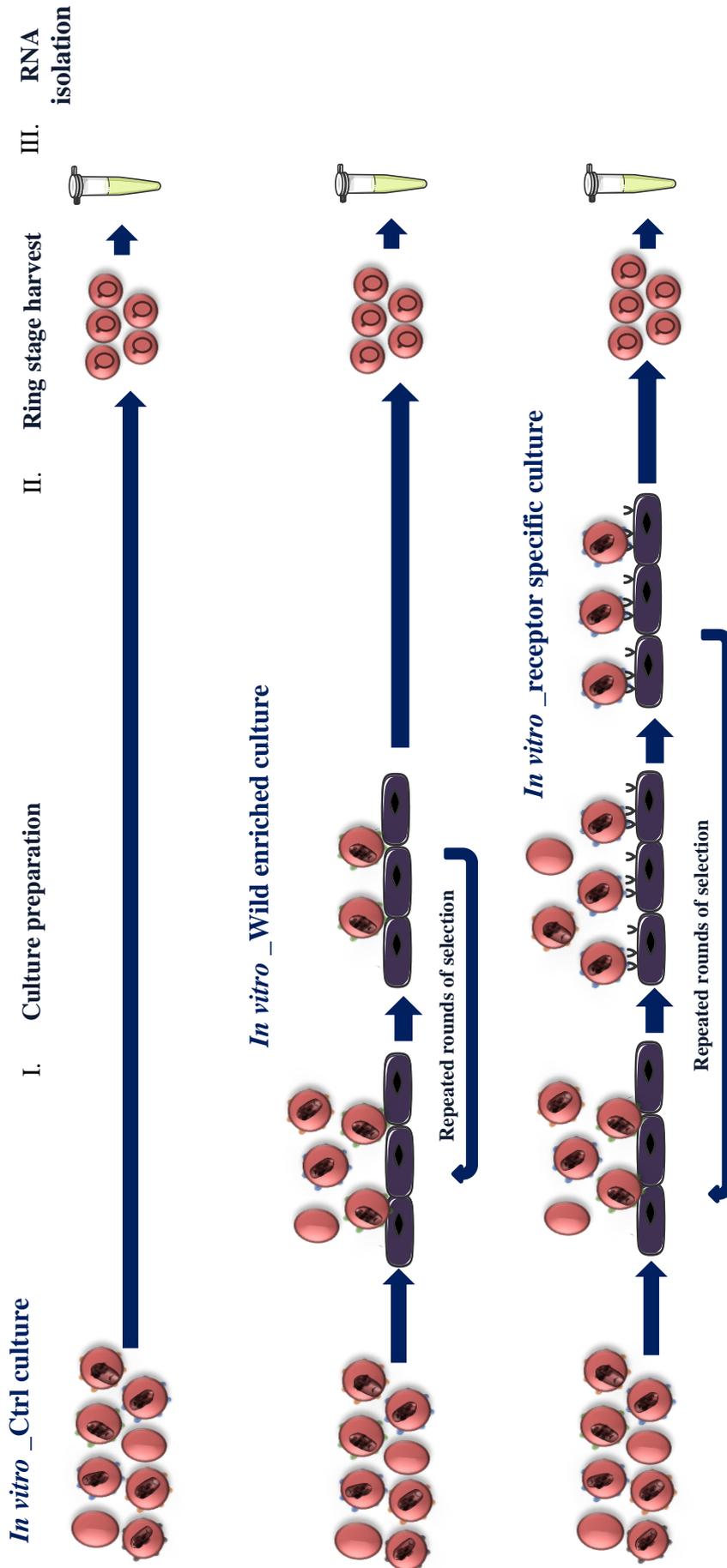
washed eight times with binding medium to get rid of all the unbound cells. The binding of erythrocytes was confirmed by examining under inverted microscope. Finally, *P. falciparum* complete culture medium was added with blood as mentioned above and the flasks were incubated at 37°C in 5% CO<sub>2</sub>, 1% O<sub>2</sub> and 94% N<sub>2</sub> gas mixture until the next day.

### **3.2.1.5. Harvest of *P. falciparum* culture after the selection**

On the next day of selection, the ring stages IEs were harvested as follows, the culture flask was gently shaken in circular movements in two directions and was set in upright position. The cell suspension was then aspirated and spun down for 5 minutes at 800 x g. Next, the pellet was resuspended in fresh pre-warmed (37°C) 7 ml culture medium. In order to separate CHO cells that might be mixed with the IEs culture, Ficoll separating solution was used as follows, 7 ml of sterile pre-warmed Ficoll solution was added in a sterile 15 ml falcon tube then the parasite culture suspension was piled carefully over the Ficoll solution. The gradient was then centrifuged for 20 minutes at 1300 x g without brakes. Erythrocytes have an average density of 1.1 g/ml and therefore they will migrate through the Ficoll solution. Whereas, the density of CHO cells is an average of about 1.05 g/ml, which stop them from migration and leave them suspended between the two layers of the gradient. The supernatant containing the CHO cells was removed and the pellet was washed one time with pre-warmed culture medium. The next step depends on the experiment requirements. After recovery of ring stages, the parasitemia level was estimated with Giemsa stain so that if enough parasitemia was reached for RNA isolation (at least 8%) then the culture was harvested for RNA isolation if not this meant it needed more rounds of enrichments so it would be cultured again as mentioned above until at least 10% parasitemia was reached to be able to repeat the round of enrichment again.

### **3.2.1.6. Harvest using Trizol reagent**

The highly synchronized ring stage IEs culture were harvested from three different stages as shown in Fig. 3.1. Then the pellet was washed 1x with culture medium then it was rapidly lysed in 15 volumes of pre-warmed (37°C) TRIzol and stored at -80 °C.



**Fig 3.1 Working scheme main stages.** First, three different types of culture were prepared (*in vitro* control culture (\_Ctrl), culture enriched over CHO wild type (\_wild) and receptor specific culture, then early ring stage cultures were harvested and finally RNA isolation followed by mRNA sequencing using NGS were performed.

### **3.2.2. Molecular biology methods**

#### **3.2.2.1. RNA isolation**

The harvested lysate from step 3.2.1.6 was divided as 1 ml in 1.5 ml RNase free safe lock tube. Chloroform was then added (200  $\mu$ l) and the tube was closed and vigorously shaken for 15 seconds. Tubes were then incubated for 2-3 minutes at room temperature. Thereafter, the tubes were centrifuged for 30 minutes at 12000 x  $g$  and 4 °C. Quickly, the upper aqueous phase containing RNA was aspirated (~ 400  $\mu$ l) and transferred into new 15 ml RNAase free falcon placed over ice. Directly, equal volume of 70% ethanol was added, then the tube was inverted three times slowly to mix place it over ice. After that PureLink<sup>®</sup> RNA Mini Kit was used according to the protocol of the manufacturer as follows: 700  $\mu$ l from the RNA sample was transferred over the spin column and centrifuged (supplied within the kit) for 15 seconds at 13000 x  $g$  by room temperature. The last step was repeated until all the sample was processed. Washing buffer I was then added (700  $\mu$ l) and centrifuged as mentioned above, then the column replaced in new washing tube. The column was then washed two times with 700  $\mu$ l of Washing buffer II and centrifuged as above mentioned. Then to dry the column completely it was centrifuged for one minute at 13000 x  $g$  by room temperature. The column was then transferred in the recovery tube. The RNA was then eluted two times by adding 25  $\mu$ l RNase free water, then the column was centrifuged for 15 seconds at 13000 x  $g$  at room temperature. Quickly, the RNA containing tube was placed over ice. Nanodrop was used to measure the concentration of RNA as well as the contamination of the RNA with DNA or proteins (for RNA A260/280 ratio ~ 2,0 is accepted). RNA was then stored at -80 °C until further steps were performed.

#### **3.2.2.2. Turbo DNase treatment**

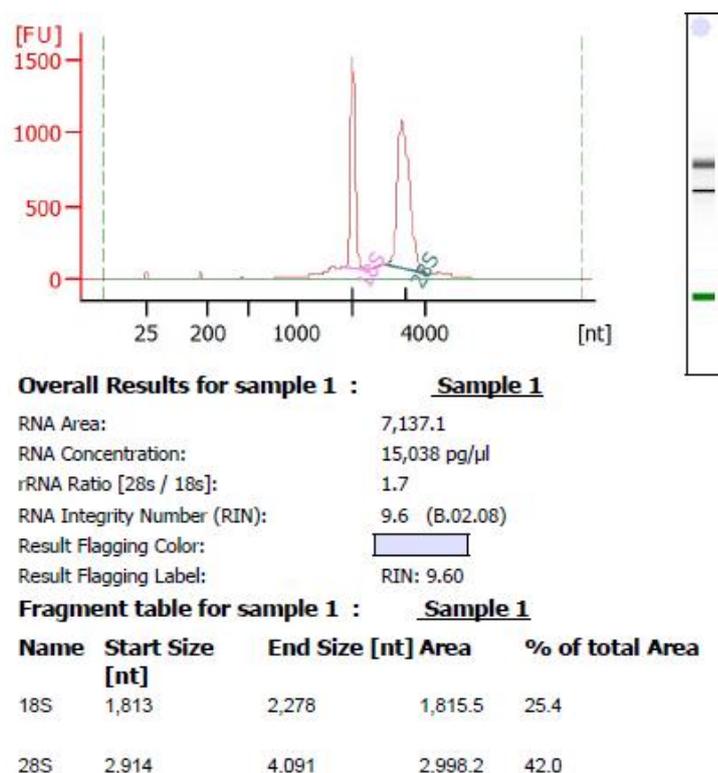
To avoid genomic DNA contamination, RNA samples were treated with TURBO DNase<sup>™</sup> according to the kit protocol as follows. For a volume of 50  $\mu$ l RNA, 5  $\mu$ l 10x Turbo DNase buffer was added as well as 1  $\mu$ l of Turbo DNase. The mixture was mixed gently by pipetting, incubated in 37°C for 20-30 minutes. Then, 5  $\mu$ l from DNase inactivation reagent (resuspended) were added and mixed well. The mixture was incubated at room temperature for five minutes (was mixed occasionally).

Finally, the reaction was centrifuged for 1.5 minutes at 10000 x g in room temperature. The upper aqueous phase containing RNA was transferred to a new RNase free 1.5 ml tube. RNA was kept on ice until the next step.

### **3.2.2.3. Magnetic beads treatment (AGENCOURT® RNAClean up kit)**

The magnetic beads were resuspended by shaking the bottle gently several times. 1.8 x reaction volume was added to the RNA (for 60 µl RNA, 108 µl magnetic beads solution was added). The mixture was mixed by pipetting 15 times, incubated at room temperature for 10 minutes. The tube was then placed on a magnetic stand for 10 minutes (to separate the beads). Slowly, the clear solution was aspirated and discarded while the tube was still on the magnet. 500 µl 70% ethanol were added (without disturbing the beads), incubated for 30 seconds then quickly the ethanol was aspirated and discarded. Ethanol wash was repeated two more times. The reaction plate was then left to dry for 10 minutes. 60 µl of RNase free water were added and mixed 10 times by pipetting (while still on the magnet). Then the reaction was incubated 3-4 minutes until the solution became clear. Then the aqueous phase containing RNA was transferred to a new RNase free 1.5 ml tube. RNA was kept on ice until the next step.

**Agilent 2100® bioanalyzer system** was used to assess the integrity of RNA. The system used chip-based nucleic acid separation technique (electrophoretic separations) (Mueller et al. 2000), where RNA molecules are stained with an intercalating dye then tiny amounts of RNA samples are separated in the channels of the microfabricated chips according to their molecular weight and detected by means of LASER-induced fluorescence. The result is a virtual electropherogram where the amount of measured fluorescence correlates with the amount of RNA of a given size. The integrity of RNA is measured by a software algorithm that allows the calculation of an RNA Integrity Number (RIN) generating a value 1-10. An intact, non-degraded and non-fragmented RNA of very high integrity correspond to RIN 10, whereas a RIN 1 corresponds to RNA completely degraded with minimal quality (Schroeder et al. 2006). According to the kit protocol shortly, total RNA from the samples was diluted and denatured, the RNA pico chip was loaded with gel dye mix, marker and the ladder followed by the RNA samples. Example of a good quality total RNA sample in this study is shown in Fig. 3.2. Degraded RNA samples were excluded from the study see supp. fig. 5/6.



**Fig. 3.2 Bioanalyzer electropherogram summary.** Total RNA was isolated from IT4 *P. falciparum* isolate enriched over CHO cells expressing CD151 on the surface (IT4\_CD151). The 18S-region and 28S-region cover the 18S peak and 28S peak, respectively. RIN is 9.6 which shows a high quality of the extracted RNA.

#### 3.2.2.4. NGS library preparation (steps performed at BNITM for most of the samples).

Next generation sequencing is a method to determine the exact order of nucleotides present in a given DNA or RNA molecule. In the past decade, the use of nucleic acid sequencing was increased, the first generation sequencing known as Sanger sequencing (the chain-termination method), developed in 1975 by Edward Sanger, was considered the gold standard for nucleic acid sequencing, this method had accomplished the Human Genome Project (Sanger et al. 1977). Nowadays NGS platforms provides the researches more advantages than the classic Sanger method. NGS is significantly cheaper, quicker, needs significantly less DNA/cDNA and is more accurate and reliable (Metzker 2010). RNA-Seq (NGS of RNA) provides entire transcriptomic information of a sample without any need of previous knowledge related to genetic sequence of an organism. RNA-Seq provides a strong alternative approach to Microarrays for gene expression studies. Currently RNA-seq has two limitations which are, first, these technologies require micrograms of total RNA to be able to proceed to library preparation steps (Tariq et al. 2011). In this

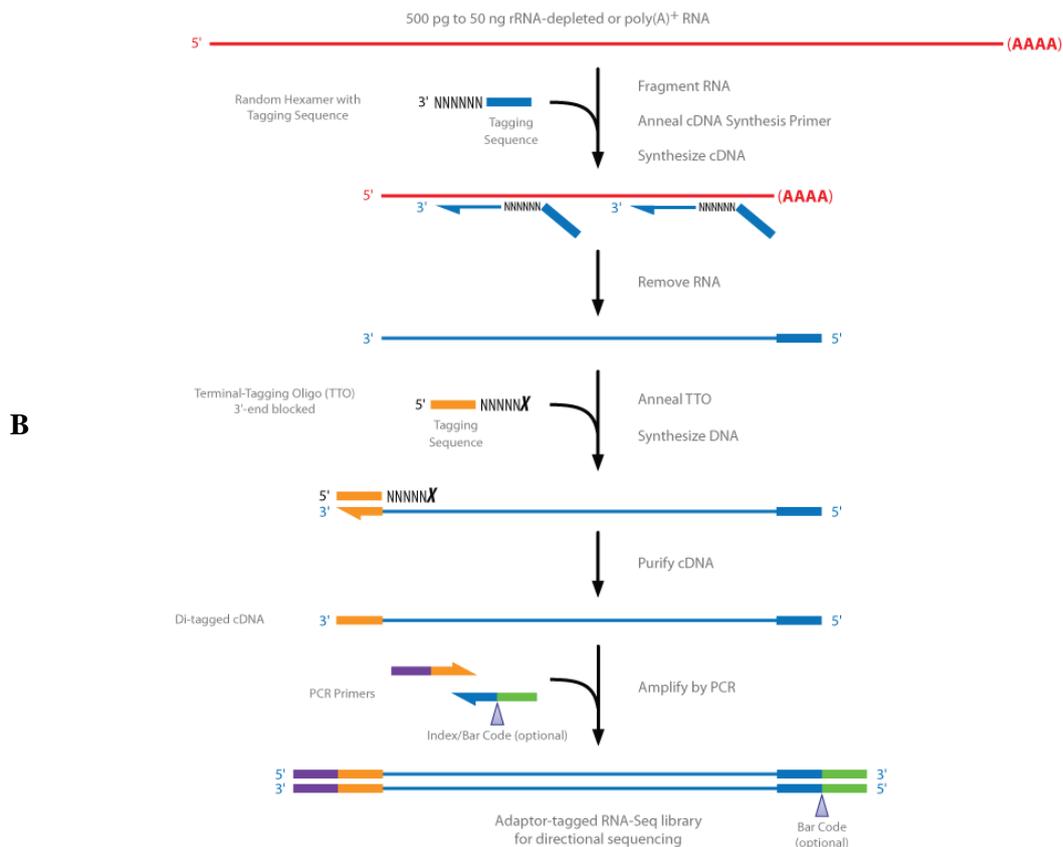
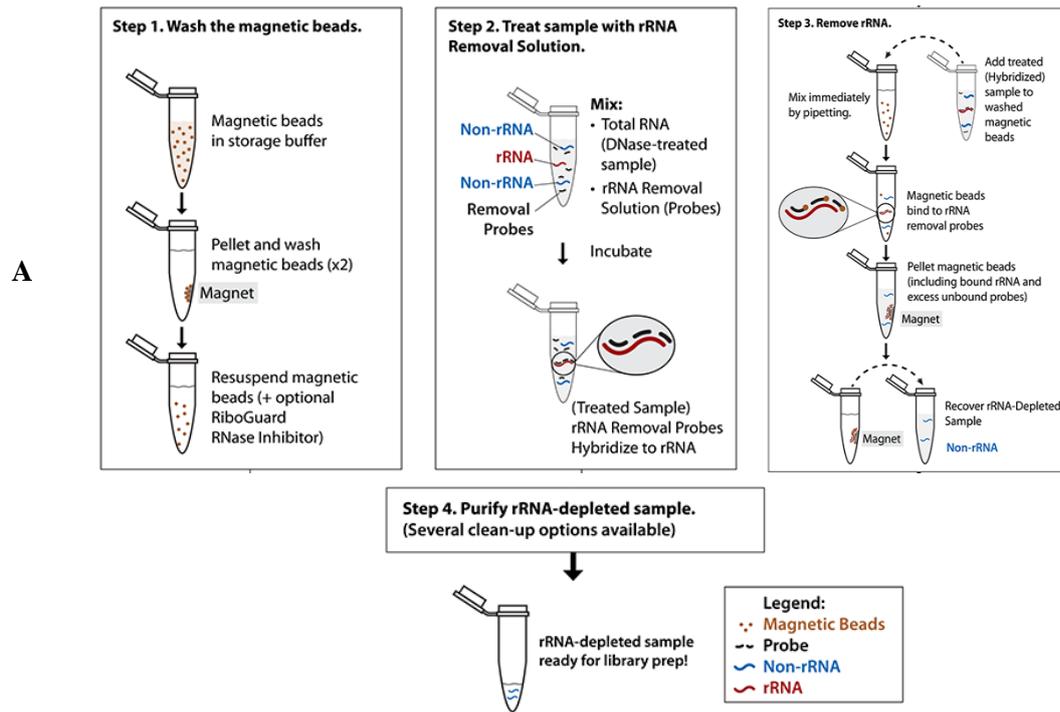
study this was avoided by enrichment process which provided large number of IEs and thus accepted amount of total RNA. It is also important to mention that, recent NGS technology allows the analysis from a single cell but these techniques still very expensive (Gawad et al. 2016). The second limitation was the sequence depth, due to the presence of large percentage of ribosomal RNA within the total RNA (Tariq et al. 2011). In this study the large amount of rRNA reads were avoided by depletion of rRNA using Ribo-Zero™ rRNA removal Kit.

### **1. Ribosomal RNA depletion (Ribo-Zero™ rRNA removal Kit)**

The Kit was used according to the protocol of the manufacturer. Hybridization capture was used to catch rRNA followed by binding to magnetic beads for subtraction (Fig 3.3A) and then a purification step for rRNA depleted sample using RNeasy MinElute Cleanup Kit from Qiagen. By using this method, minimal amount of RNA with the MinElute spin columns based on silica-membrane technology can be recovered. Typically, 2-8% of the total RNA amount was recovered in the rRNA-depleted sample. The quality of the rRNA-depleted RNA was assessed by Agilent 2100® bioanalyzer system using Agilent RNA6000 Pico Chip (2 ng of the rRNA-depleted RNA was loaded in the chip).

### **2. Template preparation**

The cDNA library was constructed and amplified using ScriptSeq™ v2 library preparation kit according to the manufacturer protocol. The main steps were as follows, rRNA-depleted RNA was fragmented and reverse transcribed using random primers containing a 5'-tagging sequence. The 5'-tagged cDNA was then tagged at its 3' end by the terminal-tagging reaction to yield di-tagged, single-stranded cDNA. cDNA was then purified using MinElute™ PCR Purification Kit from Qiagen. After purification, the second strand of cDNA was generated, Illumina adaptor sequences were added followed by addition of Illumina index primers which specifies a barcode for each sample. Finally, the library was amplified by PCR. The cDNA library as purified using AMPure™ XP purification kit which removes the primer dimer that might be formed during the PCR. The library concentration and fragment size were assessed using a Qubit® dsDNA HS Assay Kit and a Bioanalyzer Agilent High Sensitivity DNA Kit.



**Fig. 3.3 (A) Ribo-Zero™ rRNA removal protocol.** 1. Magnetic beads were washed two times. 2. A buffer was added to hybridize with rRNA 3. The hybridized sample was added to the washed magnetic beads which will substrat the rRNA from the total RNA (Figure from Ribo-zero kit user guide). **(B) Principal of ScriptSeq™ v2 library preparation kit** four main steps were done as shown in the figure, 1. RNA fragmentation, 2. cDNA synthesis, 3. 3' terminal tagging 4. Adaptor ligation, Index barcoding and PCR amplification (figure from ScriptSeq kit user guide).

### 3. MiSeq sequencing and imaging

The indexed libraries were pooled together and adjusted as a 4 nM library so that all the samples are equally presented. The pooled library was denaturated and diluted according to the protocol described in the Illumina User guide. The libraries were paired-end sequenced ( $2 \times 150$  bp/read length). MiSeq uses single lane flow cell. Samples are applied to the flow cell through the reagent cartridge sample tube. The library fragments act as a template, of which a new DNA fragment is synthesized. Sequence depends on bridge amplification to form template clusters on a flow cell (Berglund et al. 2011, Quail et al 2012). MiSeq relies on sequencing by synthesis which uses four fluorescently labelled nucleotides to sequence the clusters on the flow cell surface. The fluorescent nucleotides are incorporated into a growing DNA strand, then they are digitally recorded as sequence. Clusters are imaged using LED and filter combinations specific for each fluorescently labelled nucleotide (Quail et al. 2012). Once sequencing is completed raw data were obtained for further analysis.

#### 3.2.3. Bioinformatics methods

##### 3.2.3.1. Quality control of NGS data

In order to assess the overall quality of a sequencing run, an initial QC (quality control) checkpoint was performed using FastQC program. Also biased nucleotide composition and adapters could be detected (Trivedi et al. 2014). The program gives a summary about the total number of sequences processed and percentage of GC content, see supp. fig. 7.

### 3.2.3.2. Data analysis

A generalized data analysis scheme for NGS data included pre-processing of the data to remove adapter sequences and low-quality reads. Reads were aligned to the *P. falciparum* transcriptome using Bowtie 2. Since raw counts are often not directly comparable within and between samples; differential expression (DE) analysis in this study was done using DEseq 1.6.0 Bioconductor package. Thereby, all the counts were normalized using a scaling factor for normalization. A negative binomial model was used to estimate the dispersion and record the relationship between the variance and the mean of each gene. By this way a balanced selection of differentially expressed genes was performed throughout the data (Anders and Huber 2010, Seyednasrollah et al. 2015). Two types of comparison were performed in this study, first Pairwise analysis between the starting *in vitro* culture and culture enriched over CHO wild type cells this was performed to find out the differentially expressed genes up on enrichment of CHO cells. Second, this data set was used in a generalized lineal model to precisely compare all the three elements considered in this study (starting culture, culture enriched over CHO cells and culture enriched over the receptor of interest). Finally, to avoid family-wise error rate, the P-value was adjusted using Benjamine- Hochberg method with FDR 10%. Data analysis was done by the help of Dr. Stephan Lorenzen BNITM.

### 3.2.4. Biotechnical methods

#### 3.2.4.1. Transmission electron microscopy (TEM)

In order to be able to identify the presence and amount of knobs on the surface of the trophozoite stage IEs, percoll gradient was used to separate trophozoite stages as mentioned above see 3.2.1.1.6. This was followed by washing in PBS puffer then the pellet was chemically fixed using 2.5% glutaraldehyde and 2.5% paraformaldehyde. The fixed probes were kept 4°C until further processing.

The following steps were done by **Dr. Katharina Höhn (BNITM)**

First the pellet was washed 2x 50 mM Cacodylate buffer then resuspended in a small volume of 1% OsO<sub>4</sub> (Osmium tetroxide) in H<sub>2</sub>O, incubated 40 min on ice in the dark. After that, it was centrifuged to remove osmium solution and washed 2-3x with H<sub>2</sub>O. The cells were then resuspended in a small volume of 0.5% UA (uranyl acetate) in water and incubated for 30 minutes at room temperature then the tube was centrifuged to remove UA. An ethanol series was used for a serial dehydration, one time ten minutes' incubation in four ascending concentrations (70%, 80%, 90%, 95%) followed by four-time incubation in 100% ethanol each time 15 minutes. The cells were then resuspended in Epon+EtOH ratio 1:1 and incubated for 1-1.5 hours at room temperature. Thereafter, the cells were resuspended in Epon+EtOH ratio 2:1 and incubated for two hours at room temperature. Then the cells were resuspended in pure Epon, divided within different capsules and embedded overnight on a rotating wheel (capsules open). On the following day labels were inserted into the moulds. Then they were incubated in the oven at 60-70 °C for 24-48 hours for polymerization. After that, samples were sectioned in 65 nm thick sections using a Leica EM UC7 and finally the sections were examined with a Tecnai Spirit transmission electron microscope at 80 kV (FEI).

In each population 30 different cells were photographed and analysed for the presence of knobs. ImageJ software was used to determine the circumference of the IEs in the cut section, then the total number of knobs were counted in each IEs and the number was divided by the circumference in µm. Significance was approved using Mann-Whitney test.

## 4. Results

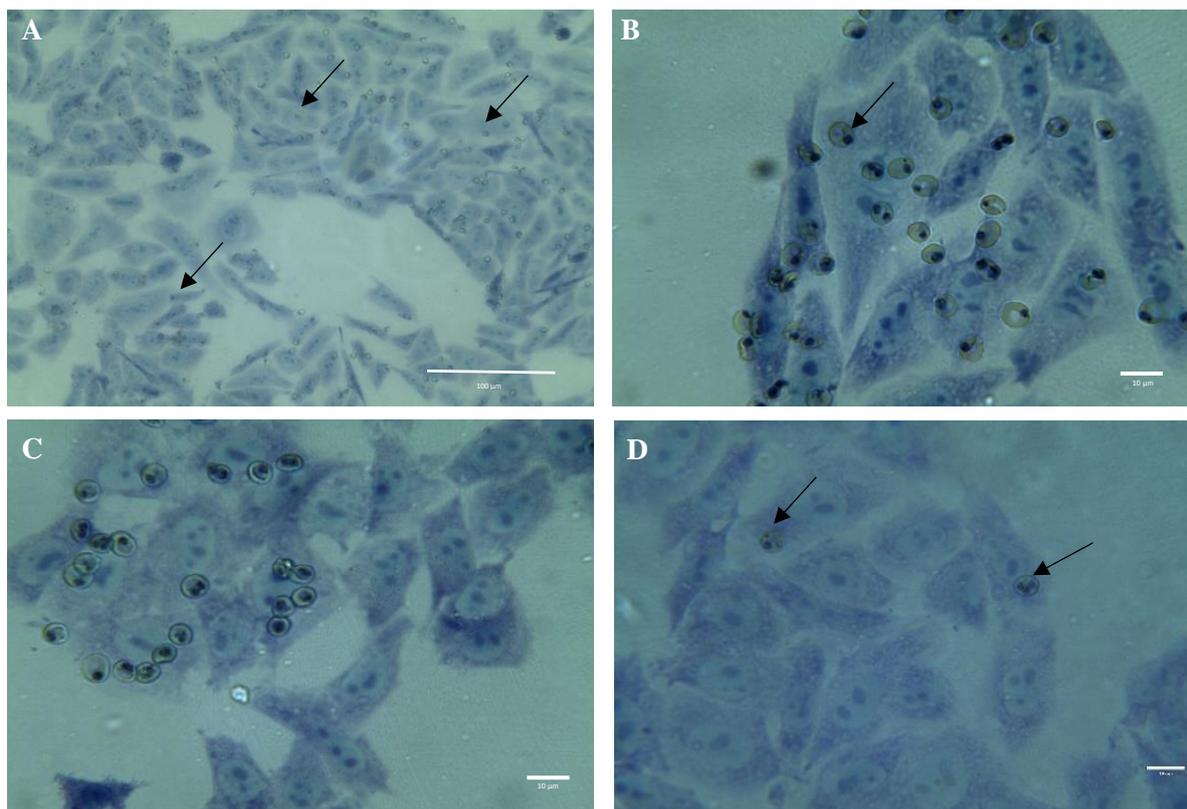
*P. falciparum* has two main mechanisms to evade the host immune system; firstly, presence and switching of variant surface antigens (VSAs) on the surface of the IEs during the course of infection thus evade recognition by host immune response (Abdi et al. 2014, Bull et al. 2008, Duffy et al. 2003, Kyes et al. 2007), secondly, sequestration within the vascular bed of vital organs, which allow the parasite to leave the circulation and avoid the splenic clearance thus surviving within the host (Andrews et al. 2005, Milner et al. 2015, Newbold et al. 1999, Smith 2014, Smith et al. 2000). Cytoadhesion mechanisms known to require an interaction between host cell receptor and a parasite ligand. Various endothelial cell receptors (22 different receptors) were identified to be used by IEs during the cytoadhesion process, reviewed in (Esser et al. 2014, Rowe et al. 2009). Research is ongoing to identify the parasites' ligands to these wide array of receptors. *PfEMP-1* variant antigens are thought to be the main players in the cytoadhesion mechanisms. Studying the molecular basis of IEs-receptors interaction mechanism is considered a very important step in identifying the pathogenesis of this deadly parasite. The aim of this study, is to identify the ligands of *P. falciparum* to different endothelial receptors (ICAM-1, selectins (P-selectin and E-selectin) and tetraspanines (CD9 and CD151).

### 4.1. Identification of the binding capacity of two *P. falciparum* laboratory isolates (IT4 and 3D7) using static cytoadhesion assays.

Firstly, it was important to estimate the binding capacity of the laboratory isolates investigated in this study to the various receptors of interest. Therefore, static cytoadhesion assays were performed using the IT4 and 3D7 *P. falciparum* laboratory isolates. This assay is based on the interaction between IEs (trophozoite stage) and endothelial receptors expressed on the surface of transgenic CHO cells (Chinese hamster ovary cells) as a protein fused to GFP which is used as a marker. CHO cells used in this study are defective in glycosaminoglycan biosynthesis (CHO-745) and were obtained from American Type Culture Collection (Esko et al. 1985, Esser et al. 2014) The binding assays were performed in triplicates, repeated three times and the average number of IEs per 100 CHO cells was calculated.

#### 4.1.1. Static cytoadhesion assays of IT4 *P. falciparum* isolate.

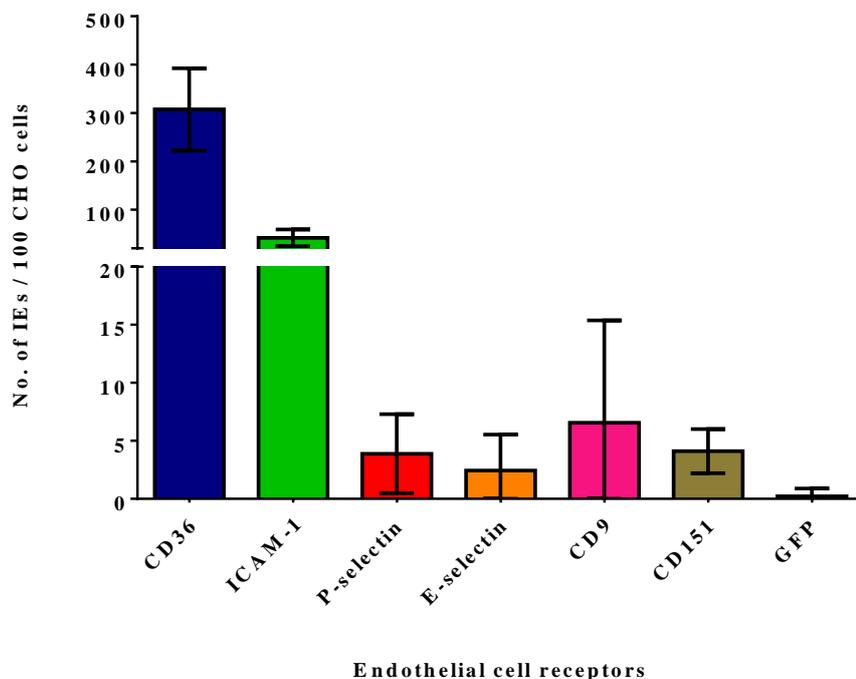
The binding capacity of IT4 *P. falciparum* *in vitro* culture to the endothelial cell receptor CD36 was very high (Fig. 4.1A/B); in average 307 bound IEs/100 CHO cells could be detected (Fig. 4.2). In contrast, only approximately 40 IEs /100 CHO cells bound to ICAM-1 (Fig. 4.1C, Fig. 4.2). Whereas the binding capacity to other receptors was very low (Fig. 4.2). An example for the binding to E-selectin is shown in Fig. 4.1D.



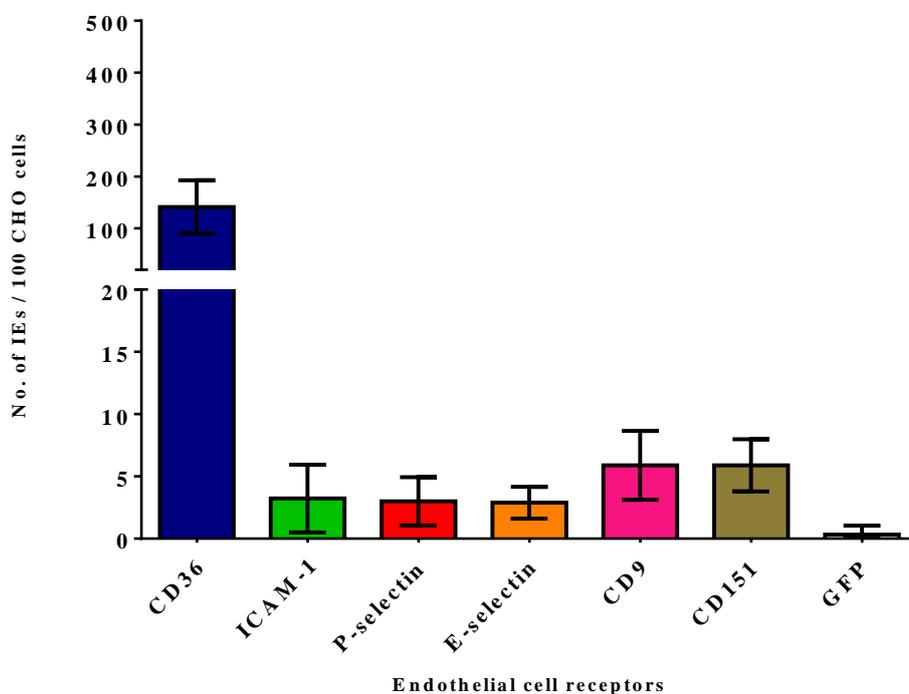
**Fig. 4.1. Cytoadhesion of erythrocytes infected with IT4 *P. falciparum* to CHO cells expressing the receptor of interest on the surface.** After incubation of trophozoite stage IEs over CHO monolayer, the slides were carefully washed, fixed with 1% glutaraldehyde and finally stained with Giemsa. Each slide was then examined under light microscope and the amount of IEs bound to 100 CHO cells was quantified three times for every slide. (A) and (B) show large amounts of IEs binding to a monolayer of CHO cells expressing CD36. A lower binding capacity was observed in case of ICAM-1 (C) and this was further reduced if the binding to E-selectin was investigated (D). Arrows indicated IEs bound to CHO cells.

#### 4.1.2. Static cytoadhesion assays of 3D7 *P. falciparum* isolate.

The binding capacity of 3D7 *P. falciparum* *in vitro* culture to the endothelial cell receptor CD36 was lower than that of IT4 isolate. In average 141 bound IEs/100 CHO cells could be detected. Very weak binding capacity for ICAM-1 was observed with average of 3 IEs /100 CHO. Whereas the binding capacity to other receptors was very low ranging from 3-5 IEs/100 CHO (Fig. 4.3).



**Fig. 4.2. Binding capacity of IT4 *P. falciparum* isolate to various endothelial receptors.** Bars represent the mean  $\pm$ SD of IEs numbers that bound to 100 CHO cells as determined by microscopic inspection and the following was observed: high binding affinity to the endothelial receptor CD36 which is considered the positive control in this assay, whereas GFP refers to the negative control (mock transfected). It was shown that the binding capacity of the IT4 isolate to the ICAM-1 receptor is higher than that to the selectins (P-selectin and E-selectin) and tetraspanins (CD9 and CD151).

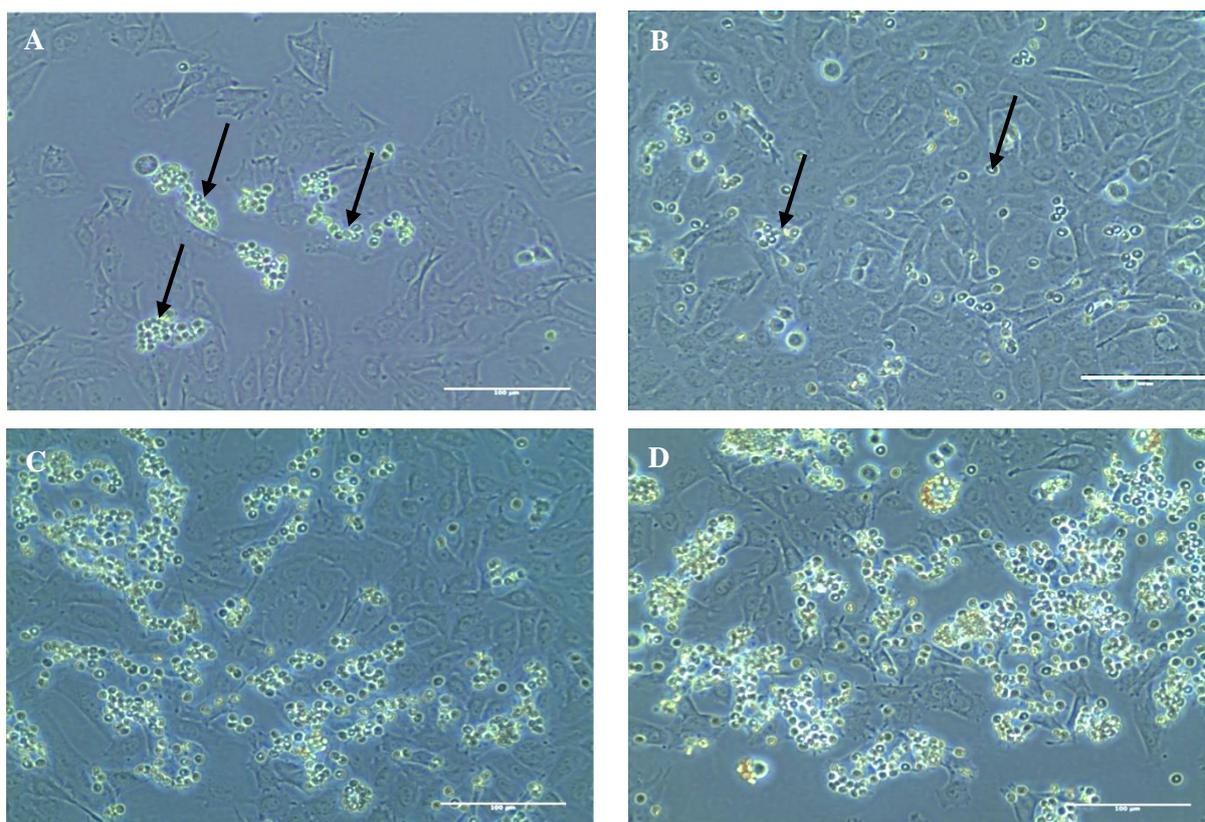


**Fig. 4.3. Binding capacity of 3D7 *P. falciparum* isolate to various endothelial receptors.** Bars represent the mean  $\pm$ SD of IEs numbers that bound to 100 CHO cells as determined by microscopic inspection and the following was observed: high binding affinity to the endothelial receptor CD36 with mean 141 IEs per 100 CHO cells, but lower than that of IT4. Adding to this, it was observed that 3D7 isolate has very low binding capacity to ICAM-1 as well as to the selectins and tetraspanines used in this assay.

## 4.2. Identification of *P. falciparum* ligands for different endothelial cell receptors using gene expression analysis of the selected parasite populations.

### 4.2.1. Initial selection and enrichment of the IEs population that could bind to the receptor of interest.

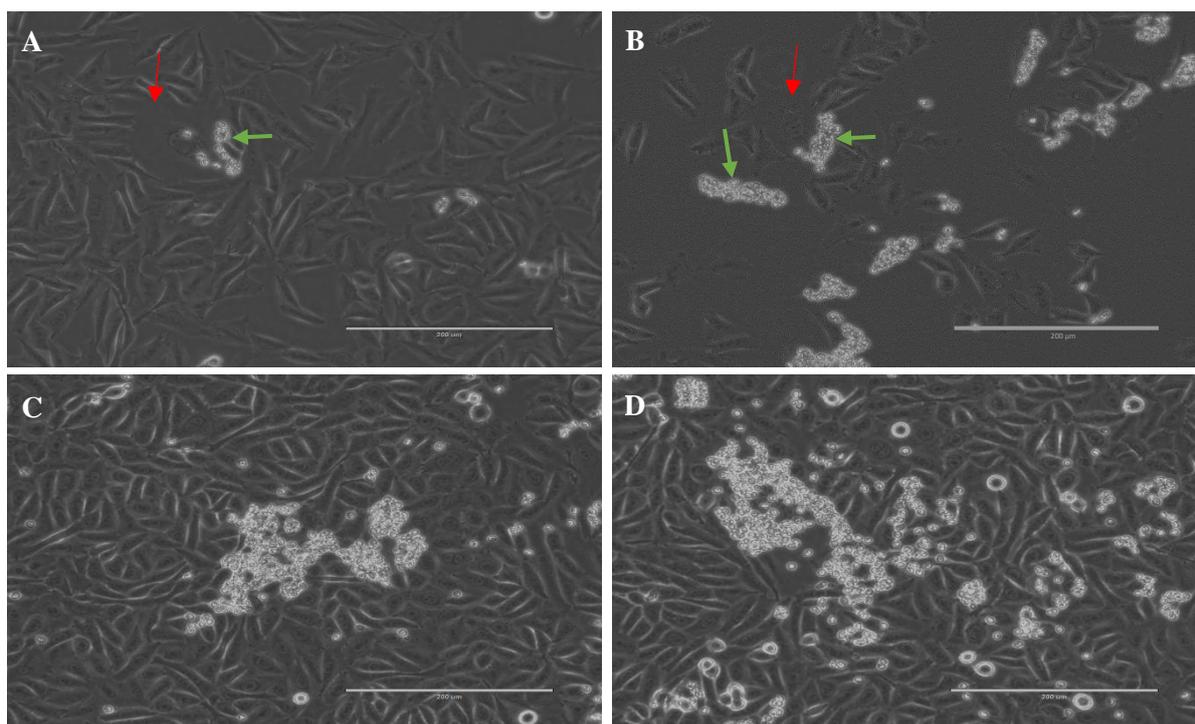
To fulfil the aim of this study, enrichment of parasite populations, that express the ligands for the receptors of interest, was performed using an approach which based on initial selection of the parasite population that have the ability to bind to the respective receptor, followed by continuous enrichment of the target parasite population through repeated rounds of enrichments; once weekly (an example is shown in Fig. 4.4). Putting into consideration that, *var* gene switching in the long term *in vitro* cultivated laboratory isolates is very slow about 2% (Bachmann et al. 2011, Horrocks et al. 2004, Scherf et al. 1998). The aim was to attain a sufficient number of IEs, which facilitates proceeding to RNA isolation followed by mRNA sequencing; to be able to analyse *var* gene expression of the target population using next generation sequencing (Gardner et al.).



**Fig. 4.4. Steps of selection and enrichment of IT4 *P. falciparum* isolate over ICAM-1.** Co-incubation of trophozoite stage IEs over CHO monolayer (expressing the receptor of interest) for 90 min. allowing binding of IEs to the receptor. Next, several steps of washing (5-8 times) to get rid of the unbound IEs was done. Then the culture was examined under an inverted microscope. Finally, culture medium and new erythrocytes were added to permit further cultivation of the selected IEs. The first round of selection was always accompanied by recovery of minimal amount of IEs. **(A)** arrows point to IEs binding to CHO cells. Performing the enrichments for two times, results in increase in the amount of bound IEs. **(B)** Subsequently, the repetition of enrichment rounds resulted in a gradual increase of the number of selected parasite population after 4<sup>th</sup> round **(C)** and 5<sup>th</sup> round, respectively **(D)** (all the co-incubation steps were done at 37°C for 75 minutes with 5% CO<sub>2</sub>).

#### 4.2.2. Enrichment of *P. falciparum* IEs to CHO wild type cells

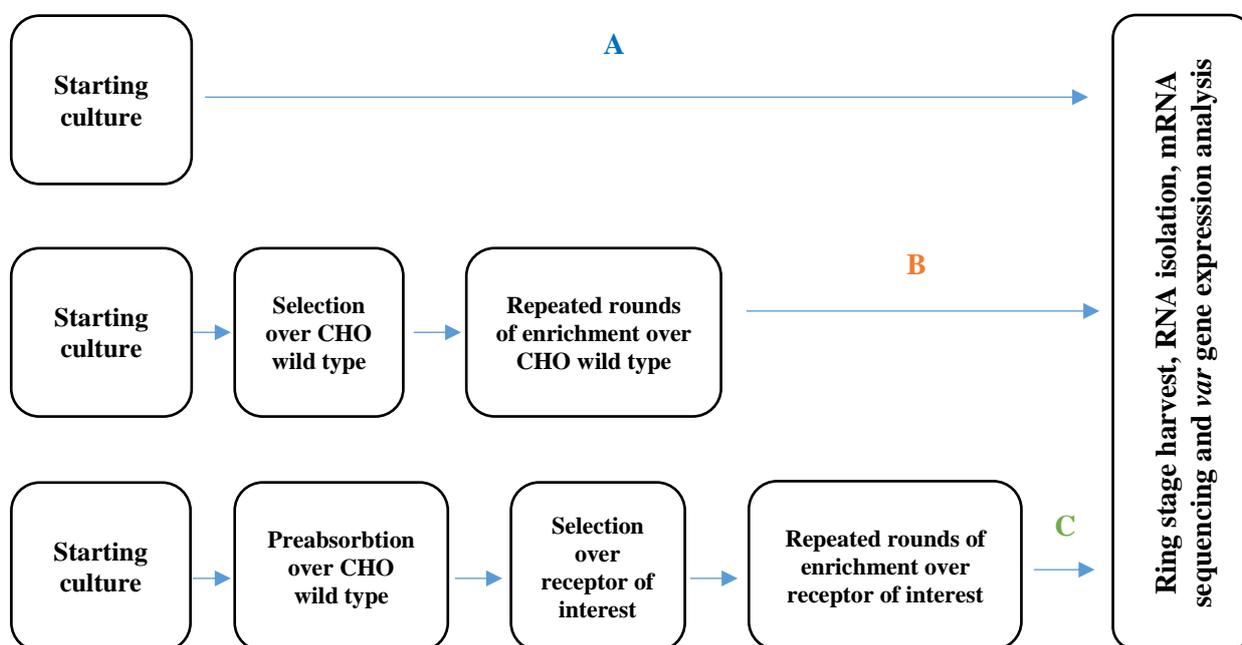
After the 3<sup>rd</sup> round of enrichment, Binding of IEs to the CHO wild type cells, was observed during the pre-absorption step (which was performed before the enrichment over the CHO cells expressing the receptor of interest). A very interesting observation was the accumulation of IEs over large senescent CHO cells (Fig. 4.5A/B). Upon repetition of enrichment rounds, the amount of IEs that bound during the pre-absorption step increased. As a result, selection and enrichment over CHO wild type cells was performed; to be able to monitor the difference between IEs that could bind specifically over the receptor of interest and those that bind to - so far- the unknown surface proteins/structures of CHO wild type cells (Fig 4.5). At the first selection round, few amount of IEs could be observed, upon repetition of enrichment rounds the amount of IEs increased dramatically and four different biological probes were isolated for NGS.



**Fig. 4.5. Enrichment of IT4 *P. falciparum* isolate over CHO wild type cells.** (A) in the first round of selection over CHO wild type cells, IEs (green arrow) were observed to bind over one large morphologically abnormal CHO cell (red arrow). A dramatic increase in the amount of binding IEs upon repetition of enrichment rounds was observed see; 2<sup>nd</sup> (B), 3<sup>rd</sup> (C) and 4<sup>th</sup> (D) round, respectively (all co-incubation steps were done at 37°C for 75 minutes with 5% CO<sub>2</sub>).

### 4.2.3. Establishing the final protocol for selection and enrichment of different populations of IEs

According to the above mentioned observations, the selection and enrichment protocol was modified and consequently three fundamental IEs populations were prepared. The first population was the starting *in vitro* culture (Fig. 4.6A), which theoretically represent the overall IEs populations. Starting culture was synchronised and early ring stages were harvested for RNA isolation. The second population was included the IEs that bind to the CHO wild type cells (Fig. 4.5/4.6B). Initial selection was done over CHO wild type cells followed by repeated rounds of enrichments until considerable amounts of IEs could be recovered for early ring stage harvest and RNA isolation. Finally, the third IEs population (Fig. 4.5C) which subsequently are divided to five different populations over the five receptors of interest analysed in this study (ICAM-1, P-selectin, E-selectin, CD9 and CD151) (Fig. 4.6). Identification of the IEs receptor-specific populations was preceded by a pre-absorption step before every enrichment round to minimize the non-specific binding and trying to specifically select the target IEs populations (Fig. 4.6C). Again, the selection of IEs over the corresponding receptor was repeated once weekly until plentiful IEs were observed binding to CHO cells. This process may take about two months of continuous cultivation and was terminated by the harvest of early ring stages and RNA isolation from different biological samples.



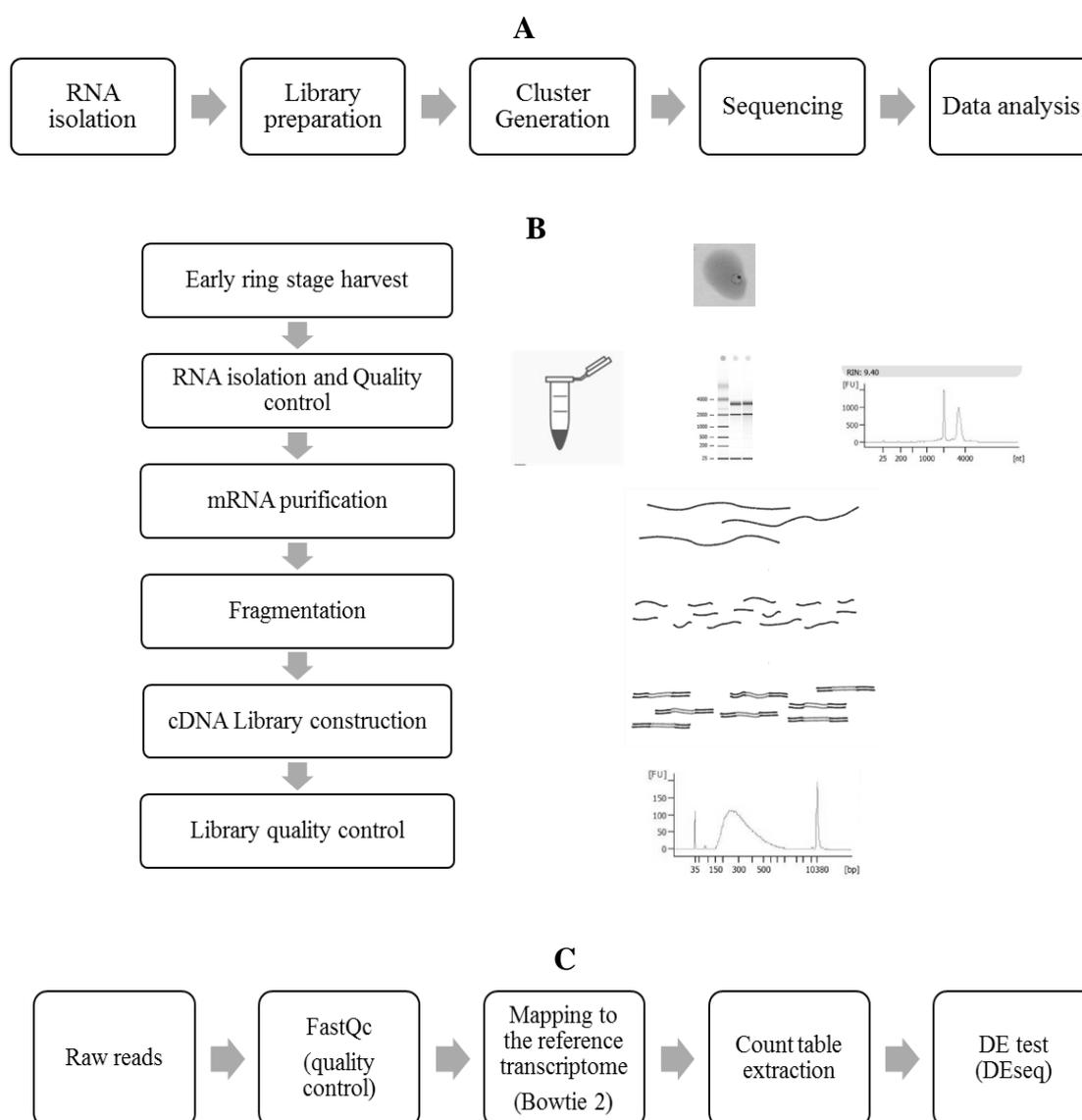
**Fig. 4.6. Schematic presentation of the preparation and enrichment of different IEs populations for RNA isolation.** Three different parasite populations were prepared, synchronised and finally early ring stages (6-8 hours post infection) were harvested for RNA isolation. These are (A) *P. falciparum in vitro* culture, (B) *P. falciparum in vitro* culture enriched over CHO wild type cells. Finally, (C) receptor specific IEs populations which were initially pre-absorbed over CHO wild types, followed by selection and further enrichment over the receptors of interest (all the co-incubation steps were done at 37°C for 75 minutes with 5% CO<sub>2</sub>).

#### 4.2.4. NGS sequencing and transcriptome analysis of the different *P. falciparum* cultures

High-throughput sequencing technologies are considered nowadays a very strong tool in comprehensive transcriptome analysis with simple and clear steps (Fig. 4.7A). Taking these advantages two Illumina platforms were used in this study MiSeq at BNITM (IT4 samples) and Hiseq 4000 at BGI TECH SOLUTIONS (3D7 samples).

Fig 4.7B shows the steps that were done for sample preparation. RNA harvest was done from highly synchronised early ring stage IEs (6-10 hours after infection; high peak of *var* gene expression is expected during this stage (Bachmann et al. 2012, Llinas et al. 2006) with at least 8-10% ring stage parasitemia to ensure enough quantity of RNA suitable to be processed for NGS (2–5 µg total RNA is needed) (Tariq et al. 2011). Harvested erythrocytes (300 µl) were rapidly lysed in 15 volumes of pre-warmed (37C°) TRIzol and stored at -80 C°. Total RNA was isolated using PureLink™ RNA Mini Kit according to manufacturing instructions. Quality and Quantity of total RNA was assessed using Agilent™ 2100 Bioanalyzer System by using Agilent™ RNA 6000 Pico kit. Degraded RNA samples were excluded from the study. To avoid genomic DNA contamination, samples were treated with TURBO™ DNase followed by magnetic beads enzymatic wash using Agencourt RNA Clean XP™. Before processing again, the integrity of RNA was proved using Agilent™ 2100 Bioanalyzer System. Total RNA include about 80% ribosomal RNA (rRNA). Consequently, if rRNA was not removed, the majority of the final sequence reads would be from rRNA (Tariq et al. 2011). Therefore, to ensure deep sequencing of mRNA a very important step is to deplete the rRNA from the total RNA. To accomplish this, Ribo-Zero™ rRNA removal Kit was used according to manufacturing guides. The rRNA depleted RNA samples were then purified using RNeasy™ MinElute Cleanup Kit. The concentration of rRNA as well as the quality of mRNA in the treated samples were assessed using Agilent 2100™ Bioanalyzer System (Agilent™ RNA 6000 Pico kit for mRNA assay). Illumina MiSeq sequencing platform is capable of providing only relatively short sequence reads; the average insert length should not exceed 600 bp. Therefore, a fragmentation step is incorporated to improve sequence coverage over the transcriptome (larger fragments don't hybridize to the MiSeq flow cell as well as smaller ones) (Liu et al. 2012, Quail et al. 2012). Fragmentation and cDNA library

construction was done by using ScriptSeq™ v2 RNA-Seq Library Preparation Kit according to the kit protocol. Abundance and size distribution of the library was checked using Agilent™ 2100 Bioanalyzer System (Agilent™ High Sensitivity DNA Kit). Libraries were diluted to 4nM each and then pooled so that each library is equally distributed. Fig 4.7C covers the general steps that were done for data analysis. Upon successful NGS run a general quality control step was performed using FastQC program. This was performed to check the overall quality of the reads, biased nucleotide composition and adapters, see supp. fig. 7. Reads were aligned to the *P. falciparum* transcriptome using Bowtie 2 software program.



**Fig. 4.7. General steps and work flow adopted in this study.** (A) Schematic presentation of NGS general approach. Both Illumina MiSeq and HiSeq 4000 platforms was used in this study. (B) Schematic presentation of RNA isolation and library preparation workflow. (C) Schematic presentation of data analysis workflow.

Since raw counts are often not directly comparable within and between samples; differential expression (DE) analysis in this study was done using DESeq 1.6.0 Bioconductor package. Taking the advantages which is provided by this software for normalisation method; with the hypothesis that most genes are not DE, so they should have similar reads in all samples.

DESeq estimates the variance considering both the coverage within each library (size factors), the expression strength under each condition, and the per-gene variance. Library coverage is determined by comparing the ratio of gene counts between replicate experiments. Expression strength is determined by the mean counts for a gene for each condition (Anders and Huber 2010).

The per-gene variance is assumed to be a function of the mean that is approximated by empirical fit to the data. The underlying idea is that non-DE genes should have similar read counts across samples, leading to a ratio of 1. Assuming most genes are not DE, the median of this ratio for the lane provides an estimate of the correction factor that should be applied to all read counts of this lane to fulfil the hypothesis. (Anders and Huber 2010, Dillies et al. 2013). Finally, gene ontology was obtained from PlasmoDB 28.

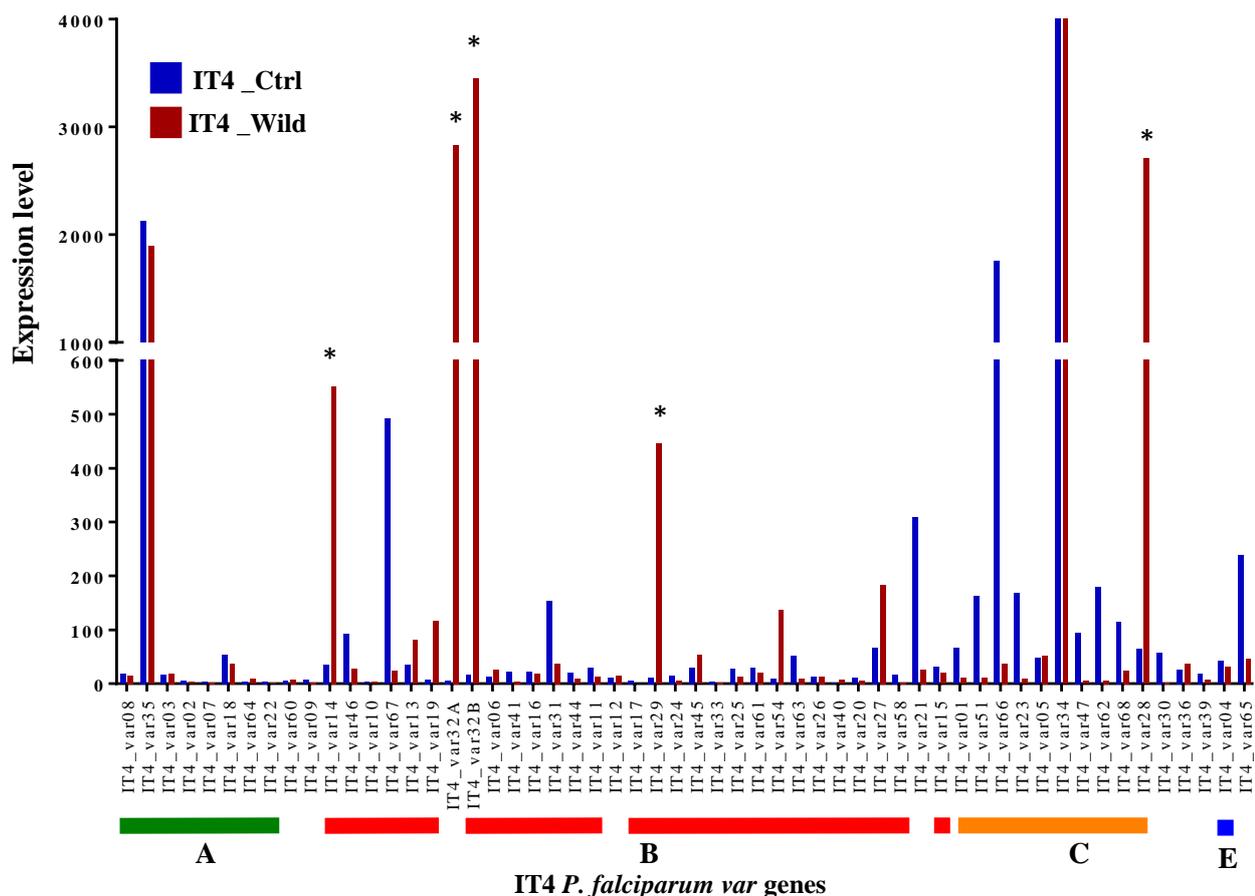
#### **4.2.5. Comparison of *var* gene expression profiles in both *in vitro* cultivated *P. falciparum* isolates and *P. falciparum* cultures enriched over CHO wild types**

As above mentioned, it was possible to enrich IEs population that bind to CHO wild type cells as mentioned in part (4.2.2). This type of binding was also mentioned in another study (Andrews et al. 2005) debating the use of CHO cells in binding assays. It was important to define the IEs populations that could bind to CHO wild type cells in general and overwhelm the receptor specific populations.

Various biological probes were isolated from both IT4\_Ctrl and 3D7\_Ctrl cultures continuously for 2 years to be sure that there was no extreme difference in the expression profile. Both IT4\_Wild and 3D7\_Wild cultures were selected and enriched over CHO wild type cells as mentioned above (part 4.2.3) until significant amount of parasites could be obtained (8-10 % of rings on the next day of enrichment round) to facilitate further sample processing and finally DE analysis as explained in part 4.2.4. Consequently, the *var* gene expression profiles between the starting *in vitro* culture (\_Ctrl) and the culture enriched over CHO wild (\_Wild) types were compared.

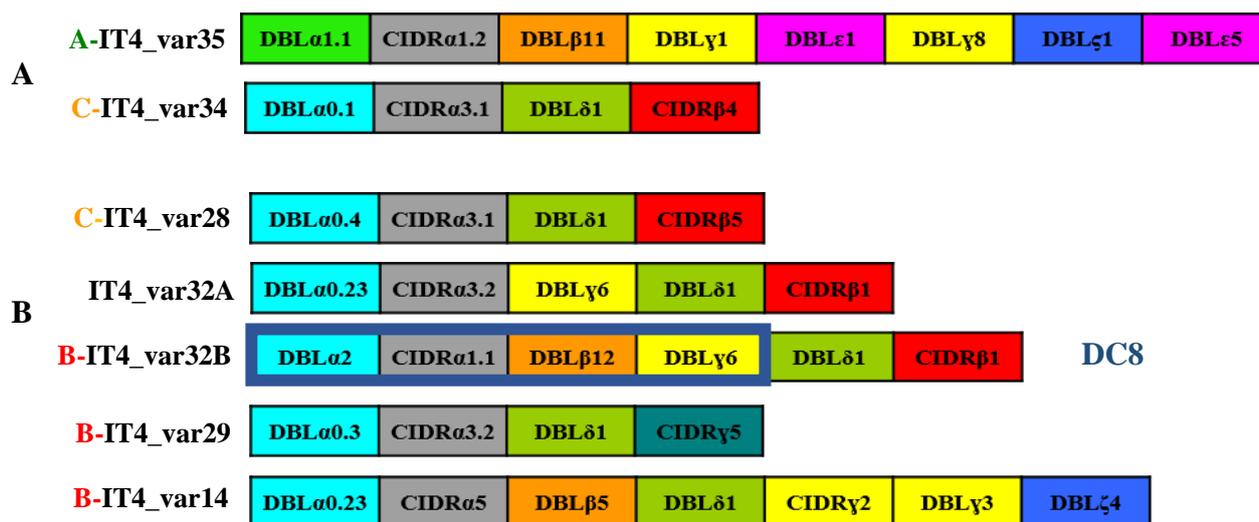
#### 4.2.5.1. *var* gene expression of IT4 *P. falciparum* isolate in both IT4\_Ctrl and IT4\_Wild populations

In both populations two *var* genes were highly expressed (IT4\_var35 and IT4\_var34) (Fig. 4.8). Fig 4.9A shows the schematic protein structure encoded by these *var* genes. IT4\_var35 or var1, is a pseudogene (other IDs: PFIT\_0536800, PFIT\_PFE1640w, or previously var1csa). It seems like its expression falls outside the classical pattern of mutually exclusively expression described of *var* genes. This phenomenon was also described by Kyes and colleagues in thier (Kyes et al. 2007). IT4\_Wild populations showed differentially expression of the *var* genes IT4\_var28, IT4\_var32A, IT4\_var32B, IT4\_var29 and IT4\_var14 compared to the IT4\_Ctrl populations (Fig. 4.8). Fig 4.9B shows the schematic protein structure encoded by these *var* genes. All the variants have the head structure required for binding to CD36. Adding to this IT4\_var32B encodes for a protein which contains Domain Cassette 8 (DC8) which consists of (DBL $\alpha$ 2 CIDR $\alpha$ 1.1 DBL $\beta$ 12 DBL $\gamma$ 6) domains.



**Fig. 4.8.** IT4 *P. falciparum* *var* gene differential expression between two IEs populations IT4\_Ctrl and IT4\_Wild. The columns represent ring stage *var* gene expression profiles, Y axis shows the expression level of 56 *var* genes, whereas X axis is divided to 4 different *var* gene groups (A) (B) (C) and (E). mRNA sequencing was done using the NGS Illumina MiSeq platform. Reads were aligned to the *P. falciparum* transcriptome using Bowtie2 and differential expression was analysed using DESeq. Gene ontology was obtained from PlasmoDB 28.

\* significant; P-value was corrected with Benjamini Hochberg method with FDR 10%.

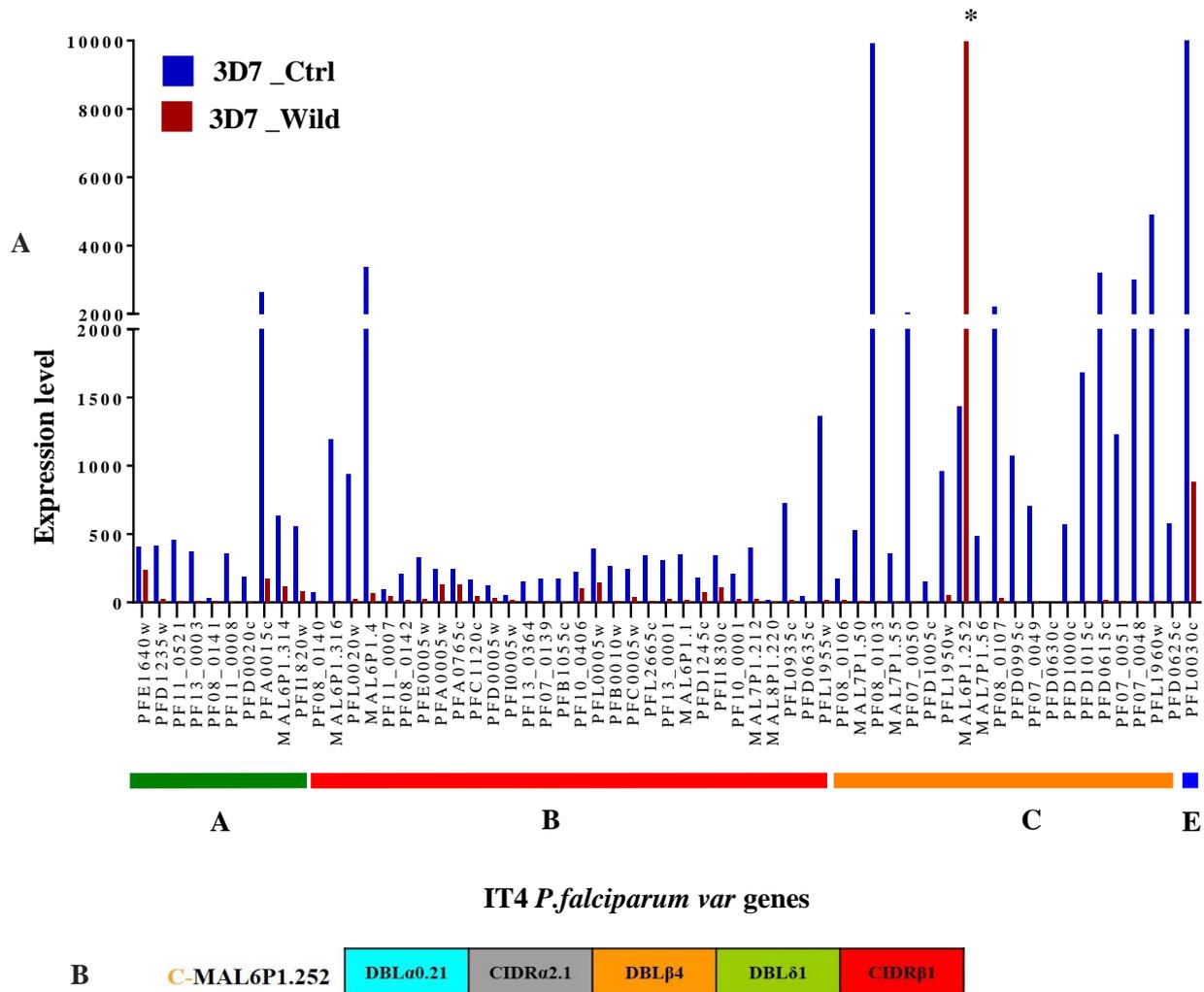


**Fig. 4.9.** Schematic domain structure of (A) *PfEMP1* proteins encoded by *var* genes IT4\_var35 (group A) and IT\_var34 (group C). Both genes were equally expressed in both parasite populations (IT4\_Ctrl and IT4\_Wild).

(B) *PfEMP1* proteins encoded by *var* genes which were differentially expressed in IT4\_Wild populations compared to IT4\_Ctrl ones. It could be observed that they contain the classic head structure (DBL $\alpha$  and CIDR $\alpha$ ) domains which are described in literature as binding domains for endothelial cell receptor CD36. IT4\_var32B showed the structure of DC8. Mapping of domain structure was done according to (Rask et al. 2010).

#### 4.2.5.2. *var* gene expression in 3D7 *P. falciparum* isolate in both 3D7\_Ctrl and 3D7\_Wild populations

3D7 *P. falciparum* isolate used in this study was selected and enriched for binding to CHO wild type cells as mentioned in part 4.2.2; to be able to define the IEs populations that could bind to CHO cells and compare them later in this study with those which specifically bind to the receptors of interest. 3D7\_Wild populations showed only one *var* gene that was differentially expressed MAL6P1.252 ( $\text{Padj} \leq 0.05$ ) compared to 3D7\_Ctrl populations (Fig. 4.10A). Fig. 4.10B shows the schematic protein structure encoded by MAL6P1.252 which contain the head structure required for binding to CD36. The whole transcriptome profile analysis of 3D7\_Wild IEs population showed surprisingly differential gene expression of 50 *rif* genes see supp. table 13.



**Fig. 4.10. 3D7 *P. falciparum* var gene differential expression between two IEs populations 3D7\_Ctrl and 3D7\_Wild.** (A) The columns represent ring stage *var* gene expression profiles, Y axis shows the expression level of *var* genes, whereas X axis is divided to 4 different *var* gene groups (A) (B) (C) and (E). Reads were aligned to the *P. falciparum* transcriptome using Bowtie 2 and differential expression was analysed using DESeq. Gene ontology was obtained from PlasmoDB 28. \*: Significant; P-value was corrected with Benjamini Hochberg method with FDR 10%. (B) **Schematic domain structure of *PfEMP1* protein encoded by *var* gene MAL6P1.252** which was expressed in 3D7\_Wild enriched populations. Mapping of domain structure was done according to (Rask et al. 2010).

#### 4.2.6. Comparison of different transcriptome profiles of differently selected cultures

##### 4.2.6.1. ICAM-1 ligands identification as a proof of concept

ICAM-1 (Intra cellular adhesion molecule-1) is an endothelial cell receptor which play a role during inflammation through facilitating the migration of leukocytes to the site of inflammation (Brown et al. 2013, Frank and Lisanti 2008). In previous studies, it was suggested to be involved in malaria pathogenesis; ICAM-1 was accumulated together with IEs in brain vessels of post-mortem samples from CM patients (Turner et al. 1994). In addition, some studies reported increase in the binding capacity to ICAM-1 receptors of IEs samples obtained from patients with severe malaria (Ochola et al. 2011, Newbold et al. 1999). The exact role of ICAM-1 in cerebral malaria is still a big issue in research. DBL $\beta$  domains of *PfEMP-1* variants mediates the binding to ICAM-1 are identified and described in details in the literature (Bengtsson et al. 2013, Smith et al. 2000). Owing to these facts, ICAM-1 ligand's identification was used in this study as a proof of concept. IT4 *P. falciparum* IEs were selected for ICAM-1 binding as mentioned above (part 4.2.3) (**but without the preabsorbtion step**). Biological probes were harvested at early ring stage after reaching at least a parasitemia of 8-10%. In case of ICAM-1, it took only 3-4 weeks of enrichment (3-4 rounds of selection). RNA isolation and mRNA sequencing using NGS were performed as mentioned above (part 4.2.4). Data analysis was performed using DESeq<sup>®</sup> Bioconductor package to determine the abundance of the differentially expressed genes in the three populations. A generalised lineal model (GLM) was used to fit the normalized values.

##### 4.2.6.1.1. Identification of IT4 *P. falciparum* ligands mediating binding to ICAM-1

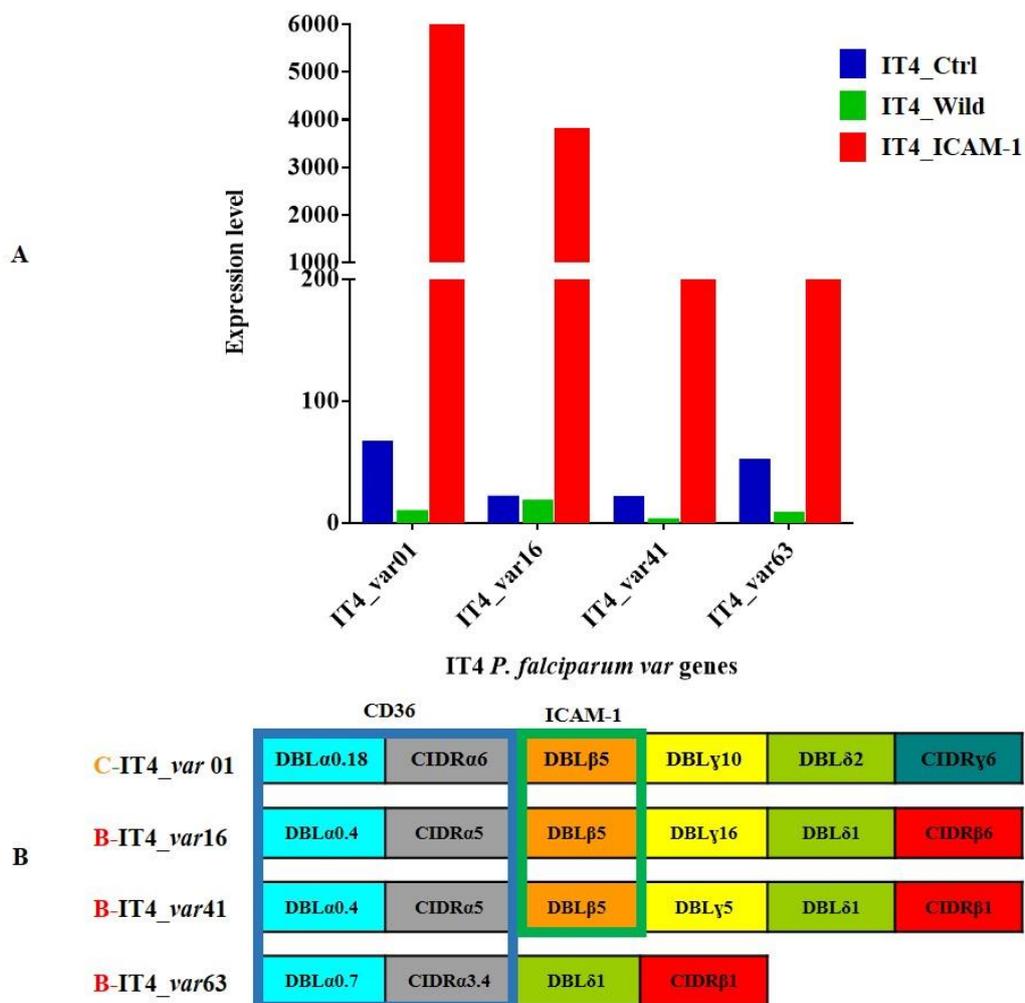
Table 1 represents part of the GLM data analysis done to identify the ICAM-1 ligands in the parasite population IT4\_ICAM-1. It was observed that the IT4\_var01 gene was the most highly expressed gene with mean expression level of 55911.55 in the IT4\_ICAM-1 population compared to the other 2 populations IT4\_Ctrl and IT4\_wild (67.53674 and 10.51116, respectively) with  $P_{adj} \leq 0.0001$  (Fig. 4.11A). Schematic domain structure of the *PfEMP-1* variant encoded by IT4\_var01 showed that it contains a DBL $\beta$ 5 domain which is described in previous studies as an ICAM-1 binding domain (Bengtsson et al. 2013, Smith et al. 2000) Adding to this, it also contains DBL $\alpha$ 0.18 and CIDR $\alpha$ 6 responsible for CD36 binding (Fig. 4.11B) (Smith et al. 2000).

In addition, IT4\_var16 and IT4\_var41 genes were highly expressed in the IT4\_ICAM-1 population with mean expression level of 3790.63 and 567.8845, respectively ( $P_{adj} \leq 0.0001$ ). Both *PfEMP-1* variants encoded by the two genes contain DBL $\beta$ 5 domain as well as CD36 binding domains (Fig. 4.9B). Looking up for the var genes that were highly expressed in the IT4\_wild IEs population (part 4.2.5.1) IT4\_var28, IT4\_var32A and IT4\_var32B were found to be low expressed in IT4\_ICAM-1 IEs population which signifies the efficacy of this system in ligands identification.

Analysing the whole transcriptome of IT4\_ICAM-1 parasite population revealed that from the 140 *rif* gene candidates which encodes RIFIN surface proteins only type A PFIT\_0616600 gene was highly expressed compared to other 2 populations. Interestingly, this gene is located on chromosome 6 near to position of IT4\_var01 which is the highly expressed *var* gene in this IEs population. On the other hand, neither *stevor* nor *Pfmc-2tm* genes was found to be highly expressed. A list of the genes which were found differentially expressed in IT4\_ICAM-1 population is attached in supp. table 4.

**Table 1. GLM mathematical model for differential *var* gene expression in the three IEs enriched populations**

var genes	IT4_Ctrl Mean expression	IT4_wild Mean expression	IT4_ICAM-1 Mean expression	X-interception	CHO true	ICAM-1 true	deviance	convergence	P value	P adjustment
IT4_var01	67.53674	10.51116	55911.55	6.075681336	-2.90695362	12.60212671	2.112549128	TRUE	<0.0001	<0.0001
IT4_var16	22.4147	18.94426	3790.63	4.461296603	-0.65011713	8.07693219	7.596047094	TRUE	<0.0001	<0.0001
IT4_var41	22.16423	3.902658	567.8845	4.433083287	-1.92603794	6.642156541	4.976719788	TRUE	<0.0001	<0.0001
IT4_var63	52.65066	9.40192	660.2013	5.69613298	-2.05399622	5.724344317	6.645582205	TRUE	<0.0001	<0.0001
IT4_var28	65.568096	3206.219	73.91961	6.033283887	5.613056444	-5.44075911	2.244453999	TRUE	0.000115864	0.007068
IT4_var32B	17.055281	3536.398	11.98239	4.092855127	7.694764193	-8.205799152	1.382329373	TRUE	0.000280186	0.016811
IT4_var32A	5.544333	2904.98	9.07756	2.48756282	9.016331138	-8.33592307	1.615680923	TRUE	0.000542202	0.03199

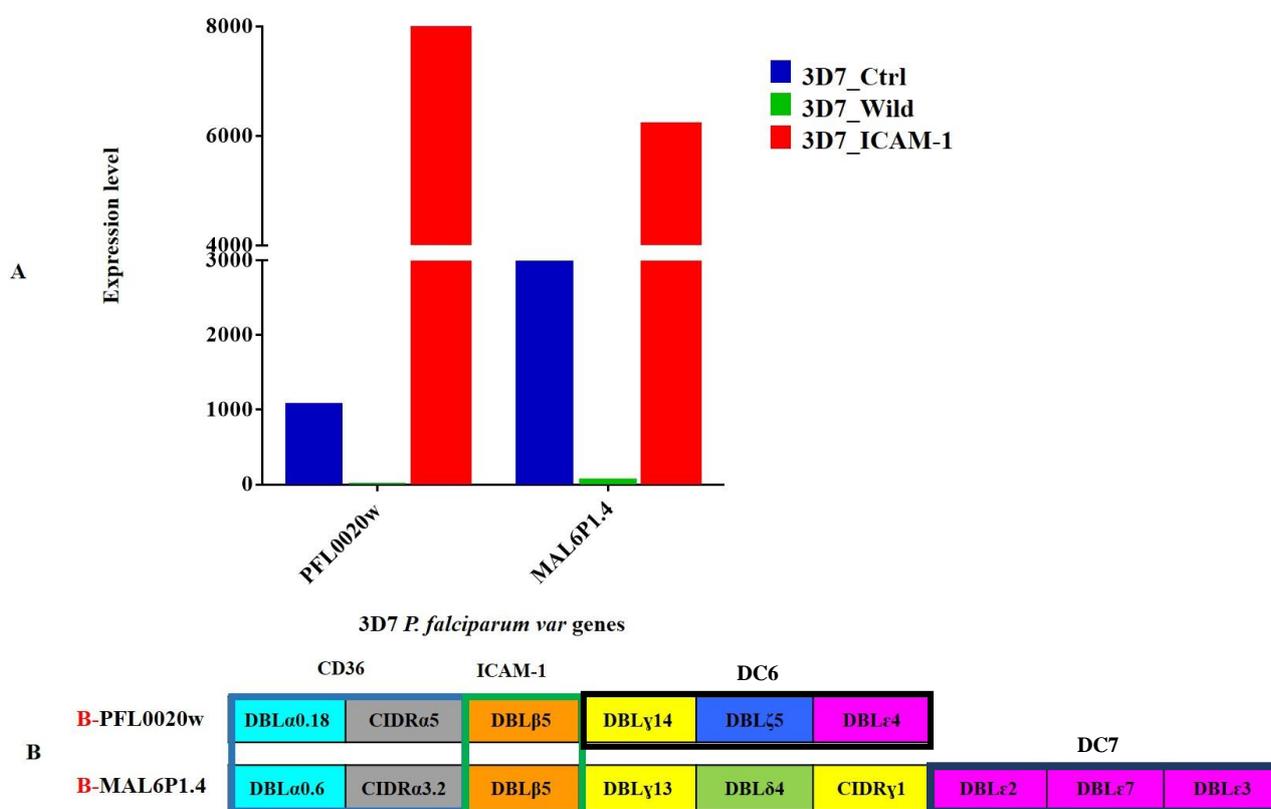


**Fig 4.11. var genes that were differentially expressed in IT4\_ICAM-1 IEs population.** (A) Differential gene expression of the four var genes that found to be highly expressed in IT4\_ICAM-1 culture compared to the IT4\_Ctrl and IT4\_Wild cultures. DEseq Bioconductor package was used for data analysis,  $\text{Padj} \leq 0.0001$ . P-value was corrected with Benjamini Hochberg method with FDR 10% (B) Schematic domain structure of *Pf*EMP1 proteins encoded by var genes IT4\_var01, IT4\_var16, IT4\_var41 and IT4\_var63 respectively. In the four variants binding domains for CD36 were observed (blue) as well as DBL $\beta$ 5 ICAM-1 binding domain (green) in the first three variants. Mapping of domain structure was done according to (Rask et al. 2010).

#### 4.2.6.1.2. Identification of 3D7 *P. falciparum* ligands mediating binding to ICAM-1.

Enrichment of 3D7 *P. falciparum* isolate took somehow longer time to be able to recover enough amount of parasites (5-7 weeks) which was expected due to the weak binding capacity showed by this isolate during the static binding assays (part 4.1.2). Differential expression analysis showed the two var genes PFL0020w and MAL6P1.4 were highly expressed in 3D7\_ICAM-1 parasite populations compared to both 3D7\_Ctrl and 3D7\_Wild (Fig. 4.12A). Interestingly, the protein domain structure encoded by these two var genes contains DBL $\beta$ 5 which is the binding domain for ICAM-1 (Fig. 4.11B) (Bengtsson et al. 2013, Smith et al. 2000).

Petter and colleagues identified PFL0020w *var* gene as the main expressed *var* gene after IEs selection for ICAM-1 binding (Petter et al. 2011). According to the recent DC classification, PFL0020w gene encodes for *PfEMP1* variant with DC6 (DBL $\gamma$ 14/ DBL $\zeta$ 5/ DBL $\epsilon$ 4) and MAL6P1.4 gene encodes for *PfEMP1* variant with DC7 (DBL $\epsilon$ 2/DBL $\epsilon$ 7/ DBL $\epsilon$ 3). Looking up in the whole transcriptome data, PF3D7\_0115400 *stevor* variant was highly expressed in 3D7\_ICAM-1 population compared to 3D7\_Wild and 3D7\_Ctrl populations (P<sub>adj</sub> <0.05). Other genes which was found differentially expressed in 3D7\_ICAM-1 population is attached in supp. table 15.



**Fig. 4.12. *var* genes that are highly expressed in 3D7\_ICAM-1 IEs population:** (A) Differential gene expression of *var* genes that found to be highly expressed in 3D7\_ICAM-1 (red) compared to the 3D7\_Ctrl (blue) and 3D7\_Wild cultures (green) IEs populations. DEseq Bioconductor package was used for data analysis, P<sub>adj</sub>  $\leq$  0.0001. P-value was corrected with Benjamini Hochberg method with FDR 10%.

(B) Schematic domain structure of *PfEMP1* proteins encoded by *var* genes **PFL0020w** and **MAL6P1.4** respectively. In the two variants binding domains for CD36 were observed (blue) as well as DBL $\beta$ 5 ICAM-1 binding domain (green). Mapping of domain structure was done according to (Rask et al. 2010).

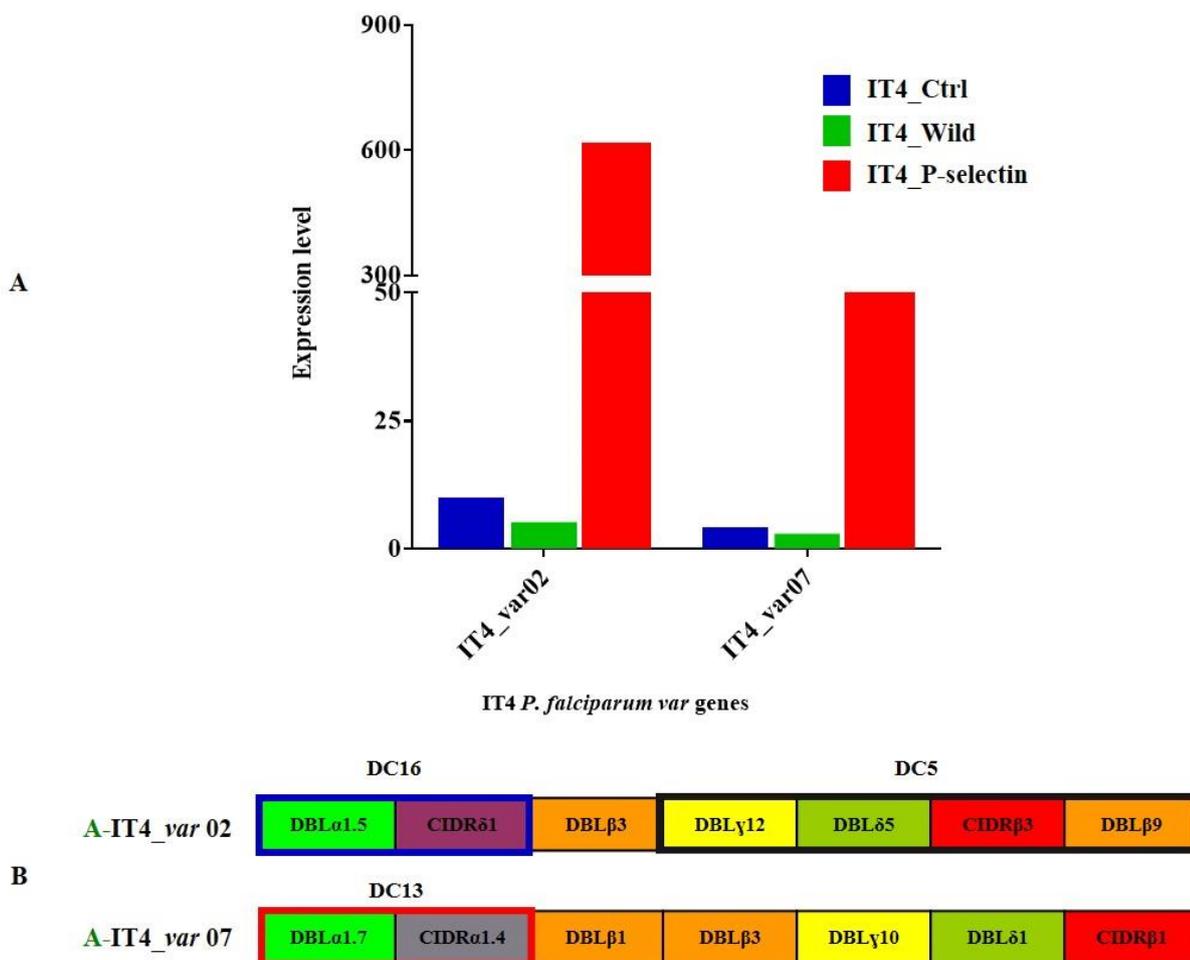
#### 4.2.6.2. P-selectin ligands identification

P-selectin receptor is expressed on surface of activated platelets and endothelial cells. It is described as a leukocyte adhesion receptor which plays a role in leukocyte rolling on the vessel wall and its accumulation at the site of inflammation (Bevilacqua and Nelson 1993, Mayadas et al. 1993). There are some evidences that suggest its role in pathogenesis of cerebral malaria for example P-selectin was up-regulated in brain vessels of cerebral malaria-susceptible mice but not of cerebral malaria resistant mice (Combes et al. 2004). Also in a recent study, it was shown to be able to interact with *PfMSP7* (*Plasmodium falciparum* merozoite surface protein 7) (Perrin et al. 2015), but until now the parasite ligands that interact with P-selectin are not yet identified. *P. falciparum* IEs were selected for P-selectin binding as mentioned above (part 4.2.3). Biological probes were harvested at early ring stage after the 5<sup>th</sup> enrichment round. RNA isolation and mRNA sequencing using NGS were performed as mentioned above (part 4.2.4). **Selection and enrichment over P-selectin was done by Dr. Ann-Kathrin Tilly and mentioned in her dissertation (2014). In this study her data were re-analysed using DESeq<sup>®</sup> Bioconductor package.** Data analysis was performed using DESeq<sup>®</sup> Bioconductor package to determine the abundance of the differentially expressed genes in the three populations, where a generalised lineal model (GLM) was used to fit the normalized values.

##### 4.2.6.2.1. Identification of IT4 *P. falciparum* ligands mediating binding to P-selectin

Looking up in the transcriptome profile of the IT4\_P-selectin IEs compared to the other populations IT4\_Ctrl and IT4\_Wild both group A *var* genes IT4\_var02 and It\_var07 were found to be differentially expressed (Fig. 4.13A). It was interesting to find IT4\_var02 as a candidate for binding to P-selectin. According to (Rask et al. 2010), it encodes a *PfEMP1* variant which contains the two domain cassettes DC16 (DBL $\alpha$ 1.5 and CIDR $\delta$ 1) as well as DC5 (DBL $\gamma$ 12 DBL $\delta$ 5 CIDR $\beta$ 3 DBL $\beta$ 9) (Fig. 4.13B). Another interesting candidate is IT4\_var07, its *PfEMP1* variant contains DC13 which could be associated with severe disease as well as other members of group A *var* gene (Lavstsen et al. 2012). These findings may support the assumption that P-selectin plays a role in CM and urge further studies to identify the interaction site between P-selectin and IEs. Searching the whole transcriptome of

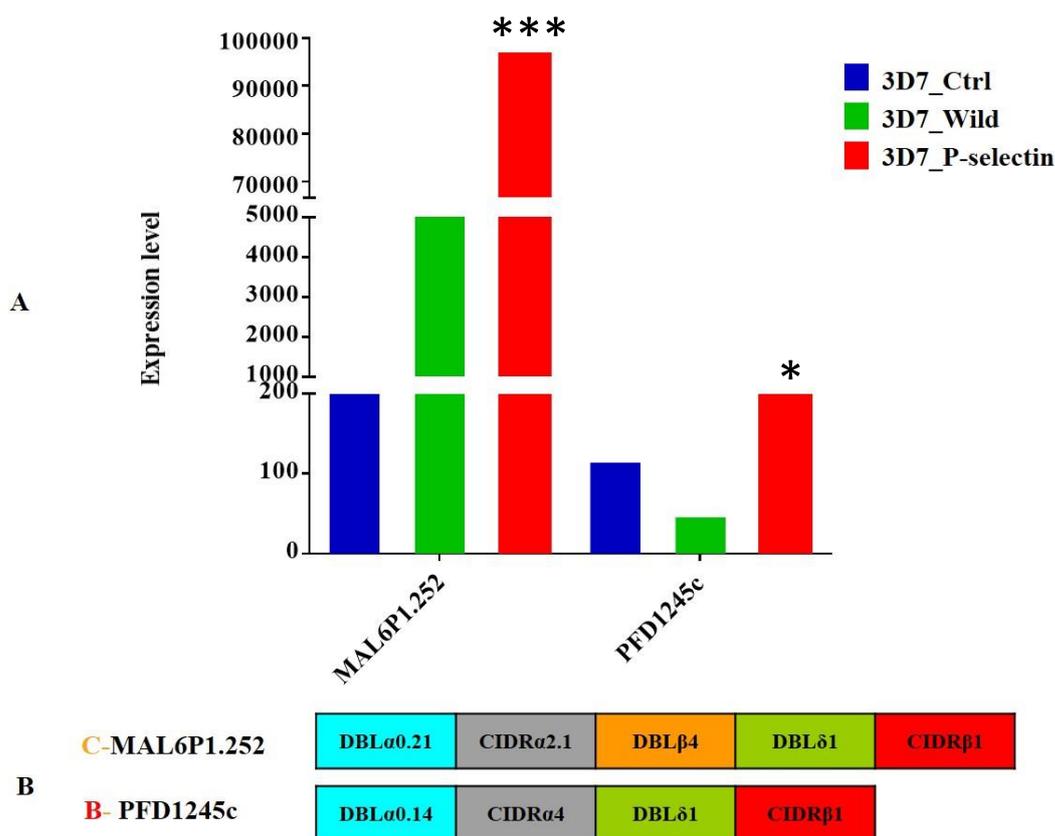
IT4\_P-selectin parasite population showed one *rif* variant to be differentially expressed compared to the other two populations (IT4\_Ctrl and IT4\_Wild) which is PFIT\_0801600 *rif* pseudogene (Padj <0.0001). On the other hand, neither *stevor* nor *Pfmc-2tm* genes were found to be highly expressed. For other genes which were found differentially expressed in the IT4\_P-selectin see supp. table 6 A/B.



**Fig. 4.13. var genes that were differentially expressed in IT4\_P-selectin IEs population.** (A) Differential gene expression of the four *var* genes that found to be highly expressed in IT4\_P-selectin (red) IEs population compared to the IT4\_Ctrl (blue) and IT4\_Wild ones (green); DEseq Bioconductor package was used for data analysis, Padj  $\leq$  0.0001. P-value was corrected with Benjamini Hochberg method with FDR 10%. (B) Schematic domain structures of *PfEMP1* proteins encoded IT4\_var02 gene, which contains domain cassette DC 16 (blue) and DC5 and IT4\_var07 gene, which contains DC13 (red). Mapping of domain structure was done according to (Rask et al. 2010).

#### 4.2.6.2.2. Identification of 3D7 *P. falciparum* ligands mediating binding to P-selectin

The transcriptome profile of 3D7\_P-selectin IEs showed two *var* genes differentially expressed compared to the other two populations (3D7\_Ctrl and 3D7\_Wild). Surprisingly one of them is MAL6P1.252 (Padj <0.0001) which was found to be highly expressed in the parasite population selected and enriched over CHO wild type (part 4.2.4.2). The other *var* variant highly expressed in this population was PFD1245c (Padj <0.05) (Fig. 4.14A). Schematic domain structures of both proteins encoded by these two *var* genes are shown in Fig. 4.14B. On the other hand, neither *rif*, *stevor* nor *PfMC-2TM* genes were found to be highly expressed. Interestingly, the gene PF3D7\_0202000 encoding KAHRP (Knob associated histidine rich protein) was found to be highly expressed in 3D7\_P-selectin IEs populations compared to 3D7\_Ctrl and 3D7\_Wild (Padj <0.05). Other genes which were found differentially expressed in 3D7\_P-selectin population were listed in supp. table 18.



**Fig. 4.14. *var* genes that are differentially expressed in 3D7\_P-selectin IEs population.** (A) Bars represents the expression level of *var* genes in 3D7\_P-selectin (red) 3D7\_Wild (green) and 3D7\_Ctrl (blue). Differential gene expression of **MAL6P1.252** and **PFD1245c** *var* genes; DEseq Bioconductor package was used for data analysis, \*\*\*Padj < 0.0001; \*Padj < 0.05. P-value was corrected with Benjamini Hochberg method with FDR 10% (B) Schematic domain structure of *PfEMP1* variants encoded by the two genes. Mapping of domain structure was done according to (Rask et al. 2010)

#### 4.2.6.3. E-selectin ligands identification

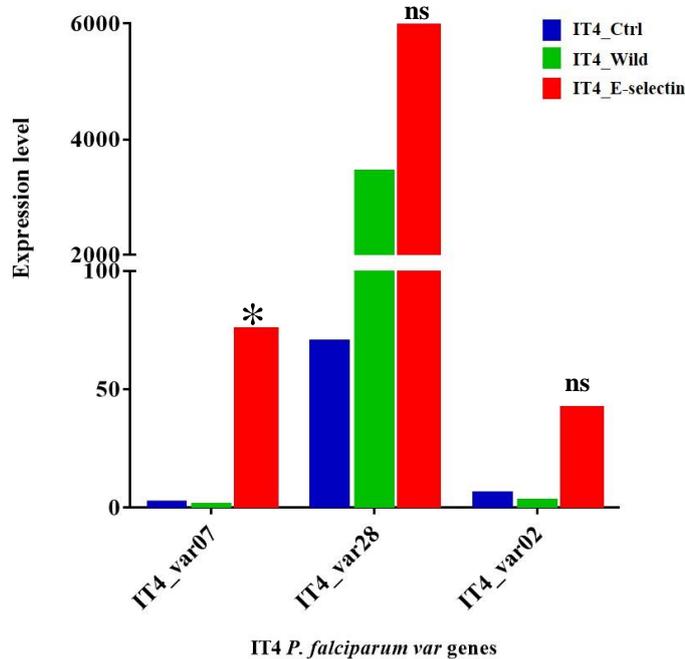
E-selectin is an endothelial cell receptor involved in the initial binding and rolling of leukocytes at the site of inflammation. It is expressed on the endothelial surface in response to cytokines (e.g. TNF $\alpha$  and IL-1 $\beta$ ) (Preston et al. 2016). The role of E-selectin receptor in development of CM is not yet clear. There were evidences of co-localisation of IEs with expression of E-selectin in post-mortem pathological samples from children with CM (Armah et al. 2005). On the other hand, some authors believe that E-selectin has no role in IEs cytoadhesion (Rowe et al. 2009) based on the following facts; some studies failed to detect any tethering or rolling of IEs over E-selectin under flow conditions and other observed very low binding capacity of IEs from African patients to E-selectin (Newbold et al. 1997, Udonsangpetch et al. 1997, Udonsangpetch et al. 1996). In this study, *P. falciparum* IEs from the two isolates 3D7 and IT4 were selected and enriched for binding to E-selectin using CHO cells permanently express E-selectin on the surface as a GFP fusion protein. All the steps were done as mentioned in parts 4.2.3 and 4.2.4.

##### 4.2.6.3.1. Identification of IT4 *P. falciparum* ligands mediating binding to E-selectin

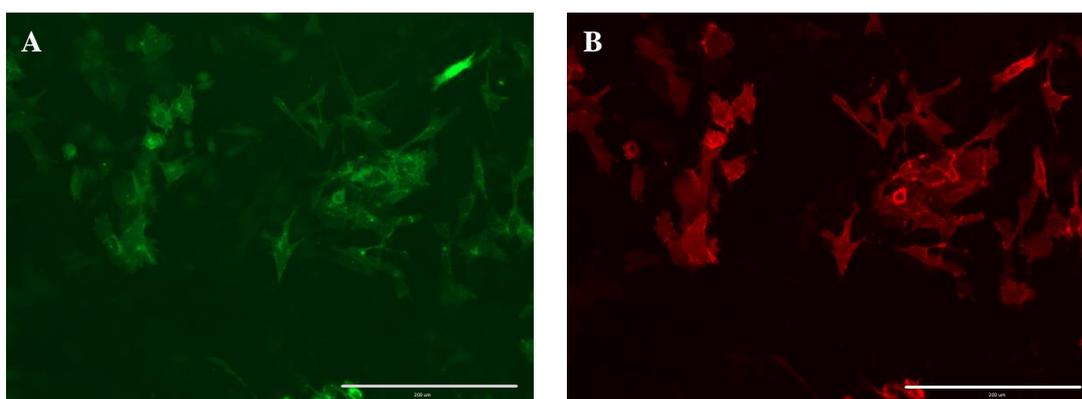
Starting the selection and enrichment of IT4 *P. falciparum* IEs for binding to E-selectin receptor, two experiments were performed; the first one was performed during the establishment phase where **no preabsorbtion step was performed**. Four different biological samples were isolated upon reaching at least 8% ring stage parasitemia. Data analysis showed that IT4\_var07 was differentially expressed among IT4\_E-selectin IEs populations (P<sub>adj</sub> <0.05). On the other hand, IT4\_var28 showed high expression in IT4\_E-selectin IEs population compared to IT4\_Wild but the difference was not statistically significant. Besides, IT4\_var02 showed some tendency to be highly expressed among IT4\_E-selectin IEs population but also no statistical significance could be detected (Fig. 4.15). Surprisingly, the genes IT4\_var32A, IT4\_var32B and IT4\_var14 exhibited approximately the same expression level between the two enriched IEs populations IT4\_E-selectin and IT4\_Wild but on the other hand, the expression level was higher than that of IT4\_Ctrl (Table 2). Adding to this, the two *var* genes IT4\_var35 and IT4\_var34 showed the same expression level in the three IEs populations.

Looking up in the whole transcriptome data one *rif* pseudogene (PFIT\_0801600) showed tendency to be highly expressed in IT4\_E-selectin IEs population but with no statistical significance. On the other hand, neither *stevor* nor *PfMC-2TM* genes were found to be highly expressed. Other genes which were found differentially expressed in IT4\_E-selectin IEs population are attached, in see supp. table 7/8. A second experiment for E-selectin enrichment was performed. First, a new cell line of CHO cells expressing E-selectin was produced. Surface expression of E-selectin was confirmed using immunofluorescence microscopy test and FACS sorting of the new cell line was performed regularly to assure the selection of the cells expressing E-selectin on the surface (Fig. 4.16) (for more details Part 2.2.1.2.5.) In this experiment, the pre-absorption step mentioned in part 4.2.3 was performed. Three different biological probes were harvested followed by NGS and data analysis as mentioned in part 4.2.4.

Data analysis showed significant decrease in the expression level of IT4\_var28 gene in IT4\_E-selectin IEs population compared to IT4\_Wild ( $P_{adj} < 0.05$ ) (Table 2). Adding to this, no differential expression of both IT4\_var07 and IT4\_var02 was identified. Furthermore, IT4\_var32A, IT4\_var32B and IT4\_var14 exhibited approximately the same expression level between the two enriched IEs populations IT4\_E-selectin and IT4\_Wild but on the other hand, the expression level was higher than that of to IT4\_Ctrl (Table 2). Besides, the two *var* genes IT4\_var35 and IT4\_var34 showed the approximately same expression level in the three IEs populations. Looking up in the whole transcriptome data, no other genes were observed to be differentially expressed within IT4\_E-selectin IEs populations.



**Fig. 4.15. var genes that showed high expression level in IT4\_E-selectin IEs population.** Bars represents the expression level of var genes in 3D7\_E-selectin (red) 3D7\_Wild (green) and 3D7\_Ctrl (blue). Differential gene expression of IT4\_var07 compared to the IT4\_Ctrl and IT4\_Wild cultures; \* $P_{adj} < 0.05$ . ns: not significant. DEseq Bioconductor package was used for data analysis. P-value was corrected with Benjamini Hochberg method with FDR 10%



**Fig. 4.16. CHO cells expressing E-selectin receptor on the surface) (A)** CHO-745 cells were transfected using Lipofectamine 2000 (Invitrogen) complexed with pAcGFP1-N1. Cells were harvested and subjected to FACS sorting using a BD FACSAria cell sorter. For immunofluorescence cells were grown on coverslips and fixed with 4% para-formaldehyde. **(B)** Surface-exposed E-selectin was labelled using the  $\alpha$  E-selectin antibody (R&D systems) and secondary antibodies conjugated to Alexa-Fluor-594 (Invitrogen). Secondary antibody did not show any reaction without the first antibody.

**Table 2. GLM mathematical model used for data analysis**

First experiment										
var genes	Mean expression level			X-interception	CHO true	E-selectin true	deviance	convergence	P value	Padj
	IT4_Ctrl	IT4_Wild	IT4_E-selectin							
IT4_var28	71.04921	3479.025	7910.482	6.143701	5.619524	1.186348	10.17727	TRUE	0.008941	0.599033
IT4_var32A	6.008675	3151.228	4551.621	2.640556	8.9795	0.532122	6.724565	TRUE	0.42991	1
IT4_var32B	18.47751	3835.369	4961.883	4.210022	7.693022	0.37364	7.271392	TRUE	0.518199	1
IT4_var29	12.65205	495.7137	709.7534	3.659442	5.29029	0.52152	5.937752	TRUE	0.515597	1
IT4_var14	39.15491	613.7492	674.2963	5.290938	3.957338	0.149198	7.403633	TRUE	0.803949	1
IT4_var35	2361.988	2110	1636.98	11.20524	-0.16252	-0.36623	6.504241	TRUE	0.342218	1
IT4_var34	2956.787	2761.262	2148.607	11.52936	-0.09598	-0.36449	7.364115	TRUE	0.31925	1
Second experiment										
var genes	Mean expression level			X-interception	CHO true	E-selectin true	deviance	convergence	P value	Padj
	IT4_Ctrl	IT4_Wild	IT4_E-selectin							
IT4_var28	46.12724	2214.793	168.5305	5.525433	5.587158	-3.98938	8.24197	TRUE	0.000304	0.020336
IT4_var32A	3.846081	2003.932	4164.628	1.950432	9.018002	1.053328	2.468141	TRUE	0.639546	1
IT4_var32B	11.91337	2440.779	5643.691	3.574581	7.678388	1.20794	2.365707	TRUE	0.609793	1
IT4_var29	8.016961	314.7946	340.1166	3.002833	5.294378	0.068586	3.505123	TRUE	0.967906	1
IT4_var14	25.36391	390.9292	575.9386	4.664534	3.944515	0.538365	3.090636	TRUE	0.773302	1
IT4_var35	1499.84	1350.057	1773.567	10.54994	-0.15194	0.414113	5.407202	TRUE	0.262127	1
IT4_var34	1880.163	1765.718	1591.772	10.87629	-0.08851	-0.10539	8.077642	TRUE	0.806597	1

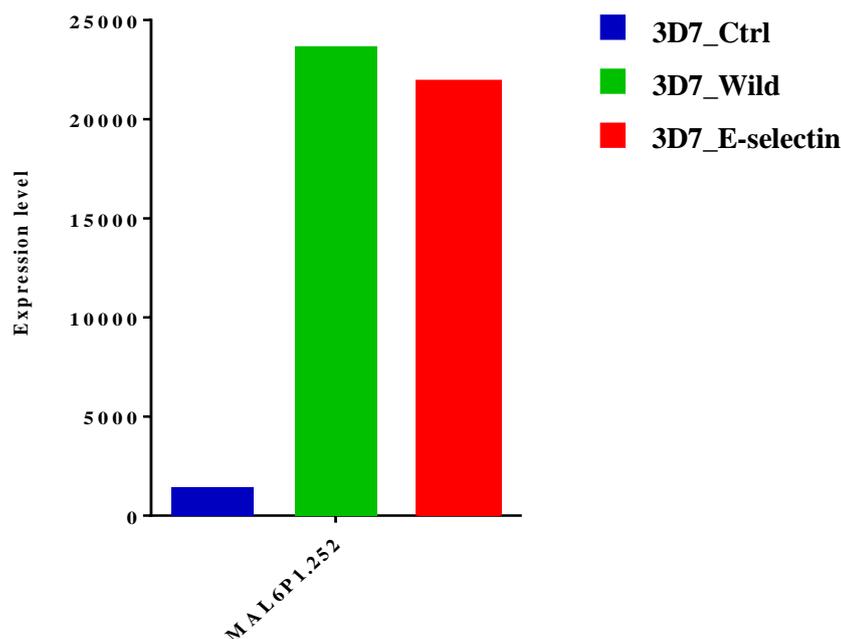
Here some *var* genes are listed which are important for the aim of discussion of the enrichment process during the two experiments. The most significant observation is, in the first experiment, the expression level of IT4\_var28 in IT4\_E-selectin IEs population was higher compared to its level in IT4\_Wild IEs population. Whereas in the second experiment its expression level was significantly decreased (Padj<0.05). DEseq Bioconductor package was used for data analysis. P-value was corrected with Benjamini Hochberg method with FDR 10%.

#### 4.2.6.3.2. Identification of 3D7 *P. falciparum* ligands mediating binding to E-selectin

Initial selection and enrichment of 3D7 *P. falciparum* IEs over E-selectin receptor was done using the newly transfected CHO-E-selectin cell line mentioned in part 4.2.6.3.1. Generally, it was observed that the enrichment using 3D7 IEs needed more rounds than in case of IT4. Four biological samples were isolated for RNA isolation and NGS sequencing. Sequencing was done using Illumina HiSeq 4000 platform (BGI Hongkong). The four samples were multiplexed in two lanes so that each sample was represented with two technical samples for assurance of the results. Combination of the raw reads between the two lanes was performed. Data were analysed using DESeq<sup>®</sup> Bioconductor package to determine the abundance of the differentially expressed genes in the three populations were a generalised lineal model (GLM) was used to fit the normalized values.

Data analysis showed that MAL6P1.252 *var* gene was the only gene that is highly expressed among 3D7\_E-selectin as well as 3D7\_Wild compared to 3D7\_Ctrl IEs populations showing mean expression levels of 1448.291, 23681.89, and 21899.83 respectively. These data showed no statistical significance according to the model used in the analysis but it may also mean that the selection and enrichment were only over the CHO cells and not over the E-selectin receptor (Fig. 4.17).

Analysis of the data using DEseq 2 version also showed no difference in the *var* genes expression profile. In addition, the whole transcriptome profile of 3D7\_E-selectin IEs population showed no differential gene expression compared to the other two populations see supp. table 19/20.



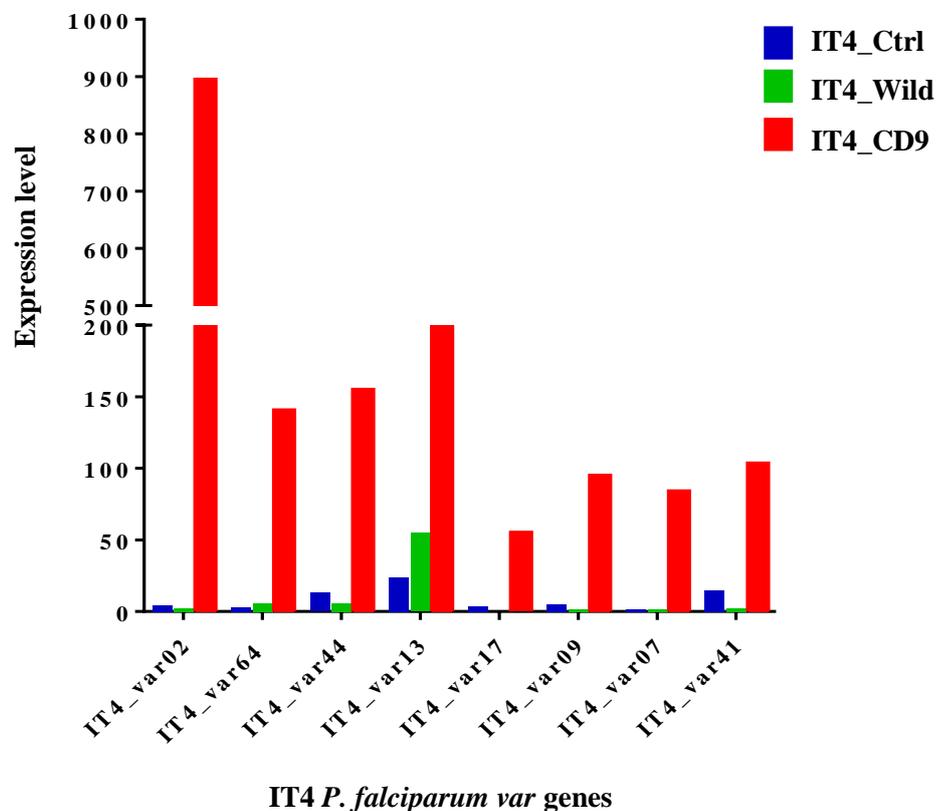
**Fig. 4.17. MAL6P1.252 var gene that was highly expressed in 3D7\_E-selectin and 3D7\_Wild IEs populations.** Bars represents the expression level of MAL6P1.252 var gene in 3D7\_E-selectin (red) 3D7\_Wild (green) and 3D7\_Ctrl (blue). Data showed only difference when compared to 3D7\_Ctrl IEs population but no statistical significance in the GLM model used. DEseq Bioconductor package was used for data analysis. P-value was corrected with Benjamini Hochberg method with FDR 10%.

#### 4.2.6.4. CD9 ligands identification

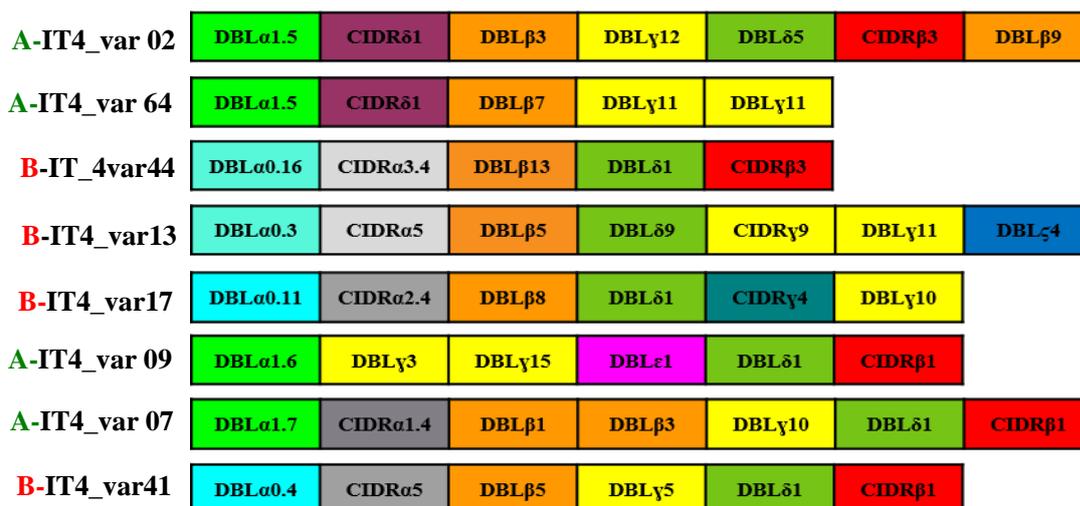
CD9 receptor is a member of transmembrane 4 superfamily known as tetraspanins. Tetraspanins are found in all cell types but not in the erythrocytes (Charrin et al. 2001). CD9 recently is known to play a role in reproduction; it was proved that CD9 deficient mice suffer female infertility due to impaired gamete fusion. One tetraspanin member (CD81) is known to be used by *P. falciparum* parasite to enter the hepatocyte (Chambrion and Le Naour 2010). CD9 is recently described as a receptor that might be used by *P. falciparum* in the cytoadhesion interaction but until now the parasite ligands for this interaction is not yet known (Esser et al. 2014). In this study, CD9 was expressed on CHO cells as a fusion protein and *P. falciparum* IEs from the two isolates 3D7 and IT4 were selected and enriched for binding to this receptor. All the steps were done as mentioned in parts 4.2.3 and 4.2.4 (with the help of Dr. Ann-Kathrin Tilly).

#### 4.2.6.4.1. Identification of IT4 *P. falciparum* ligands mediating binding to CD9

Surprisingly, in IT4\_CD9 IEs population 8 *var* genes were differentially expressed compared to the other two IEs populations (IT4\_Ctrl and IT4\_Wild);  $P_{adj} < 0.0001$  (Fig. 4.17/18). Interesting fact that four of them belong to group A UPS *var* genes and they are IT4\_var02, IT4\_var64, IT4\_var09 and IT4\_var07. The other four *var* genes belong to group B UPS *var* genes IT4\_var44, IT4\_var13, IT4\_var17 and IT4\_var41. Searching for other variant surface antigens in the whole transcriptome profile, only one *rif* gene was differentially expressed (PFIT\_0801600) in IT4\_CD9 IEs population.



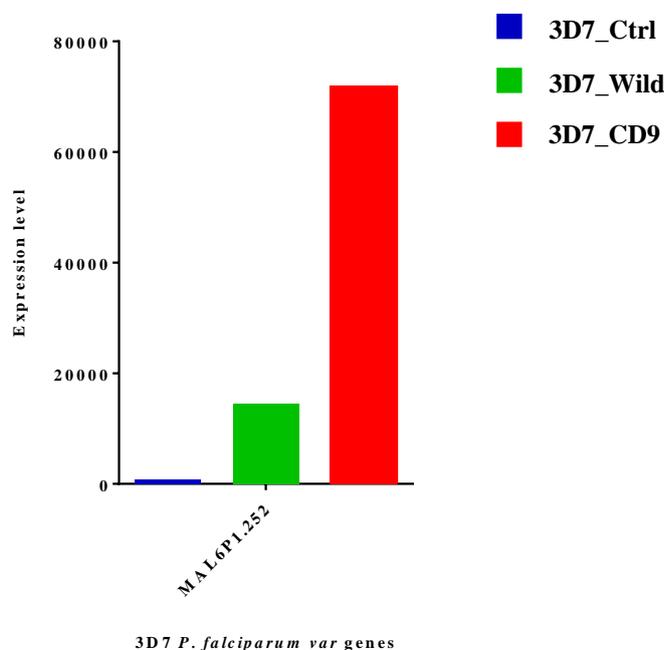
**Fig. 4.18. *var* genes that were differentially expressed in IT4\_CD9 IEs population.** Bars represents the expression level of *var* genes; IT4\_CD9 (red) IT4\_Wild (green) and IT4\_Ctrl (blue); DEseq Bioconductor package was used for data analysis,  $P_{adj} < 0.0001$ . P-value was corrected with Benjamini Hochberg method with FDR 10%.



**Fig. 4.19. Schematic domain structure of *PfEMP1* variants encoded by the *var* genes.** Four of them belong to group A UPS *var* genes and they are IT4\_var02, IT4\_var64, IT4\_var09 and IT4\_var07 Mapping of domain structure was done according to (Rask et al. 2010).

#### 4.2.6.4.2. Identification of 3D7 *P. falciparum* ligands mediating binding to CD9

The transcriptome profile of 3D7\_CD9 IEs showed one *var* genes differentially expressed compared to the other two populations (3D7\_Ctrl and 3D7\_Wild). Surprisingly, this *var* gene was MAL6P1.252 (Padj <0.0001) (Fig. 4.20). This gene was found to be highly expressed in the parasite population selected and enriched over CHO wild type (part 4.2.4.2).



**Fig. 4.20. MAL6P1.252 *var* gene that was highly expressed in 3D7\_CD9 IEs populations.** Bars represents the expression level of MAL6P1.252 *var* gene in 3D7\_CD9 (red) 3D7\_Wild (green) and 3D7\_Ctrl (blue) DEseq Bioconductor package was used for data analysis, Padj <0.0001. P-value was corrected with Benjamini Hochberg method with FDR 10%.

#### 4.2.6.5. CD151 ligands identification

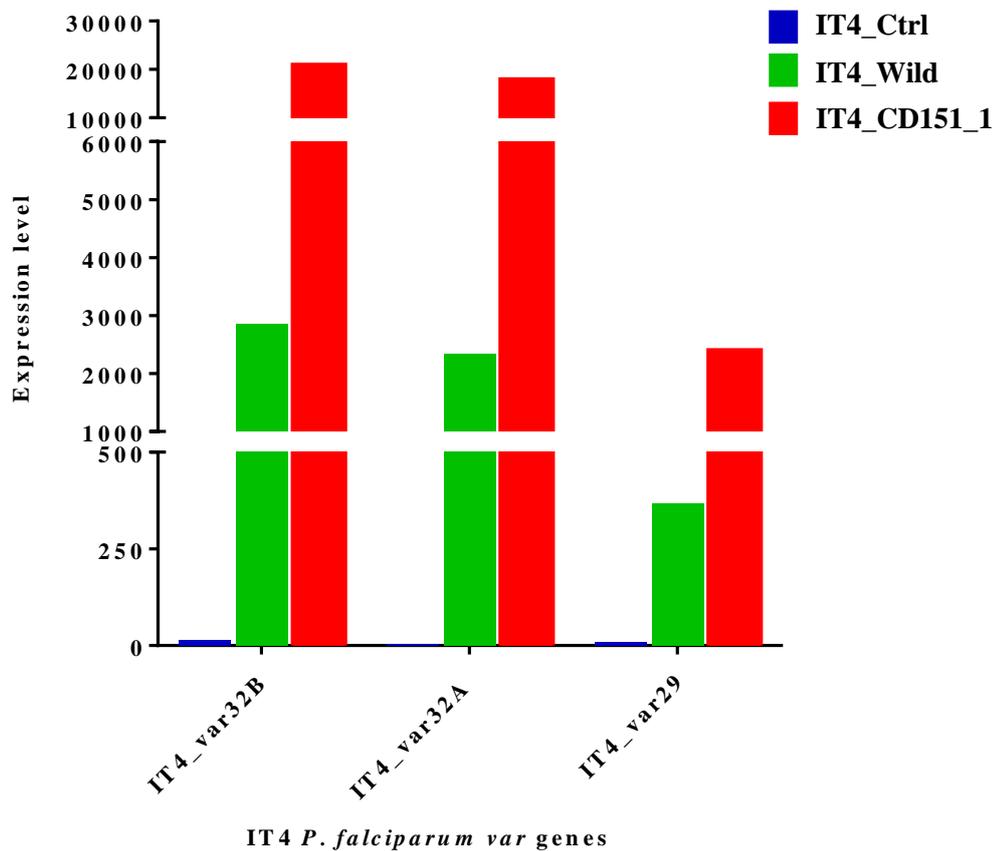
A member of tetraspanin family. It is composed of four transmembrane domains, two extracellular loops, and two short cytoplasmic tails. CD151 is known to be involved in endocytosis of HPV16 (van Sriel and Figdor 2010). It was first described as a receptor for *P. falciparum* by Esser and colleagues where they tested the binding to CD151 using a pool of patient isolates and the laboratory isolate IT4 (Esser et al. 2014). In this study the two isolates IT4 and 3D7 were enriched over CD151 in order to identify the probable ligands for this interaction.

##### 4.2.6.5.1. Identification of IT4 *P. falciparum* ligands mediating binding to CD151

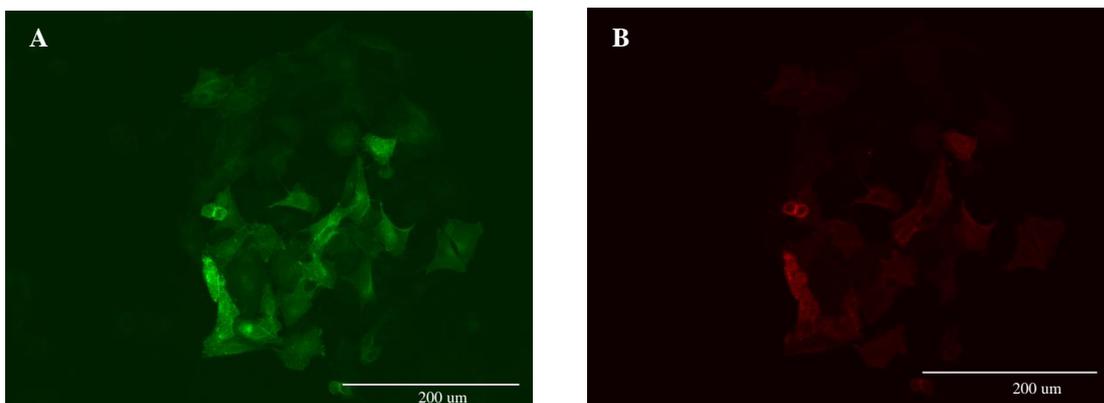
Two experiments were performed; the first one was performed during the establishment phase, **without the preabsorbtion step**. Four different biological samples were isolated upon reaching at least 8% ring stage parasitemia. Library preparation as well as NGS run were performed as described above (part 4.2.4). Data analysis showed that, three *var* genes were highly expressed in IT4\_CD151 IEs populations (Fig. 4.21). These were IT4\_var32B, IT4var32A and IT4var29. Part of the whole transcriptome data of this experiment is attached in Supp. table 9.

A second experiment for CD151 enrichment was performed. CHO cell lines were tested for the surface expression of CD151 using immunofluorescence microscopy test and FACS sorting of the CHO transfectant was performed regularly to assure the selection of the cells expressing CD151 on the surface (Fig. 4.22). In this experiment, the pre-absorption step mentioned in part 4.2.3 was performed. Five different biological probes were harvested followed by NGS and data analysis as mentioned in part 4.2.4.

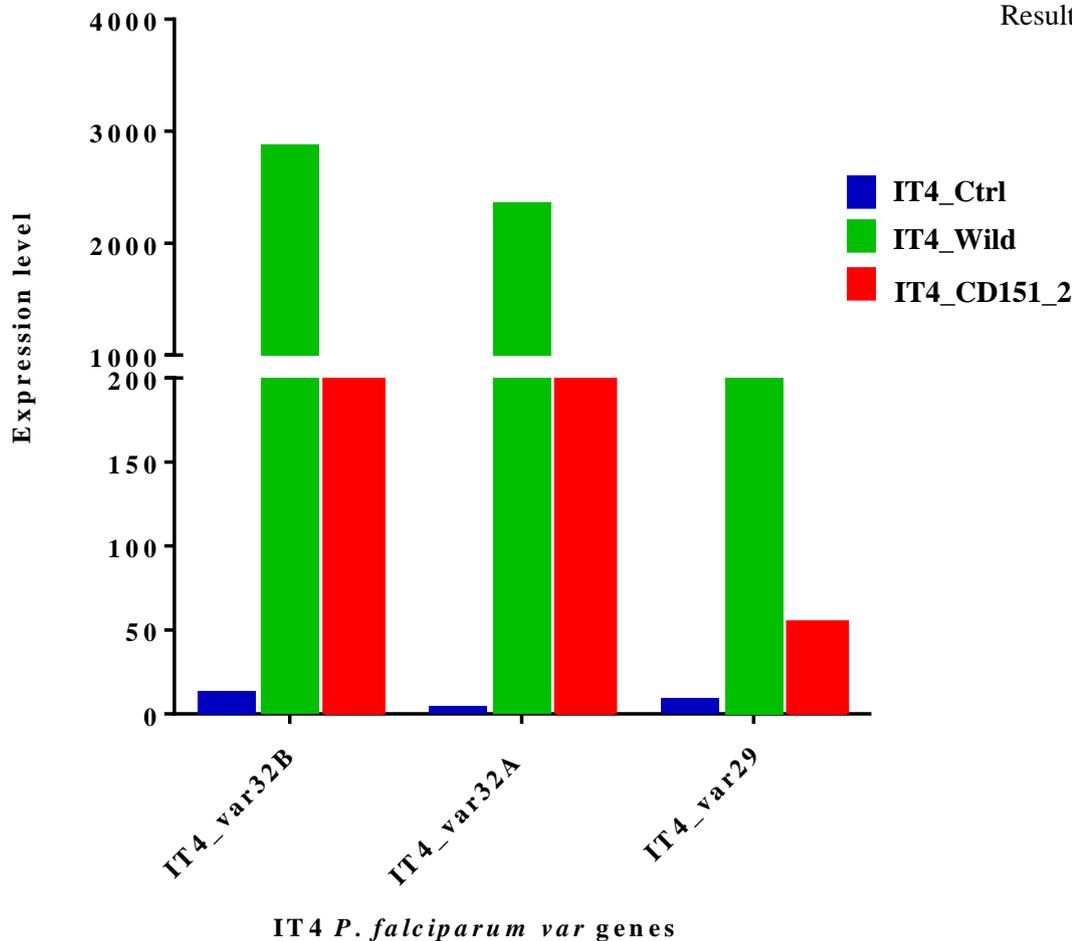
Data analysis showed that, IT4\_var32B, IT4\_var32A and IT4\_var29 -that were highly expressed in the first experiment- had a lower expression level in the second experiment (Fig. 4.23) which gave an indicator for the effect of the preabsorbtion step and proved that these IEs populations were only binding to the unknown receptor/structure over CHO wild type cells. For more details on the transcriptome, see Supp. table 10.



**Fig. 4.21. var genes that were highly expressed in IT4\_CD151 IEs population.** Bars represents the expression level of var genes; IT4\_CD151 (red) IT4\_Wild (green) and IT4\_Ctrl (blue); DEseq Bioconductor package was used for data analysis. P value was corrected with Benjamini Hochberg method with FDR 10%.



**Fig 4.22. CHO cells expressing CD151 receptor on the surface) (A)** CHO-745 cells were transfected using Lipofectamine 2000 (Invitrogen) complexed with pEGFP1-N1. Cells were harvested and subjected to FACS sorting using a BD FACSAria cell sorter. For immunofluorescence assay, cells were grown on coverslips and fixed with 4% para-formaldehyde. **(B)** Surface-exposed CD151 was labelled using homemade  $\alpha$  CD151 monoclonal antibody and secondary antibody conjugated to Alexa-Fluor-594 (Invitrogen).



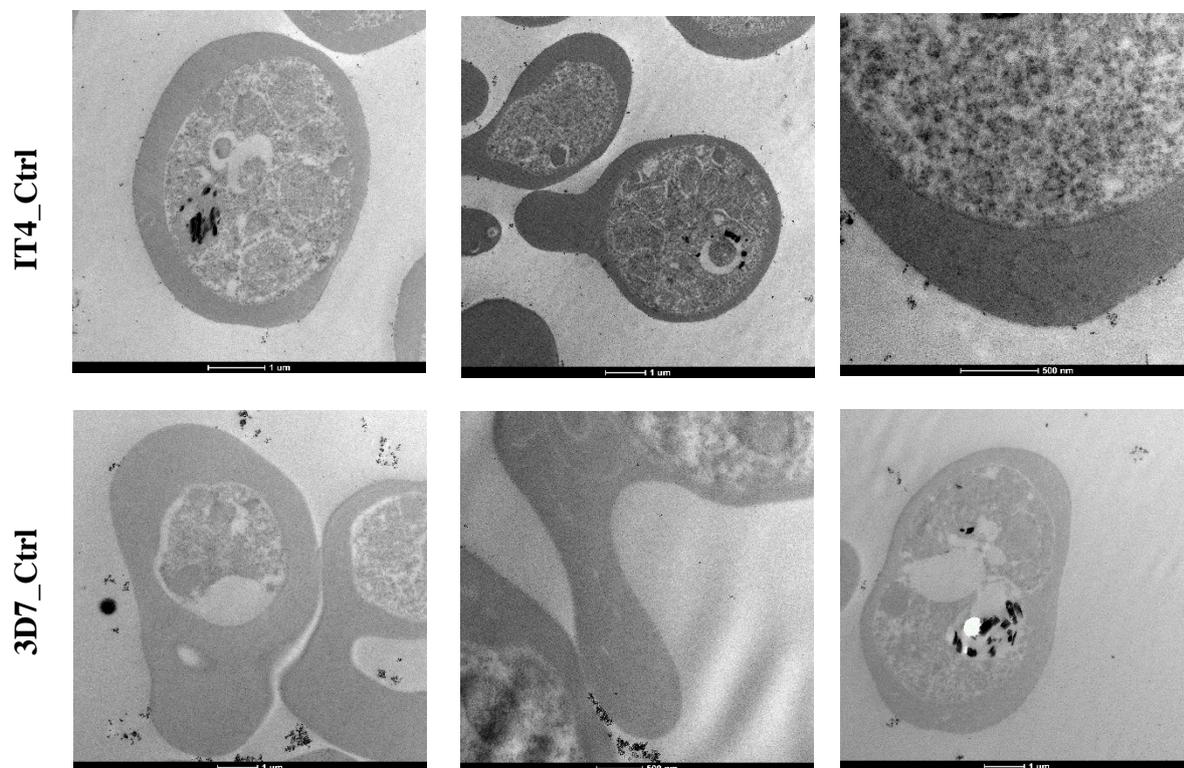
**Fig. 4.23. Expression level of IT4\_var32B/32A/29 var genes in the second experiment.** Bars represent the expression level of var genes in the three populations; IT4\_CD151 (red) IT4\_Wild (green) and IT4\_Ctrl (blue); DESeq Bioconductor package was used for data analysis. P-value was corrected with Benjamini Hochberg method with FDR 10%.

#### 4.2.6.5.2. Identification of 3D7 *P. falciparum* ligands mediating binding to CD151

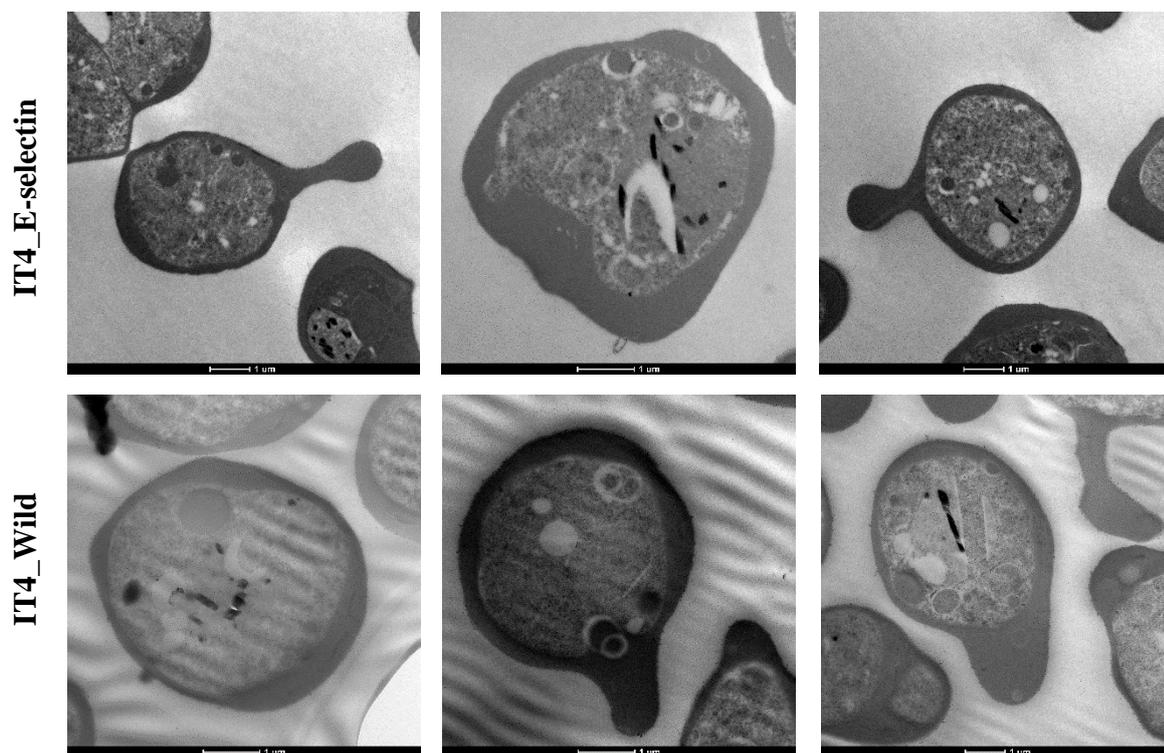
Initial selection and enrichment of 3D7 *P. falciparum* IEs over CD151 was done as described before. Four biological samples were isolated for RNA isolation and NGS sequencing. Sequencing was done using Illumina HiSeq 4000 platform (BGI Hongkong). The four samples were multiplexed in two lanes. For data analysis, the raw reads between the two lanes were combined. Data were analysed using DESeq<sup>®</sup> Bioconductor package. MAL6P1.252 var gene was the only gene that is highly expressed among 3D7\_CD151 as well as 3D7\_Wild compared to 3D7\_Ctrl IEs populations. These data showed no statistical significance according to the model used in the analysis but it may also mean that the selection and enrichment were only over the CHO cells and not over CD151. The whole transcriptome profile of 3D7\_CD151 IEs population showed no differential gene expression compared to the other two populations, see supp. table 23.

### 4.3. Knob characterization of different *P. falciparum* isolates using TEM

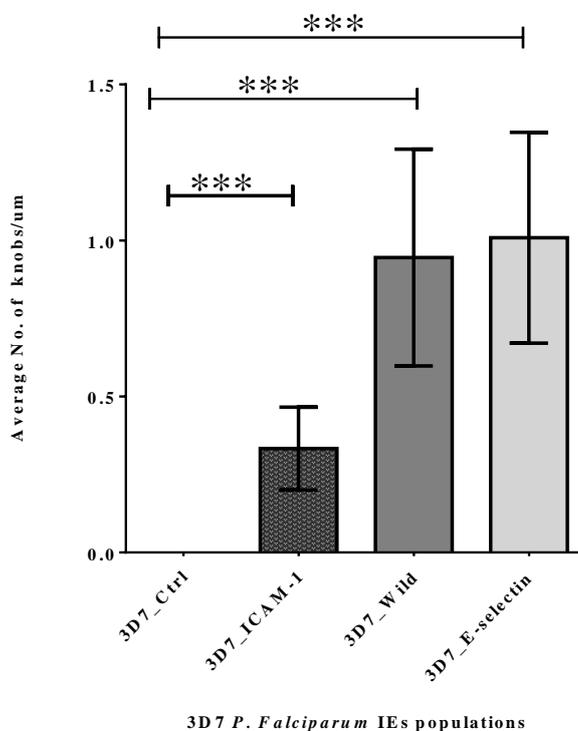
In order to be able to identify the presence and amount of knobs on the surface of the trophozoite stage IEs, percoll gradient was used to separate trophozoite stages followed by washing in PBS puffer then the pellet was chemically fixed using 2.5% GA and 2.5% PFA. For staining, 1% osmium tetroxide (OsO<sub>4</sub>) and 0.5% Uranyl acetate (UA) were used. First, the Ctrl cultures from both populations (IT4 and 3D7) showed absence of knobs (Fig. 4.24). Surprisingly IT4\_Wild and IT4\_E-selectin populations showed also a smooth IEs surface without knobs (Fig. 4.25). On the other hand, 3D7 selected IEs populations showed knobs on the surface. Higher knob density was observed in both 3D7\_Wild and 3D7\_E-selectin compared to 3D7\_ICAM-1. The knob densities were as follows, in case of 3D7\_ICAM-1 0.33 knobs / 1 $\mu$ m  $\pm$  0.13 SD whereas 3D7\_Wild showed 0.94 knobs / 1 $\mu$ m  $\pm$  0.34 SD and finally 3D7\_E-selectin exhibited 1.01 knobs / 1 $\mu$ m  $\pm$  0.33 SD (Fig. 4.26/4.27).



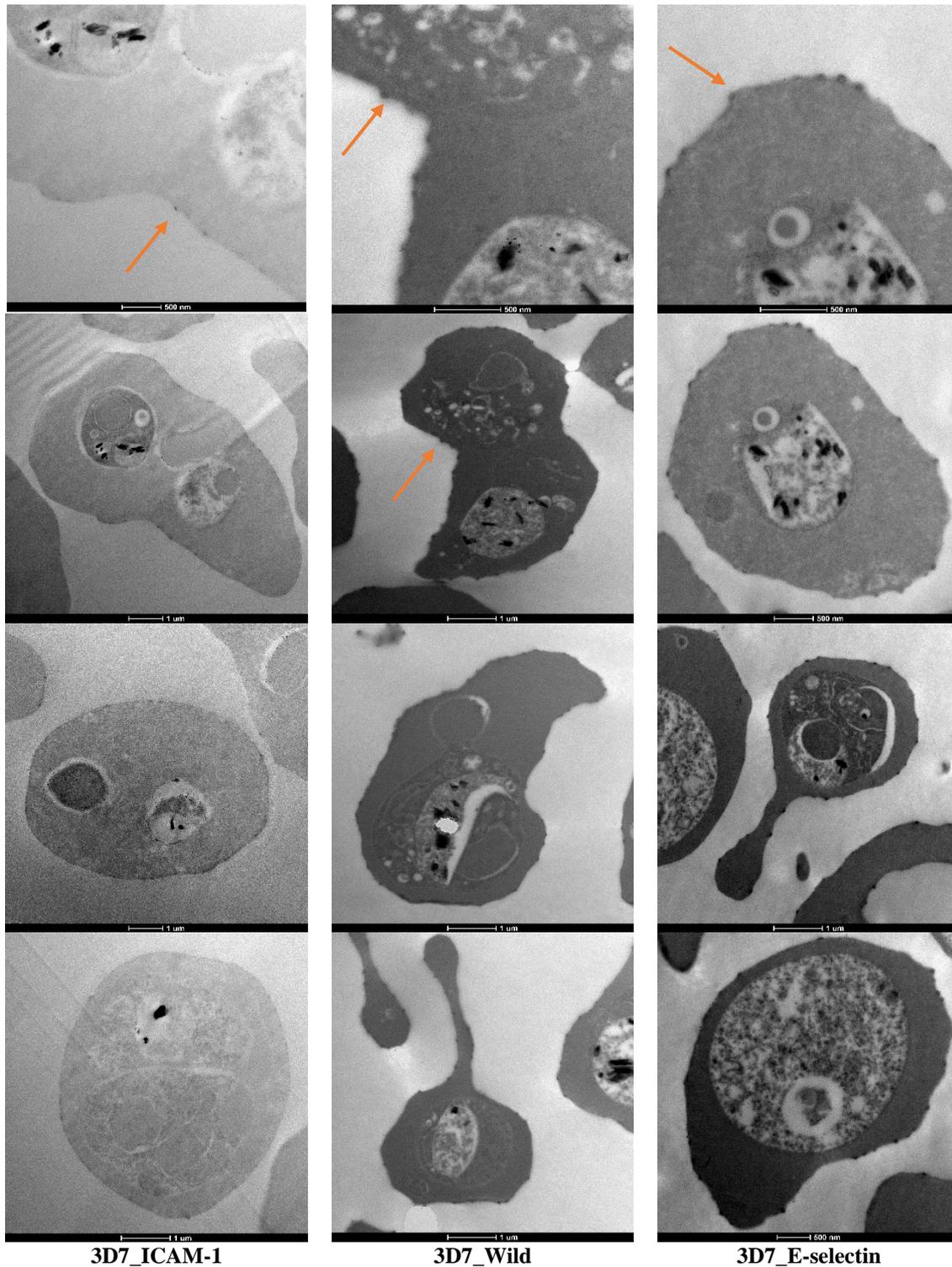
**Fig. 4.24. Transmission electron microscopy of trophozoite stage of IT4\_Ctrl and 3D7 Ctrl IEs.** Both isolates did not show any knobs on the surface. For staining 1% OsO<sub>4</sub> and 0.5% Uranylacetat UA were used. Samples were sectioned in 65 nm thick sections using a Leica EM UC7. Sections were examined with a Tecnai Spirit transmission electron microscope at 80 kV (FEI). 30 IEs were counted to estimate the knob density.



**Fig. 4.25. Transmission electron microscopy of trophozoite stage of IT4 IEs receptor selected populations.** Both selected populations (IT4\_Wild and IT4\_E-selectin) did not show any knobs on the surface. For staining 1% OsO<sub>4</sub> and 0.5% Uranylacetat UA were used. Samples were sectioned in 65 nm thick sections using a Leica EM UC7. Sections were examined with a Tecnai Spirit transmission electron microscope at 80 kV (FEI). 30 IEs were counted to estimate the knob density.



**Fig. 4.26. Transmission electron microscopy of trophozoite stage of 3D7 IEs receptor selected populations.** Bars represent the average number of knobs per 1 μm. In each population 30 different cells were analysed for the presence of knobs. ImageJ software was used to determine the circumference of the IEs in the cut section, then the total number of knobs were counted in each IEs and the number was divided by the circumference in μm. \*\*\* significance was approved using Mann-Whitney test.



**Fig. 4.27. Transmission electron microscopy of trophozoite stage of 3D7 IEs receptor selected populations.** Higher knob density was observed in both IEs populations 3D7\_Wild and 3D7\_E-selectin compared to 3D7\_ICAM-1. (Arrows point to knobs). For staining 1% OsO<sub>4</sub> and 0.5% Uranylacetat UA were used. Samples were sectioned in 65 nm thick sections using a Leica EM UC7. Sections were examined with a Tecnai Spirit transmission electron microscope at 80 kV (FEI). 30 IEs were counted to estimate the knob density.

## 5. Discussion

Decades of research and malaria is still a big burden for human. About 70% of deaths caused by malaria infection occurred in children under five years of age according to WHO report in 2015. *P. falciparum* is an intense form of malaria infection in human; this is attributed to the severe complications that might occur during the course of infection; principally, cerebral malaria (CM). Aiming to survive inside its host, *P. falciparum* parasite distinctively modify the physiological characters as well as the antigenic properties of the IEs; accordingly, avoid the splenic clearance and evade the host immunity. In the last decade, sequestration of the IEs in the vascular bed of the body organs especially the brain has been a central point in field of malaria research. Studying the molecular mechanisms behind this sequestration will be of great help to take the first step towards the development of a new vaccine that could block the adhesion thus the spleen will be able to remove the unsequestered parasites and circumvent any further complications.

### 5.1. The starting point

The story behind this research issue started in Italy in 1892, where the two malariologists -Marchiafava and Bignami- reported a thoroughly description to *P. falciparum* infection after autopsies of patients died from this infection. They started to describe all the blood stages of *P. falciparum*, with the observation that only the ring stages appear in the peripheral circulation whereas the mature forms are collected in the small vessels of large organs mainly in the following order, the brain, lungs, spleen, bone marrow, liver and intestine. They reported also the appearance of gametocytes in the spleen and bone marrow. Describing the picture of the cause of CM at this time took several years of research to accept this theory (White et al. 2013). Through their assumption, the doors were opened for unlimited questions in field of malaria research which until now are not completely answered due to the complexity of the mechanisms used by this parasite.

## 5.2. Unanswered questions behind the interaction between *P. falciparum* IEs and different endothelial receptors

In literature, the parasite's surface exposed protein; *PfEMP1* was identified to interact with many endothelial receptors mediating *P. falciparum* IEs sequestration. For some of these receptors, their parasite's ligand is identified and the others not yet. The interactions with CD36 and ICAM-1 with the IEs were studied in details but until now there was no straightforward association between these two receptors and occurrence of severe complications, that's why the question of the significance of anti-adhesion therapies against the binding domains to these two receptors is still not answered (Madkhali et al. 2014, Rowe et al. 2009, Turner et al. 1994). Recently, Turner and colleagues (Turner et al. 2013) highlighted EPCR as a receptor impinged in pathogenesis of severe malaria, but another study revealed another role of this receptor in CM (Moxon et al. 2013).

Still lots of assumptions in this puzzling problem, Esser and colleagues (Esser et al. 2014) decided to open more and more questions by revealing that seven new receptors may be accused of playing a role in CM pathogenesis but *Plasmodium* ligands still unidentified. By this, a total number of 22 endothelial receptors still under investigations.

## 5.3. Binding capacity of *P. falciparum* IEs to different endothelial receptors

In this study, the binding capacity of the two laboratory isolates 3D7 and IT4 was tested using static binding (assays see part 4.1.2). Principally, the two isolates showed high binding capacity to CD36 and that is logically in agreement with the protein structure of *PfEMP1* variants expressed by these two isolates, where it could be observed that in both isolates most of *PfEMP1* variants (42 variants in IT4 and 47 variants in 3D7) contain CIDR $\alpha$ 2-6 in a well-known head structure known for binding to CD36 (Robinson et al. 2003, Smith 2014, Smith et al. 2000). The role of CD36 in pathogenesis of malaria is controversial despite of the high binding phenotype to CD36. Due to the lack in data that correlate between severe malaria and CD36 binding phenotype, it may be postulated that the interaction with CD36 may provide the survival interactions which facilitate the parasite's reproduction within the host (Cabrera et al. 2014, Serghides et al. 2003). In his study -my colleague Pedro Lubiana- (unpublished data), observed that under flow conditions the IEs roll over CD36 which may support the postulation that the interaction

between CD36 and IEs meant to slow down the IEs through rolling until they firmly adhere to another receptor; that's why it is important for most of the IEs to exhibit the binding domains for this interaction.

The binding assays showed very weak binding capacity of both isolates IT4 and 3D7 to the other endothelial receptors studied here (ICAM-1, P-selectin, E-selectin, CD9 and CD151). The binding to ICAM-1 showed different pattern between the two isolates; where IT4 IEs showed stronger affinity (40 IEs/100 CHO cells) than those of 3D7 (3 IEs/100 CHO cells). Whereas, the binding capacity to the other four receptors in IT4 was as weak as in 3D7 ranging from 3-6 IEs/100 CHO cells. Esser and colleagues measured the binding capacity of the IT4 isolate and a pool of field isolates to the two tetraspanins CD9 and CD151 (Esser et al. 2014). Their results showed almost the same, considering the weak binding capacity of the IT4 isolate to CD9 and especially CD151. On the other hand, they reported stronger binding capacity using field isolates. In a previous study, both isolates showed also weak binding capacity to the selectins as well as to the tetraspanins (Tilly et al. 2015).

#### **5.4. Selection and enrichment over CHO cells “the unknown receptor”**

##### **5.4.1. The history of CHO cells**

CHO cell lines are considered unique in their culture characteristics; they were established in 1957 in the laboratory of Dr. Theodore T. Puck from 0.1 gram of ovary tissue of a Chinese hamster where they developed some kind of spontaneous immortalization and appeared as a fibroblast type mostly with a diploid karyotype (Puck 1958, Puck et al. 1958, Wurm 2013). This special character was not observed in primary cells of human origin; where the fully diploid karyotype goes through senescence (Hayflick limit) followed by death of the cells after about 50 population doubling (Hayflick and Moorhead 1961, Puck 1958, Puck et al. 1958, Wurm 2013). In cytoadhesion studies of *P. falciparum*, two CHO cell lines were adopted as an *in vitro* model for these experiments; the first one is CHO-K1 which is used to study the cytoadhesive phenotype related to placental malaria due to the presence of chondroitin-4-sulfate on its surface. The second cell line, which was **used in this study** is CHO-745 ATCC, a glycosaminoglycan deficient CHO-K1 (Esko et al. 1985). Both cell lines don't express any known *P. falciparum* receptor on the surface (such as CD36, ICAM-1, thrombospondin, VCAM-1, E-selectin or CD31)

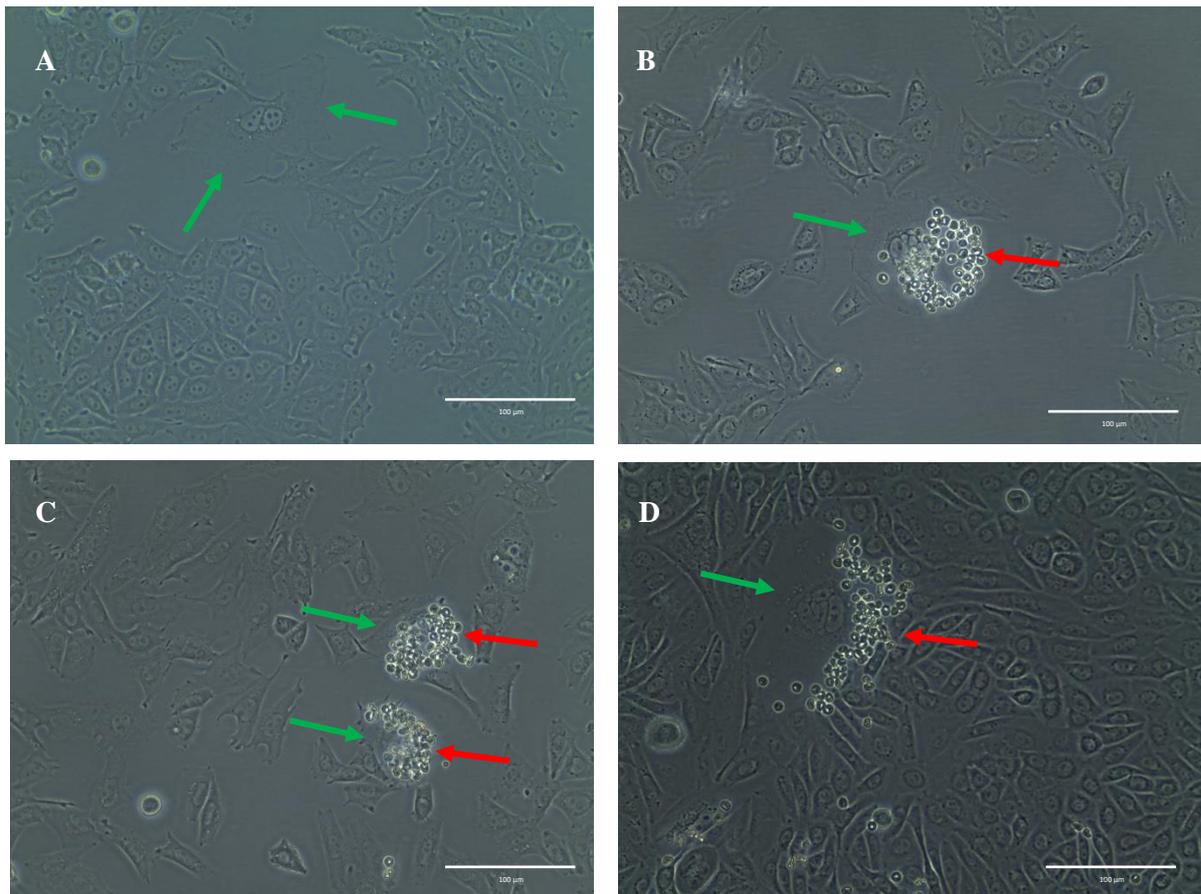
(Andrews et al. 2005, Pouvelle et al. 2000). Therefore, transgenic CHO-745 ATCC overexpressing the receptor of interest can be used to facilitate the study of other binding phenotypes than the placental one. In another studies as well as in this study, binding of IEs to wild-type CHO cells was observed and consequently it was suggested that these cell lines may harbour a new unknown *P. falciparum* receptor (Andrews et al. 2005, Andrews et al. 2003).

#### **5.4.2. Interesting binding phenotype**

During the enrichment process in this study particular forms of IEs collections were regularly observed binding over the monolayer of CHO cells. Trying to figure out the nature of this binding phenotype the following were evidenced; during selection over CHO wild type cells, collections of IEs observed together in a clump-like form adhering to a single morphologically abnormal CHO cell (Fig. 1B/C/D). These kind of cells appear repetitively throughout the cultivated monolayer not only in the wild type culture but also within the transfected cell lines. These cells were larger in size than the other CHO cells with multiple nuclei (Fig. 5.1A). In literature, cells which lose their original morphology known as ``senescent cells``. It was described that these cells are larger in size with many vacuoles in the cytoplasm, containing a larger nucleus or are multinucleated with an increase in the number of lysosomes and Golgi. Also these cells known to lose their ability to multiply, they still metabolically active (Campisi and d'Adda di Fagagna 2007, Hayflick and Moorhead 1961, Robbins et al. 1970).

Now, the question is, on which structure do the IEs bind on the surface of these cells?

The senescence of endothelial cells was investigated by Mun and his colleagues (Mun and Boo 2010), where they postulated that there might be strong adhesive molecules present on the surface of these cells. In their study, they found that CD44 gene was higher expressed in senescent cells than in younger ones. High binding capacity of monocytes -in a cluster form- to senescent endothelial cells in comparison the younger ones was also reported in another study (Lowe and Raj 2014). CD44 is a cell surface glycoprotein which is known to be involved in different cell to cell interactions like lymphocyte rolling and monocyte migration across the blood-brain barrier (Ponta et al. 2003).



**Fig. 5.1. Special binding phenotype to CHO senescent cells** (A) Red arrow points to morphologically abnormal binucleated CHO cells. (B), (C) and (D) demonstrate different examples of this binding phenotype in three independent experiments. Green arrow points to IT4 *P. falciparum* IEs clump-like collection of cells.

In order to investigate the expression of CD44 on the surface of CHO cells used in this study, the cells were stained using  $\alpha$ CD44 antibodies and secondary antibody conjugated to Alexa-fluor 594. Cells were then examined by fluorescence microscopy confirming the expression of CD44 on the surface of CHO cells used in this study, see Supp. fig. 1. It was not possible to correlate the amount of fluorescence to the senescent cells than the others; it was equally the same all over the cells. That's why further methods need to be found to investigate whether CD44 acts as a receptor for the IEs or not, and if not then to which structure on the senescent cells IEs were able to bind?

### 5.5. Absence of mutually exclusive *var* gene expression phenomena in ring stage IEs of IT4 *P. falciparum* isolate

According to previous studies, it is important for the parasite to limit the exposure of the surface antigens to keep its own ammo; evading the immune response and survive within the host. Therefore, only one *var* gene is expressed and consequently only one specific *PfEMP1* is exposed on the surface of mature trophozoite stage IEs (Chen et al. 1998, Rowe et al. 2009, Scherf et al. 1998). It was reported in some studies that in few *in vitro* cultivated isolates multiple *var* genes are transcribed in ring stage IEs but only one *var* gene is detected in the mature stages (Noviyanti et al. 2001, Scherf et al. 1998).

An interesting observation in our data analysis is the highly and equally expression pattern of the two *var* genes IT4\_var35 (*var1* variant, pseudogene) and IT4\_var34 in all IT4 populations investigated. This means that the pattern of mutually exclusive expression of *var* genes in ring stage IEs does not apply at least in the IT4 isolate. IT4\_var35 or *var1* an exclusive *var* pseudogene, is often found as a truncated gene according to Rask and colleagues (Rask et al. 2010). They found that in IT4 isolate *var1* lacks the exon2 sequence. Adding to this they found that both *var1* and *var2CSA* have shortened ATS which might be puzzling about the function of *var1*. Some studies suggest that the function of pseudogenes is to act as an archive for *var* diversity like in case of VSG of *Trypanosoma* (Barry et al. 2007). Kyes and colleagues postulated that, the conserved *var1* variant falls outside the normal mutually exclusive expression of *var* genes. They summarized their findings that in ring stage IEs, one dominant *var* transcript is highly expressed which will be translated to surface as a *PfEMP1* variant in the trophozoit stage, whereas, in the trophozoit stage two *var* transcripts exists; the dominant *var* gene and *var1* variant (Kyes et al. 2007).

Another interesting finding was that, in the whole transcriptome analysis of IT4 isolate used in this study, the PFIT\_bin09700 gene was also highly equally expressed, BLAST analysis indicated that the mRNA sequence identified is a fragment of IT4\_var34; it seems that this gene exists as two fragments in IT4 isolate used in this study which might affect its translation to a surface *PfEMP1*.

It is important to mention that in 3D7 isolate used in this study, *var1* gene (PF3D7\_0533100 or PFE1640w) was not found to be highly expressed in ring stage IEs as in case of IT4. This finding was also mentioned by Taylor and colleagues (Taylor et al. 2000). They reported that this isolate behaves differently where *var1* pseudogene transcript detected in pigmented trophozoite stages.

## **5.6. *var* gene variants that were differentially expressed in IT4 IEs (IT4\_Wild) and 3D7 IEs (3D7\_Wild) enriched over wild-type CHO cells**

### **5.6.1. DBL $\delta$ 1- CIDR $\beta$ 1 domain combination**

Data analysis showed that five *var* genes were differentially expressed in IT4\_Wild IEs population. According to the last classification done by Rask and colleagues, three of them belong to group B *var* genes (IT4\_var32B, IT4\_var29, IT4\_var14), one belongs to group C *var* genes (IT4\_var28) and one its UPS group is undetermined (IT4\_var32A) (Rask et al. 2010). The *Pf*EMP1 variants encoded by these genes showed that, four variants contain the classical head structure that binds to CD36 (DBL $\alpha$ 0-CIDR $\alpha$ 2-6) (Baruch et al. 1996, Robinson et al. 2003, Smith et al 2013), Kraemer and Smith 2003, Petter and Duffy 2015, Rask et al. 2010). The highly significant three variants IT4\_var28/32A/32B contain in common DBL $\delta$ 1 domain in combination with CIDR $\beta$ 1/5 domains. The other two variants IT4\_var14/29 contain in common DBL $\delta$ 1.

In 3D7\_Wild only one *var* variant was differentially expressed; MAL6P1.252 which belongs to group C *var* genes. The *Pf*EMP1 encoded by this gene showed the classical head structure that binds to CD36 receptor (DBL $\alpha$ 0-CIDR $\alpha$ 2). It is interesting to mention that, this protein also contains the combination of DBL $\delta$ 1 and CIDR $\beta$ 1 domains found in IT4\_Wild populations. Looking at the whole protein structure of *Pf*EMP1 variants found in IT4 and 3D7 isolates, it could be concluded that the above mentioned combination is frequently repeated. It is found in 27 variants in both isolates (Supp. fig. 3/4) which may bring the question about the function of these two domains. It is important also to mention that this combination also present frequently in other isolates studied by Rask and colleagues (for example 13 times in DD2, 15 in HB3 and 19 in PFCLIN) (Rask et al. 2010). This frequent repeated manner may give the impression that these domains might be important for the surveillance of the parasite through binding to a specific receptor (unknown until now) like in case of CD36 binding domains and their frequent

repetition in different variants. This question may open the doors for further investigations for the binding affinity of these two domains to different endothelial receptors which may be a helpful clue to know more about the cytoadhesive capacity of *P. falciparum* IEs.

### 5.6.2. IT4\_var32B

An interesting *var* candidate was found to be differentially expressed in IT4\_Wild IEs populations is also IT4\_var32B. This protein contains the domain cassette DC8 which composed of the domains DBL $\alpha$ 2, CIDR $\alpha$ 1.1, DBL $\beta$ 12 and DBL $\gamma$ 6 (Rask et al. 2010).

In a recent study Lavstsen and colleagues compared the *var* gene transcriptome in children with different malaria syndromes and found that children presented with severe malaria symptoms were infected with parasites expressing *var* genes which encode *PfEMP1* variants containing DC8 (Lavstsen et al. 2012). These data were also supported by another study in Benin (Bertin et al. 2013). In addition, binding of *PfEMP1* variants containing DC8 to brain, heart, lung and bone marrow endothelial cells was found to be through interaction with EPCR receptor and parasite's domain CIDR $\alpha$ 1 (Turner et al. 2013). In Malawian, children who died of severe malaria the EPCR was not found in their brain microvasculature, which may be explained there might be another receptor that interact with another domain harboured by the *PfEMP1* variants (Moxon et al. 2013).

Another interesting finding in this study, was the absence of knobs in all IT4 enriched populations including those which could bind to IT4\_Wild and expressing high levels of IT4\_var32B among their *var* transcriptome. This seemed to be inconsistent with a recent study conducted by Subramani and colleagues. They were aiming to identify the link between the knob density of IEs and the *PfEMP1* variant expressed on the surface. So they selected a parasite line expressing IT4\_var32B variant by incubating IT4 IEs with human monoclonal antibodies against this protein. Then they used atomic force microscopy to compare the knob density. They found that the parasite population expressing IT4\_var32B on the surface showed increased knob density compared to those expressing IT4\_var04 or IT4\_var60 (Subramani et al. 2015). It was suggested that knob density is linked to the *PfEMP1* variant expressed. But this was not the case here in this study, where high binding capacity was identified in absence of knobs (IT4\_Wild, IT4\_E-selectin) (Fig. 4.25).

### Outstanding questions from the above mentioned data in this section:

- Which receptor/structure is present on the surface of CHO cells that might perform the interaction with the IEs? Or does CD44 act as a receptor as suggested above?
- What is the function of DBL $\delta$ 1 and CIDR $\beta$ 1 domains, or in other words to which receptor do they act as ligands?
- Does the knob density play a role in the capacity of IEs binding to CHO cells in this study?

### 5.7. ICAM-1 ligands (proof of concept)

The proof of concept used in this study was selecting the trophozoite stage IEs of the two isolates IT4 and 3D7 over ICAM-1; which is a well described receptor known to interact with *P. falciparum* IEs.

#### 5.7.1. DBL $\beta$ 5 domains

IT4\_ICAM-1 IEs population showed differential gene expression of four *var* gene variants (IT4\_var01/var16/var41/var63). *PfEMP1* domains encoded by these *var* genes contain the head structure required for binding to CD36 (DBL $\alpha$ 0-CIDR $\alpha$ 2-6). In addition, three of them (IT4\_var01/var16/var41) interestingly exhibit a **DBL $\beta$ 5** domain in their protein structure. These three *var* variants were also identified in the study of Bengtsson and colleagues, where they identified the ICAM-1 binding and non-binding DBL $\beta$  domains. From 16 DBL $\beta$  ICAM-1 binding domains 7 were DBL $\beta$ 5 and the rest were DBL $\beta$ 3 belong to DC4 (Bengtsson et al. 2013). On the other hand, IT4\_var63 *PfEMP1* variant contain the DBL $\delta$ 1-CIDR $\beta$ 1 domains discussed above which may be enriched due to unknown binding interaction with the CHO cells. Adding to this, 3D7\_ICAM-1 IEs populations showed a differential expression of PFL0020w and MAL6P1.4 from group B *var* genes. Both genes contain also **DBL $\beta$ 5** as well as CD36 binding domains in their protein structure. PFL0020w candidate was also identified by Petter and colleagues upon selection of 3D7 isolate for binding to ICAM-1 (Petter et al. 2011). Despite the identification of the *PfEMP1* domains interacting with ICAM-1 but until now, no crystal structure for a DBL $\beta$ \_ICAM-1 binding domain exists. Some studies found that in ICAM-1, the binding site of DBL $\beta$  domains is the D1 domain. Whereas, DBL $\beta$  domain

interact with ICAM-1 using its C-terminal end (Chattopadhyay et al. 2004, Lennartz et al. 2015, Mayor et al. 2005, Smith et al. 2000).

### 5.7.2. Efficacy of our enrichment system

Identifying the above mentioned candidates as ICAM-1 binders' proofs the potency of using the enrichment method used in this study. Although there might be some disadvantages like the unknown receptor on the surface of CHO cells (discussed in part 5.3), we overcame this problem through two steps. Firstly, performing the preabsorbtion step by incubating the IEs over the CHO wild type cells prior to the enrichment step. Secondly, identifying the transcriptome of IEs that could be enriched on CHO wild type cells and according to results -see table 1 part 4.2.6.1.1- all the *var* genes that were differentially expressed in IEs\_Wild populations, were not highly expressed in case of IEs\_ICAM-1 populations. Adding to this, we took the advantage of using this system which the natural folding of the proteins that might be required for the *Pf*EMP1\_receptor interactions.

### 5.7.3. Role of ICAM-1 binding in CM

The evidence of ICAM-1 involvement in CM is not yet conclusive inspite of many studies. Knowing that ICAM-1 is a receptor that is upregulated due to inflammation and endothelial cell activation and this means it needs an initiatory mechanism to play a role in IEs sequestration. The most important evidence of its involvement in CM was an observed colocalization of IEs as well as ICAM-1 expression in a brain sample of patient died with CM (Turner et al. 1994). Tripathi and colleagues found that exposure of human brain endothelial cells to IEs was accompanied by high expression of ICAM-1 (Tripathi et al. 2006). Adding to this the high binding capacity to ICAM-1 by patient isolates from Africa with severe malaria compared to patients with uncomplicated and mild malaria (Turner et al. 2013). On the other hand, patient isolates from Asia showed low affinity to bind to ICAM-1 (Ockenhouse et al. 1991, Udomsangpetch et al. 1996). It is also important to take a look on the expermental cerebral malaria studies with animal models to be able to see the whole picture. Studies showed that, brain vessels from mice with CM showed a marked upregulation of ICAM-1 level (Falanga and Butcher 1991, Lou et al. 2001). In addition ICAM-1 deficient mice showed resistance to CM; they survived longer than the wild types (Favre et al. 1999, Shikani et al. 2012). Another

interesting study on the murine cerebral malaria showed that, there was a significant upregulation of ICAM-1 upon immunohistochemical analysis of the inner ear in mice with hearing loss during the course of severe CM (Schmutzhard et al. 2011).

## 5.8. P-selectin ligands

### 5.8.1. Does P-selectin plays a role in pathogenesis of CM?

Chang and his colleagues tried to answer this question using animal model in experimental malaria, where they found that P-selectin deficient mice infected with *P. berghei* experience few pathologic changes compared to controls with the same parasitemia level. Adding to this they observed upregulation of P-selectin in the endothelial lining of several organs; brain, lungs and kidneys (Chang et al. 2003). Another study revealed actually the same results, in which they reported that, P-selectin immunostaining in brain vessels of *P. berghei* infected mice showed upregulation of P-selectin in brain vessels of CM-susceptible mice in contrast to CM resistant mice. Mice with P-selectin knocked out gene did not show any sign of CM or any neurological complications. They also ruled out the role of platelet P-selectin by using platelet P-selectin deficient mice, which showed signs of CM similar to that seen in wildtype mice (Combes et al. 2004).

Considering *P. falciparum*, the role of P-selectin is not yet clear; it was described to serve for IEs rolling in field isolates from Thailand (Ho et al. 1998, Udomsangpetch et al. 1997). Yipp and colleagues reported that P-selectin is able to capture the rapidly moving IEs; due to its unique extension beyond the glycocalyx, thus facilitating the IEs binding to other endothelial receptors (Yipp et al. 2007). It was reported by MacDonald and his colleagues that IEs produces histamine releasing factor found in sera of patients with *P. falciparum* infection which stimulate the release of histamine from human leukocytes which in turn cause upregulation of P-selectin most probably to help in the cytoadhesion process (MacDonald et al. 2001). An elevated level of soluble P-selectin in serum of patients with severe malaria compared to those of non-severe symptoms was also reported, confirming the role of P-selectin in severe malaria pathogenesis (Facer and Theodoridou 1994).

A recent study suggested P-selectin plays another role in severe malaria other than its adhesive role. Where they found that, *PfMSP7* interact with P-selectin to limit the inflammatory response and help the parasite to survive (Perrin et al. 2015).

The above mentioned data raised the possibility that the interaction between the parasite and P-selectin may benefit the parasite in two ways; first, allowing the sequestration of trophozoite stage IEs to vascular endothelium and thus escape the splenic clearance and second, competition of merozoites with leukocytes for binding to P-selectin thus decrease the inflammatory response which may harm the parasite. All these facts promote to the importance of identifying the parasite's ligand(s) for these interactions.

### 5.8.2. IT4\_var02

In this study, an interesting finding was that upon selection and enrichment of IT4 *P. falciparum* isolate for binding to P-selectin receptor, group A IT4\_var02 *var* gene was differentially expressed among IT4\_P-selectin population. Group A *var* genes recently were underlined as a culprit in the pathogenesis of CM (this will be discussed later in this chapter). All other *var* genes that found to be differentially expressed in case of IT4\_Wild population were not differentially expressed in IT4\_P-selectin population; which seems to be it is a specific binding to P-selectin receptor. The *PfEMP1* variant encoded by this protein which contains the head structure with a non-binding CD36 domains (DBL $\alpha$ 1.5 and CIDR $\delta$ 1) which form DC16, followed by the four domains (DBL $\gamma$ 12 / DBL $\delta$ 5 / CIDR $\beta$ 3 and DBL $\beta$ 9) which form together DC5. Identifying the exact domain mediating the interaction to P-selectin receptor needs more studies.

Interesting facts considering DC5, it was linked to children presented with symptoms of severe malaria compared to children with non-severe malaria. In addition, antibodies against DC5 was found to be acquired in patients living in endemic areas (Berger et al. 2013, Jensen et al. 2004). In their study Berger and colleagues tested the binding phenotype of IEs to DC5 in 3D7 (PF11\_0008 gene) and IT4 (IT4\_var02 gene) they found that both bind strongly to bone marrow endothelial cells expressing both ICAM-1 and PECAM-1 (Berger et al. 2013). Trying to inhibit this binding with antibodies to ICAM-1 failed while it was inhibited by antibodies to PECAM-1. Another group tested the binding of IEs expressing IT4\_var02 to human bone marrow endothelial cells and tried to inhibit the binding with anti EPCR antibodies which failed to inhibit the binding (Turner et al. 2013).

### 5.8.3. IT4\_var07

Besides IT4\_var02, IT4\_var07 was differentially expressed in IT4\_P-selectin population. Interestingly its *PfEMP1* variant contains DC13 which composed of two domains (DBL $\alpha$ 1.7 and CIDR $\alpha$ 1.4) (Rask et al. 2010). In a recent study, high transcript of DC13 was found in patients with severe malaria and in comparison to patients with mild symptoms (Lavstsen et al. 2012). Adding to this, high binding capacity of IEs expressing DC13 to variable endothelial cell lines was also recently reported (Avril et al. 2013). It was also assumed that they mediate binding to EPCR via CIDR $\alpha$ 1.4 domain subclass (Turner et al. 2013). In a recent study, IT4\_var07 expressing parasite cell line was used to determine the role of CIDR $\alpha$ 1.4 in binding to EPCR. They found that this parasite line as well as another one (IT4\_var19) do bind to endothelial cell lines of brain, lung and dermis. Surprisingly, when they blocked EPCR, the binding was only inhibited to bone marrow and lung endothelial cells only but the IEs continue to bind over brain and dermis endothelial cells, which might suggest that other *PfEMP1* domains may bind to another receptor on these endothelial cells and confirm the heterogeneity of receptor usage by IEs (Gillrie et al. 2015).

### 5.8.4. Common domains between IT4\_var02 and IT4\_var07 *PfEMP1* variants

Searching for common domains between the two *var* genes expressed in IT4\_P-selectin population may give a clue of the most propable domain that mediate the binding on P-selectin. Firstly, two similar subclasses were observed forming a part of the head structure of these two genes; DBL $\alpha$ 1.5 in IT4\_var02 and DBL $\alpha$ 1.7 in IT4\_var07. Another common domain between the two *var* genes is DBL $\beta$ 3 domain, this domain was identified in a recent study to mediate binding to ICAM-1 but as a part of DC4 (Bengtsson et al. 2013). This needs more studies to figure out which of these domains are responsible for this binding phenotype. For example, raising antibodies against every single domain and test if the binding capacity to P-selectin will be affected or not. This type of study must be done with a pre-selected IT4 parasite line expressing a single *var* gene to avoid any bias in the results due to the binding of other IEs populations to the unknown receptor on CHO cells (discussed in part 5.3).

### 5.9. Still considering E-selectin as a receptor for *P. falciparum* IEs or not?

In this study, despite many controlling steps to confirm the surface expression of E-selectin on the CHO cells, it was not possible to identify any parasite population that have affinity to bind to E-selectin. In case of IT4\_E-selectin isolate only IT4\_var07 was differentially expressed in a first experiment but upon repetition of the experiment, the result could not be verified. Unfortunately, until now in the literature no ligands could be identified to mediate the binding to E-selectin. Also no information considering its role in experimental CM could be found.

Little information was available as follows; In Thailand, post-mortem brain examination from patients died from CM showed upregulation of E-selectin on the endothelial lining. In the same study it was also possible to select IEs for binding to E-selectin coated plates (Ockenhouse et al. 1992). On the other hand, also in Thailand a study reported that upon testing 60 patient isolates for binding to L cells (mouse cells) transfectants -expressing E-selectin on the surface- minimal IEs could bind to E-selectin compared to CD36 for example (Udomsangpetch et al. 1996). In another study no binding for E-selectin was found under flow conditions comparing to other receptors (Udomsangpetch et al. 1997). Recently, Janes and colleagues in their study tested the binding of IEs expressing Group A *var* genes to CHO cells transfectants expressing E-selectin on the surface and found very minimal binding capacity (Janes et al. 2011).

The controversial data mentioned above may support the results in this study that suggest that E-selectin plays minimal or no role in cytoadhesion of IEs to vascular endothelium and its upregulation in the brain of patients may be due to the inflammatory products produced by the inflammatory process provoked by the presence of IEs.

## 5.10. CD9 ligands

### 5.10.1. CD9 a new receptor which interact with *P. falciparum* IEs

In this study, the selection and enrichment for binding to CD9 receptor revealed differential expression of eight different *var* genes in IT4\_CD9 populations. Which might suggest a strong role of CD9 in the sequestration of IEs. CD9 was firstly described by Esser and colleagues, where they tested the affinity of a pool of patient isolates as well as IT4 isolate to bind to CD9. Where they reported significant binding in case of patient isolates and marginal binding using IT4 isolate. Inhibiting this binding using  $\alpha$ CD9 antibody showed statistically significant inhibition (Esser et al. 2014). Until now the interaction between IEs and CD9 is not yet studied in details neither in patients with CM nor in experimental CM. The leukocytes aggregation cascade is always a model to imagine the cytoadhesion of IEs. A very interesting study showed that, during inflammation leukocytes aggregation in the brain are mediated by endothelial adhesive platforms (EAPs) which are adhesive clusters containing CD9 and ICAM-1. Upon leukocyte binding via CD9, the EAPs rise above the surface to facilitate leukocyte capture (Franz et al. 2016). In infectious diseases, CD9 together with CD81 play a role in HIV infection by modulation of membrane fusion (Gordon-Alonso et al. 2006). CD9 is also involved at a post-entry step in the FIV (Feline immunodeficiency virus) life-cycle, possibly by interfering with FIV assembly and/or release. In bacterial infection it was found that CD9 interacts with proHB-EGF (pro heparin binding endothelial growth factor) which is the receptor with Diphtheria toxin (van Spruiel and Figdor 2010).

But the different *PfEMP1s* variants that could interact with CD9 in this study is somehow puzzlement. Where, four *PfEMP1s* are group A *PfEMP1s* (IT4\_var02, IT4\_var64, IT4\_var09 and IT4\_var07) and the other four are group B *PfEMP1s* (IT4\_var44, IT4\_var13, IT4\_var17 and IT4\_var41) which make it somehow difficult to predict the causative binding domains. Also some questions are addressed considering the role of CD9 as follows:

- What is the exact mechanism of *PfEMP1s*\_CD9 interactions?
- Does CD9 capture the IEs from circulation with its large extracellular loop or with the small one? Or both CD9 extracellular loops perform different functions in this interaction?

### 5.10.2. The significance of group A *var* genes

A very interesting finding in this study is that, four group A *var* genes (IT4\_var02, IT4\_var64, IT4\_var09 and IT4\_var07) were differentially expressed upon enrichment to CD9. Whereas, two of them were also expressed upon enrichment to P-selectin (IT4\_var02 and IT4\_var07). Group A *var* genes comprise about 20 % of *var* gene repertoire, they encode for *PfEMP1* variants that contain DC5, DC8 and DC13 which were previously discussed (see above). Actually group A *var* genes attracted the attention in the last few years. A study in Kenya, they measured the expression of *var* genes in 216 samples from children infected with *P. falciparum* they found that group A *var* genes were highly expressed and this is in association with severe symptoms such as impaired consciousness (Warimwe et al. 2012). In Tanzania, a similar study was conducted to compare between the *var* gene transcript in children with asymptomatic, symptomatic and severe malaria, Group A *var* genes had the highest expression profile among children with severe malaria compared to others (Rottmann et al. 2006).

In *P. falciparum* infection sterile immunity requires antibodies against broad range of *PfEMP1s* variants. Recently, a model of infection in Africa was suggested based on several studies and was reviewed by (Duffy et al. 2016) as follows, Children and naïve individuals suffer from severe malaria which is accompanied by group A and B *PfEMP1s* that stimulate the production of antibodies against these two groups. Whereas, semi-immune individuals exhibit some antibodies against both group A and B *PfEMP1s*. There was also a suggested sequelae for the infection cascade as follows; firstly, parasites with DC8, DC15 and DC13 will cause initial infection and severe disease symptoms which is then controlled by the immune response generating antibodies against them. The next step to evade the immunity, therefore the parasites with group C *PfEMP1s* and other *PfEMP1s* will dominate the infection without being recognized by the host antibody response (Kalmbach et al. 2010, Warimwe et al. 2009, Jensen et al. 2004, Gupta et al. 1999, Chan et al. 2012).

The above mentioned data raised the importance of characterization of the interactions of group A *PfEMP1s* with host receptors and in this study four members of group A *PfEMP1s* were identified to interact with two host receptors (P-selectin and CD9).

### 5.11. No ligands for CD151

In this study two attempts to select for CD151 binding were performed. For this the surface expression of CD151 on CHO cells was confirmed by immunofluorescence microscopy and the cells were sorted using FACS. In two independent experiments nine different biological probes were investigated. Data analysis showed that, the selection was only made over the unknown receptor over CHO cells, this because the *var* gene transcripts in both populations (IT4\_Wild and IT4\_CD151 as well as 3D7\_Wild and 3D7\_CD151) were exactly the same. As a result, it could be concluded that CD151 most probably do not play a role in the cytoadhesion of IEs.

But these conclusion is inconsistent with the study made by Esser and colleagues where they identified CD151 as a receptor that could interact with IEs from a pool of patient isolates as well as with IT4 isolate. But in their results they mentioned some facts that might be helpful to explain the data in these two studies. First, the binding capacity of IT4 isolate seemed to be negligible considering CD151 receptor compared to CD9 receptor to which IT4 showed a higher binding capacity. Second, in their trial to inhibit the binding with antibodies against the two tetraspanins, the antibody against CD9 inhibited more percentage of IEs than in case of the antibody against CD151 which seemed to cause minimal inhibition. This might suggest that the binding phenotype described in their study might be to the unknown receptor on the surface of CHO cells (discussed in part 5.3). These explanations and the results of this study could rule out CD151 as a receptor used by *P. falciparum* IEs in the cytoadhesion.

### 5.12. The ligands in 3D7 isolate

Actually, the results of 3D7 enrichment were disappointing in all IEs populations with different receptors; every attempt to select and enrich 3D7 over any receptor the same IEs population expressing MAL6P1.252 *var* gene was the dominant one. In case of 3D7\_P-selectin and 3D7\_CD9 IEs populations, the expression level of MAL6P1.252 *var* gene was statistical significant. Whereas, in case of 3D7\_E-selectin and 3D7\_CD151, its level was highly differentially expressed compared to 3D7\_Ctrl but not to 3D7\_Wild.

The interpretation of these results will have many points to discuss. Step by step through 3D7 IEs populations dynamics, we could observe the following. First, the expression levels of other *var* genes in the 3D7\_Ctrl population, it was observed that all *var* genes were expressed at different levels this means that there is no error in *var* gene expression. The second evidence that, it was possible to select for ICAM-1 binding which revealed differential expression of other two *var* genes (see results part 4.2.6.1.2), which means, it is possible to select other populations. Third, it is obviously that, the selection and enrichment were over the unknown receptor expressed by CHO wild type cells (discussed in part 5.3). The unbelievable expression of this *var* gene in all the IEs populations may be explained by the presence of more knobs on the IEs expressing this gene, which facilitates its binding in tremendous amount (Fig. 4.27). Another alternative explanation for the abundance of this population might be any cause in the culturing techniques which favours the survival and rapid growth of this population than the others.

In order to test the binding of 3D7 isolate to these receptors, we could try the other way on. For example, selecting 3D7 cell line expressing genes that encodes for PfEMP1 domains similar to those found in case of IT4 isolate such as PF13\_0003 which encodes for a protein with both domains (DBL $\alpha$ 1.6 and CIDR $\delta$ 1) representing DC16 which was found in IT4\_var02. Also the two *var* genes PFD1235w and PFD0020c encodes for two sub classes (DBL $\alpha$ 1 and CIDR $\alpha$ 1) which were encoded by both IT4\_var07 and IT4\_var64. A method for limited dilution cloning for *P. falciparum* culture was mentioned by (Gillrie et al. 2015, Janes et al. 2011). Selecting these cell lines and adopting them in culture for few cycles may enable the puzzling testing of domains adhesion affinity; blocking a domain, a test for the binding affinity to a certain receptor will be an approach to test the binding capacity of 3D7 isolate and correlate the results to those from IT4 isolate.

### 5.13. Is it possible to apply this system to select patient isolates for binding?

The success of the system used in this study in identifying IEs ligands to different receptors and exclude the role of other receptors, brought this question for discussion. Well, the most important stone in building this system was the stability of *var* genes switching pattern in the two laboratory adopted isolates used in this study. On the other hand, using patient isolates might bring more variability considering the different *var* gene repertoires and their interactions with different host receptors. The problem upon using patient isolates will be the quick *var* gene switching pattern which will make it difficult to identify the *var* gene encodes for the *PfEMP1*s that interact with host receptors. The solution may be to use *ex vivo* isolates which might bring more interesting and detailed results considering the host parasite interactions. In *ex vivo* isolates the rate of *var* gene switching is slower than fresh patient isolates and quicker than the laboratory adapted ones. So in order to use this system with *ex vivo* isolates, a careful measurement of the *var* gene switching pattern must be considered.

### 5.14. To summarize

The current study identified *PfEMP1* candidates that could interact with two host receptors; the first one, is P-selectin which was described as an old adhesion partner with *PfEMP1* but its ligand is not yet identified. The second is, CD9 which was newly described as a receptor used by *P. falciparum* IEs. The most interesting finding is that, four from the *PfEMP1* variants identified in this study belong to group A *PfEMP1*s which is linked to patients presented with severe malaria. Also this study ruled out two receptors from being accused to interact with IEs which are E-selectin and CD151. As a proof of concept ligands for ICAM-1 were also identified and the results were compatible with results from previous studies. Finally, an interesting binding phenotype was observed between IEs and senescent cells present in the CHO cell culture, which opens many questions to the nature of this binding phenotype.

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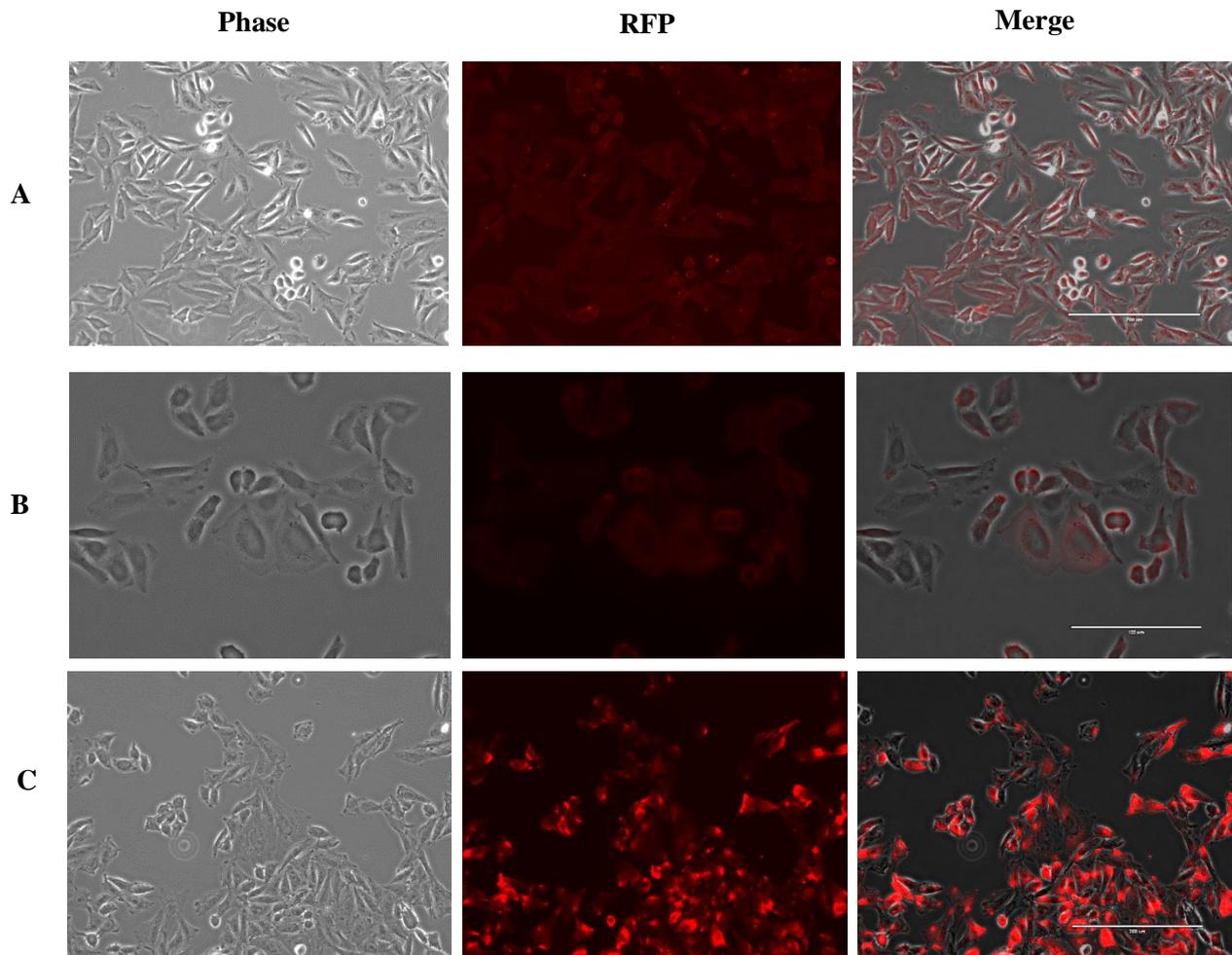
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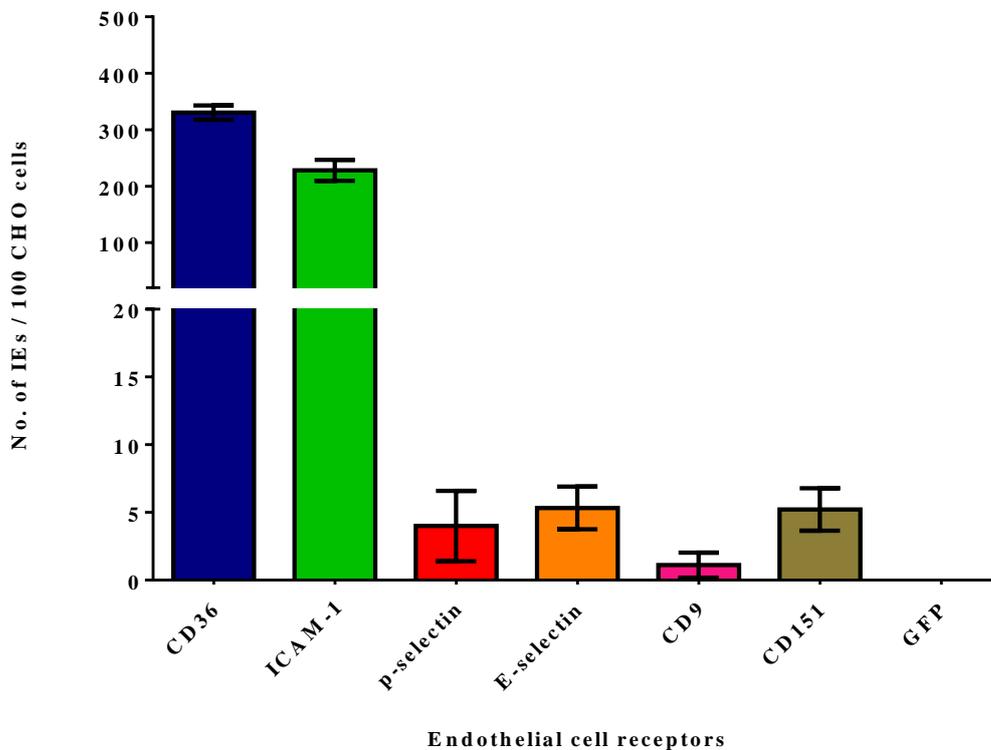
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## 7. Supplements



**Supp. fig. 1. Immunofluorescence assays to test CD44 expression on the surface of CHO cells - Wild types.** Cells were grown on coverslips and fixed with 4% para-formaldehyde. (A) and (B) CHO wild types were labelled using  $\alpha$  CD44 antibody (rat-anti mouse) (monoclonal KM114) and secondary antibody conjugated to Alexa-Fluor-594 (Invitrogen) which did not show any reaction without the first antibody. (C) CHO wild types were labelled using  $\alpha$  CD44 antibody (rat-anti-mouse/human) (monoclonal IM7) conjugated to PE/Cy7.



**Supp. fig. 2. Static binding assays of IT4\_ICAM-1 enriched cell line to various endothelial receptors.** After incubation of trophozoite stage IEs over CHO monolayer, the slides were carefully washed, fixed with 1% glutaraldehyde and finally stained with Giemsa. Each slide was then examined under light microscope and the amount of IEs bound to 100 CHO cells was quantified three times for every slide. Bars represent the mean  $\pm$ SD of IEs numbers that bound to 100 CHO cells as determined by microscopic inspection and the following was observed: IT4\_ICAM-1 showed high binding affinity to the CD36 as well as to ICAM-1, whereas GFP refers to the negative control (mock transfected). On the other hand, minimal binding capacity was observed considering the selectins (P-selectin and E-selectin) and tetraspanins (CD9 and CD151).

Gene name	Parasite	UPS	NTS	Extracellular Domain Structure (Predicted)												
IT4var08	IT4	A1	A6	DBLa1.7	CDRy3	DBLp3	DBLp4	CDRy1	DBLp8	DBLp9						
IT4var35	IT4	A2*	A1	DBLa1.1	CDRa1.2	DBLp11	DBLy1	DBLa1	DBLy6	DBLp1	DBLp5					
IT4var03	IT4	A3	A6	DBLa1.3	DBLp8											
IT4var02	IT4	A3	A6	DBLa1.5	CDRy1	DBLp3	DBLy12	DBLp5	CDRy3	DBLp9						
IT4var07	IT4	A3	A6	DBLa1.7	CDRa1.4	DBLp1	DBLp3	DBLy10	DBLp1	CDRy1						
IT4var18	IT4	A3	A3	DBLa1.2	CDRa1.6	DBLp3	DBLy16	DBLy2	DBLp1	CDRy1						
IT4var64	IT4	A3	A5	DBLa1.5	CDRy1	DBLp7	DBLy11	DBLy11								
IT4var22	IT4	A3	A2	DBLa1.4	CDRa1.7	DBLp3	DBLy10	DBLy11	DBLp1	CDRy1						
DC408104	IT4	A3	A5	DBLa1.5												
IT4var60	IT4		A7	DBLa1.8	CDRy3	DBLy7	DBLp5	DBLp11	DBLp12							
IT4var9	IT4		A8	DBLa1.8	CDRy3	DBLy15	DBLp1	DBLp1	CDRy1							
IT4var14	IT4		B3	DBLa0.23	CDRa5	DBLp5	DBLp1	CDRy2	DBLy3	DBLp4						
IT4var46	IT4	B1	B3	DBLa0.10	CDRa2.2	DBLp1	CDRy4	DBLp2	DBLp3	DBLp3						
IT4var10	IT4	B1	B3	DBLa0.5	CDRa2.6	DBLp10	DBLp1									
IT4var67	IT4	B1	B3	DBLa0.10	CDRa2.2	DBLp2	DBLp3	DBLp12								
IT4var13	IT4	B1	B3	DBLa0.3	CDRa5	DBLp5	DBLp9	CDRy9	DBLy11	DBLp4						
IT4var19	IT4	B1	B3	DBLa2	CDRa1.1	DBLp12	DBLy6	DBLp1	CDRy1	DBLy9						
IT4var32a	IT4		B3	DBLa0.23	CDRa3.2	DBLy6	DBLp1	CDRy1								
IT4var32b	IT4	B1	B3	DBLa2	CDRa1.1	DBLp12	DBLy6	DBLp1	CDRy1							
IT4var06	IT4	B1	B3	DBLa2	CDRa1.1	DBLp12	DBLy6	DBLp1	CDRy4							
IT4var41	IT4	B1	B3	DBLa0.4	CDRa5	DBLp5	DBLy5	DBLp1	CDRy1							
IT4var16	IT4	B1	B3	DBLa0.4	CDRa5	DBLp5	DBLy16	DBLp1	CDRy6							
IT4var31	IT4	B1	B3	DBLa0.16	CDRa4	DBLp3	DBLy9									
IT4var44	IT4	B1	B3	DBLa0.16	CDRa3.4	DBLp13	DBLp1	CDRy6								
IT4var11	IT4	B1	B3	DBLa0.3	CDRa2.4	DBLp10	DBLp1	CDRy1								
AA90961	IT4	B1	B3	DBLa0.16	CDRa6	DBLp4	DBLp1	CDRy1								
IT4var12	IT4		B3	DBLa0.18	CDRa6	DBLp4	DBLp1	CDRy1								
IT4var17	IT4	B1	B3	DBLa0.11	CDRa2.4	DBLp8	DBLp1	CDRy4		DBLy10						
IT4var29	IT4	B1	B3	DBLa0.3	CDRa3.2	DBLp1	CDRy5									
IT4var24	IT4	B1	B3	DBLa0.10	CDRa2.2	DBLp1	CDRy5									
IT4var45	IT4	B1	B3	DBLa0.5	CDRa2.9	DBLp1	CDRy1									
IT4var33	IT4	B1	B3	DBLa0.11	CDRa2.4	DBLp1	CDRy6									
IT4var25	IT4	B1	B3	DBLa0.11	CDRa2.4	DBLp1	CDRy1									
IT4var61	IT4	B1	B3	DBLa0.9	CDRa2.7	DBLp1	CDRy1									
IT4var54	IT4	B1	B3	DBLa0.1	CDRa3.1	DBLp1	CDRy1									
IT4var63	IT4	B1	B3	DBLa0.7	CDRa3.4	DBLp1	CDRy1									
IT4var26	IT4	B1	B3	DBLa0.23	CDRa3.3	DBLp1	CDRy1									
IT4var40	IT4	B1	B3	DBLa0.12	CDRa2.11											
IT4var20	IT4	B2	B3	DBLa2	CDRa1.1	DBLp12	DBLy6	DBLp1	CDRy1							
IT4var27	IT4	B3	B3	DBLa0.6	CDRa3.1	DBLp5	DBLy5	DBLp1	CDRy1							
IT4var58	IT4	B3*	B3	DBLa0.1	CDRa3.1	DBLp1	CDRy6									
IT4var59	IT4	B3*	B3	DBLa0.1	CDRa3.1	DBLp1	CDRy1									
IT4var21	IT4		B3	DBLa0.1	CDRa3.1	DBLp1	CDRy1									
IT4var15	IT4	B7	B3	DBLa0.8	CDRa3.5	DBLp8	DBLp1	CDRy1								
IT4var01	IT4	C1	B6	DBLa0.18	CDRa6	DBLp5	DBLy10	DBLp2	CDRy6							
IT4var51	IT4	C1	B3	DBLa0.17	CDRa3.1	DBLp1	CDRy12									
IT4var66	IT4	C1	B3	DBLa0.19	CDRa2.10	DBLp1	CDRy1									
IT4var23	IT4	C1	B3	DBLa0.1	CDRa3.1	DBLp1	CDRy10									
IT4var05	IT4	C1	B3	DBLa0.5	CDRa2.3	DBLp1	CDRy1									
IT4var34	IT4	C1	B3	DBLa0.1	CDRa3.1	DBLp1	CDRy4									
IT4var47	IT4	C1	B3	DBLa0.1	CDRa3.3	DBLp1	CDRy1									
IT4var62	IT4	C1	B3	DBLa0.1	CDRa3.1	DBLp8	CDRy2									
IT4var68	IT4	C1	B3	DBLa0.1	CDRa3.2											
IT4var28	IT4	C2	B2	DBLa0.4	CDRa3.1	DBLp1	CDRy6									
IT4var30	IT4			DBLa0.13	CDRa2.10	DBLp1	CDRy1									
IT4var36	IT4			DBLa0.8	CDRa5	DBLp1	CDRy1									
IT4var39	IT4		B3	DBLa0.5	CDRa2.5	DBLp1	CDRy6									
IT4var04	IT4	E	pam	DBLpam1	DBLpam2	CDRpam	DBLpam3	DBLpam4	DBLpam5	DBLpam6	DBLpam10					
IT4var65	IT4			CDRy1												

Supp. fig. 3. IT4 *PfEMP1s* variants (predicted extracellular protein structure) (Rask et al. 2010).

Gene name	Parasite	UPS	NTS	Extracellular Domain Structure (Predicted)														
PF13_0003	3D7	A1	A3	DBLe1.6	CIDR31	DBLβ3	DBLy12	DBLβ5	CIDRβ3	DBLβ9								
PF11_0008	3D7	A1	A6	DBLe1.5	CIDRγ3	DBLy12	DBLβ5	CIDRβ4	DBLβ9									
PF11_0521	3D7	A1	A6	DBLe1.7	CIDRα1.4	DBLβ3	DBLβ6	DBLβ3	CIDRγ2									
PFD1235w	3D7	A1	A2	DBLe1.4	CIDRα1.5	DBLβ3	DBLβ3	DBLy13	DBLβ1	CIDRβ5								
PFD0020c	3D7	A1	A7	DBLe1.2	CIDRα1.1	DBLβ12	DBLy6	DBLy11	DBLβ1	CIDRβ8								
PFE1640w	3D7	A2	A1	DBLe1.4	CIDRα1.3	DBLβ1	DBLy15	DBLβ1	DBLy8	DBLβ1	DBLβ5							
PF08_0141	3D7	A3	A5	DBLe1.8	CIDRβ2	DBLβ6	DBLy14	DBLβ5	DBLe4									
PFA0015c	3D7	A3	A3	DBLe1.3	DBLe8													
MAL6P1.314	3D7	A3	A3	DBLe1.3	DBLe8													
PF11820w	3D7	A3	A3	DBLe1.3	DBLe8													
PF10_0001	3D7	B1	B3	DBLe0.20	CIDRα3.1	DBLβ1	CIDRγ4											
PFI0005w	3D7	B1	B3	DBLe0.16	CIDRα3.4	DBLβ1	CIDRγ12											
PFL0935c	3D7	B1	B3	DBLe0.16	CIDRα3.4	DBLβ1	CIDRγ12											
PF11830c	3D7	B1	B3	DBLe0.1	CIDRα3.1	DBLβ1	CIDRγ5											
MAL7P1.50	3D7	B1	B3	DBLe0.1	CIDRα3.1	DBLβ1	CIDRγ10											
PF08_0142	3D7	B1	B3	DBLe0.9	CIDRα2.11	DBLβ1	CIDRβ1											
PFL0005w	3D7	B1	B3	DBLe0.9	CIDRα2.2	DBLβ1	CIDRβ1											
PFA0785c	3D7	B1	B3	DBLe0.5	CIDRα2.8	DBLβ1	CIDRβ1											
PFA0005w	3D7	B1	B3	DBLe0.11	CIDRα2.8	DBLβ1	CIDRβ1											
PFL2665c	3D7	B1	B3	DBLe0.19	CIDRα2.3	DBLβ1	CIDRβ1											
PF13_0001	3D7	B1	B3	DBLe0.11	CIDRα2.4	DBLβ1	CIDRβ1											
PFE0005w	3D7	B1	B3	DBLe0.11	CIDRα2.4	DBLβ1	CIDRβ1											
PFC0005w	3D7	B1	B3	DBLe0.9	CIDRα2.4	DBLβ1	CIDRβ1											
PF10_0408	3D7	B1	B3	DBLe0.9	CIDRα2.7	DBLβ1	CIDRβ1											
MAL7P1.212	3D7	B1	B3	DBLe0.17	CIDRα3.1	DBLβ1	CIDRβ1											
MAL8P1.220	3D7	B1	B3	DBLe0.1	CIDRα3.1	DBLβ1	CIDRβ1											
PFB1055c	3D7	B1	B3	DBLe0.16	CIDRα3.4	DBLβ1	CIDRβ1											
PF13_0384	3D7	B1	B3	DBLe0.16	CIDRα3.4	DBLβ1	CIDRβ1											
PF11_0007	3D7	B1	B3	DBLe0.15	CIDRα3.2	DBLβ1	CIDRβ1											
MAL6P1.1	3D7	B1	B3	DBLe0.8	CIDRα4	DBLβ1	CIDRβ1											
PFD1245c	3D7	B1	B3	DBLe0.14	CIDRα4	DBLβ1	CIDRβ1											
PFC1120c	3D7	B1	B3	DBLe0.9	CIDRα2.1	DBLβ1	CIDRβ7											
PF07_0139	3D7	B1	B3	DBLe0.16	CIDRα3.4	DBLβ1	CIDRβ1	DBLe4										
PFB0010w	3D7	B1	B3	DBLe0.7	CIDRα2.2	DBLy11												
PFD0005w	3D7	B1	B3	DBLe0.5	CIDRα2.10	DBLy5	DBLβ1	CIDRβ1										
MAL6P1.316	3D7	B2	B3	DBLe2	CIDRα1.8	DBLβ12	DBLy4	DBLβ3	DBLe12									
PF08_0140	3D7	B2	B5	DBLe2	CIDRα1.5	DBLβ12	DBLy4	DBLβ1	CIDRβ1									
PFD07_0050	3D7	B3	B3	DBLe0.18	CIDRα6	DBLβ6	DBLy9											
PFD0635c	3D7	B3	B3	DBLe0.1	CIDRα3.1	DBLβ1	CIDRγ2											
PFL1950w	3D7	B4	B3	DBLe0.7	CIDRα4	DBLβ8	DBLβ1	CIDRβ1										
MAL6P1.4	3D7	B5	B3	DBLe0.6	CIDRα3.2	DBLβ5	DBLy13	DBLβ4	CIDRγ1	DBLe2	DBLe7	DBLe3						
PFL0020w	3D7	B6	B3	DBLe0.18	CIDRα5	DBLβ5	DBLy14	DBLβ5	DBLe4									
PFD1005c	3D7	B5	B3	DBLe0.8	CIDRα4	DBLβ1	CIDRγ11											
PF08_0103	3D7	B6	B3	DBLe0.12	CIDRα2.2	DBLβ1	CIDRβ1											
MAL7P1.55	3D7	B7	B3	DBLe0.9	CIDRα2.4	DBLβ1	CIDRγ5											
PF08_0106	3D7	B7	B3	DBLe0.2	CIDRα3.1	DBLβ1	CIDRβ1											
PFL1955w	3D7	B7	B3	DBLe0.16	CIDRα3.4	DBLβ1	CIDRβ1											
MAL6P1.252	3D7	C1	B1	DBLe0.21	CIDRα2.1	DBLβ4	DBLβ1	CIDRβ1										
PFD0995c	3D7	C1	B3	DBLe0.1	CIDRα3.2	DBLβ1	CIDRγ6											
PFD1000c	3D7	C1	B3	DBLe0.1	CIDRα3.2	DBLβ1	CIDRγ11											
PF07_0049	3D7	C1	B3	DBLe0.17	CIDRα3.1	DBLβ1	CIDRγ7											
PFD0630c	3D7	C1	B3	DBLe0.1	CIDRα3.1	DBLβ1	CIDRγ2											
PF08_0107	3D7	C1	B3	DBLe0.15	CIDRα3.2	DBLβ1	CIDRγ2											
PF07_0051	3D7	C1	B3	DBLe0.1	CIDRα3.1	DBLβ1	CIDRβ1											
PFD0615c	3D7	C1	B3	DBLe0.17	CIDRα3.1	DBLβ1	CIDRβ1											
PFL1960w	3D7	C1	B3	DBLe0.20	CIDRα3.1	DBLβ1	CIDRβ1											
PFD1015c	3D7	C1	B3	DBLe0.24	CIDRα3.4	DBLβ1	CIDRβ1											
PFD0625c	3D7	C1	B3	DBLe0.1	CIDRα3.2	DBLβ1	CIDRβ1											
PF07_0048	3D7	C1	B3	DBLe0.1	CIDRα3.2	DBLβ1	CIDRβ6											
MAL7P1.56	3D7	C2	B3	DBLe0.20	CIDRα3.1	DBLβ1	CIDRβ1											
PFL0030c	3D7	E	pam	DBLpam1	DBLpam2	CIDRpam	DBLpam3	DBLpam4	DBLpam5	DBLpam6	DBLpam7	DBLpam8	DBLpam9	DBLpam10				

Supp. fig. 4. 3D7 *PfEMP1s* variants (predicted extracellular protein structure) (Rask et al. 2010).

Supp. table 1. var genes expression profiles of IT4\_Ctrl compared to IT4\_Wild IEs

var genes (Nom ID)	IT4_Ctrl	IT4_Ctrl2	IT4_Ctrl3	IT4_Ctrl4	IT4_Wild	IT4_Wild2	IT4_Wild3	baseMeanA	baseMeanB	foldChange	log2FoldChange	P-value	Padj
PFIT_0710800	862.1408	2762.334	2043.427	1342.882	39.66905	48.59286	48.59286	37.35066598	0.023110409	-5.552297884	4.7E-06	0.000312025	
PFIT_0810000	119.2468	48.38819	36.58715	52.62644	1484.504	3271.919	4617.930	64.21215009	0.486109737	312.4620846	5.004691021	4E-05	0.0004666304
PFIT_bin00100	4.797286	3.155734	7.662087	602.9696	3352.908	4543.906	5.437714519	2833.261067	521.0389506	9.025247416	7.4E-05	0.0004806304	
PFIT_bin00900	23.98643	11.57109	16.13471	15.12254	828.6424	4227.579	5287.887	16.70369127	3448.036124	206.4369127	0.00021	0.013256645	
PFIT_0419300	115.8202	328.1982	154.0751	117.5509	120.3448	0	6.488205	178.8610856	0.067570065	-4.832721104	0.00358	0.225286745	
PFIT_bin0483000	96.63104	280.8619	149.9846	120.7787	22.03836	0	10.81367	162.0640534	10.950678666	0.048075138	0.00179	0.482750555	
PFIT_0120000	398.86	97.7909	374.7343	267.5682	49.36593	0	21.62735	492.2383452	3.266426263	-4.387471942	0.01479	0.902005095	
PFIT_bin02100	45.91688	124.1262	115.8972	93.96139	12.34148	0	2.162735	94.97541972	4.834739099	0.050905162	0.04779	0.902005095	
PFIT_bin11000	6.167939	14.72684	8.87189	95.20572	469.731	772.0964	0	11.41853171	45.6776997	39.03108636	0.02628	1	
PFIT_bin07000	23.3011	93.62063	59.08484	54.23951	5.289207	0	57.56152098	1.763068911	0.030629297	-5.028943944	0.03878	1	
PFIT_0616500	18.97447	116.7628	117.58359	56.2941	160.4393	696.4977	798.0492	65.40953125	15.579479917	3.901575099	0.04664	1	
PFIT_0616500	2.05598	12.62301	9.998973	7.460453	32.61677	145.7786	168.69233	80.34603046	115.6962295	3.847971289	0.06261	1	
PFIT_0810000	171.3316	450.2205	165.1381	51.129	48.59286	38.29233	237.8047206	46.2170301	0.194348665	-2.363280894	0.07081	1	
PFIT_bin06200	4.797286	6.83067	10.68072	4.032677	31.73524	210.5691	170.8561	0.085339826	137.7201245	3.922055102	0.09297	1	
PFIT_0411000	108.9669	374.4825	118.3969	68.95878	71.73069	0	8.65094	167.7012868	8.258729667	-4.258729667	0.11056	1	
PFIT_0710600	126.1001	184.0855	78.62829	71.37839	28.2091	16.19762	28.11555	115.0480669	24.17409254	-2.250703103	0.11464	1	
PFIT_bin02200	82.23918	215.643	188.1625	123.8032	59.94434	16.19762	34.60376	152.4619747	28.29209912	-2.046160988	0.17863	1	
PFIT_bin01000	98.00169	129.3858	72.26531	66.13591	30.85371	32.39524	21.62735	91.44718084	38.29209912	-1.692539474	0.2224	1	
PFIT_1219000	41.80492	105.1917	33.86016	24.59933	12.34148	0	15.13914	51.36403176	0.178338978	-2.48730604	0.32596	1	
PFIT_0411000	190.5208	842.5856	127.4869	73.798	38.78752	16.19762	32.79008	308.5978326	26.25840709	-3.554876623	0.36728	1	
PFIT_bin04000	28.09839	7.46342	4.999487	6.25065	29.09064	0	15.13914	11.67798623	14.74326049	0.336264098	0.39331	1	
PFIT_0411000	47.97286	29.45368	29.31517	35.28593	44.07672	80.98811	116.7877	35.50690946	80.61750463	2.270473715	0.52822	1	
PFIT_0100100	25.35708	33.66135	24.99743	20.96992	35.26138	16.19762	60.55688	26.2464468	37.33852568	-0.508542837	0.55541	1	
PFIT_bin06100	8.909254	12.62301	13.18046	3.71111	29.09064	32.39524	15.13914	12.10595479	25.54167457	1.077136246	0.53894	1	
PFIT_0810000	2.05598	5.259586	9.771724	8.84738	10.57841	0	12.97641	5.733667921	7.851607657	1.369386537	0.57177	1	
PFIT_1310000	10.2799	31.55713	12.86221	12.5013	3.526138	0	2.162735	17.05023151	1.896290913	-0.453529734	0.59771	1	
PFIT_0411000	32.89567	99.93713	77.49204	56.25585	52.01053	210.5691	287.6437	66.64392451	183.4078938	2.752055572	0.60857	1	
PFIT_1150900	13.70653	25.46011	9.3162	12.90457	2.56545	0	1.466461	17.79332725	15.00970449	-0.245440738	0.60857	1	
PFIT_bin08950	10.2799	0	0.681748	2.01639	6.17041	0	0	3.244496247	2.77825369	-0.224038591	0.65078	1	
PFIT_bin03000	9.594572	7.36342	7.044751	7.662087	14.98609	16.19762	6.85094	7.916202514	13.27821551	1.677346618	0.65308	1	
PFIT_0900100	33.581	31.55713	11.58972	9.88006	7.052276	0	4.32547	21.62502351	3.792581827	-2.513252979	0.67012	1	
PFIT_1310000	0	5.259586	3.408741	2.801639	3.526138	0	2.162735	2.671166361	1.896290913	-0.494289523	0.70336	1	
PFIT_1100200	8.223918	48.54471	20.1112	12.95195	24.99415	23.85921	23.85921	24.86217208	0.975893053	-0.035205042	0.80766	1	
PFIT_bin09200	31.70653	4.207669	0	0.806535	2.644603	0	4.32547	6.367477628	4.086246645	-0.252193758	0.81289	1	
PFIT_1200200	8.909245	10.51917	4.090489	5.242481	8.815345	0	0	30.41462969	52.98799828	1.742187849	0.81744	1	
PFIT_0810900	9.594572	6.311503	3.181492	3.226142	0	0	0	30.016791	20.75755798	-0.532133023	0.82498	1	
PFIT_1200200	31.52502	31.55713	51.35836	52.82807	25.5645	32.39524	36.706649	41.81724336	31.57541168	-0.405296417	0.79452	1	
PFIT_0811900	16.65069	48.54471	20.1112	12.95195	24.99415	23.85921	23.85921	24.86217208	0.975893053	-0.035205042	0.80766	1	
PFIT_0731700	8.223918	36.8171	24.99743	14.10845	32.39524	0	6.85094	2.818514984	18.38357773	0.839618722	0.81289	1	
PFIT_bin04300	56.19678	28.40176	19.3162	17.4378	42.31365	97.18573	19.46461	30.41462969	52.98799828	1.742187849	0.81744	1	
PFIT_0835600	40.43427	32.60943	23.63394	33.38953	35.26138	16.19762	10.81367	30.016791	20.75755798	-0.532133023	0.82498	1	
PFIT_0710900	0.685327	6.311503	3.86324	4.637579	5.289207	16.19762	4.32547	3.874412072	8.604099229	1.15104676	0.82958	1	
PFIT_bin09700	2077.225	4121.411	2630.639	1837.086	2669.286	2267.667	2484.982	2666.590344	2473.978569	-0.927768517	0.91196	1	
PFIT_1151000	26.72773	19.98643	17.27095	13.30784	7.052276	0	19.46461	19.332323765	8.8389633	-1.128387767	0.91364	1	
PFIT_1240400	67.84733	71.53037	25.67918	23.7928	52.01053	80.98811	21.62735	47.21241818	51.54199588	0.126582026	0.91742	1	
PFIT_0731700	0.685327	1.051917	1.136247	1.008169	0	16.19762	4.32547	0.970415009	7.049592448	2.817539855	0.93853	1	
PFIT_1400100	27.41306	7.36342	6.362983	7.863721	7.93381	16.19762	15.13914	12.5079643	13.09019188	1.068517623	0.95532	1	
PFIT_1240000	15.76251	26.29793	16.36196	16.73561	0	6.85094	10.81367	27.6420088	13.00759239	-1.48691256	0.97831	1	
PFIT_bin10900	56.8821	39.97285	48.63137	69.56369	41.43212	16.19762	54.06837	53.76250285	37.23270449	-0.550029889	0.98747	1	
PFIT_0411300	41.11959	4.207669	4.999489	6.049016	2.644603	0	2.162735	4.842032685	1.602446095	-1.595336992	1	1	
PFIT_0419500	6.167939	0	1.363496	1.613071	1.763069	0	2.162735	2.286126548	1.308601276	-0.804879683	1	1	
PFIT_0500100	41.11959	30.5056	22.27071	26.44603	0	12.97641	29.97660005	3.140814339	0.438369074	-1.189782074	1	1	
PFIT_0536800	1436.444	4489.209	1865.718	1729.212	1939.376	1927.517	1805.884	21.93600005	18.90425457	-0.1718597	1	1	
PFIT_0537600	17.13316	50.49202	24.99743	17.94541	28.2091	0	10.81367	27.6420088	13.00759239	-0.470573339	1	1	
PFIT_0800100	15.76251	28.40176	29.9692	22.17973	15.86762	0	2.162735	24.08522994	12.49832312	-0.946414153	1	1	
PFIT_1400200	6.167939	21.03834	7.271981	5.242481	5.289207	16.19762	2.162735	15.19655862	7.88318759	-1.604271319	1	1	
PFIT_1401900	5.482612	13.67492	23.31569	19.96175	7.93381	16.19762	32.44102	15.8587435	18.85748499	0.60122258	1	1	
PFIT_bin02000	3.426633	3.155751	3.427766	3.427766	3.26138	0	0	3.468349929	1.175379274	-1.561123098	1	1	
PFIT_bin02700	4.797286	26.29793	29.08792	20.16339	9.696879	16.19762	47.58017	20.08663113	24.4915561	-0.28604882	1	1	
PFIT_bin04200	1.370653	5.259586	4.772237	4.234311	1.763069	0	4.32547	3.909196873	2.029512915	-0.945738722	1	1	
PFIT_bin07850	23.98643	11.57109	14.77121	8.670257	12.34148	0	6.488205	14.74974619	6.276562377	-0.425536975	1	1	
PFIT_bin10700	29.46904	27.34985	42.26839	23.99443	35.26138	16.19762	6.85094	6.301863423	3.338114191	-0.91674518	1	1	
PFIT_bin10800	8.909245	11.57109	9.544475	9.299443	11.45995	0	4.32547	9.925808557	5.261805919	-0.618903379	1	1	
PFIT_bin10300	0	0	0.227249	0.201634	0	0	0	0.107220817	0	-0.51165704	1	1	
PFIT_bin10100	0	0	0	0	0	0	0	0	0	-0.1915626593	1	1	

Supp. table 2. Part of the whole transcriptome analysis of IT4\_Ctrl compared to IT4\_Wild IEs populations

	IT4_Ctrl	IT4_Chr2	IT4_Chr3	IT4_Chr4	wild_1	wild_2	wild_3	baseMcanA	baseMcanB	foldChange	log2FoldChange	P-value	name
PFTT_0710800	862.1407846	2762.32446	2043.4265	1342.8816	39.6691	48.59286	33.79008409	37.5065849	0.021310409	-5.552297884	0.02598	erythrocyte membrane protein 1, PTEMPI	
PFTT_0711000	21.2468176	48.3881894	36.587152	52.626441	1484.5	3271.919	461.7439048	3124.6208646	48.6608737	5.604691021	0.11269	C-ITyae28 - erythrocyte membrane protein 1, PTEMPI, putative	
PFTT_b040100	4.797285765	3.15575148	6.1357336	7.6020872	602.97	3352.908	4543.900661	5.437714519	2833.261067	9.025247416	0.13751	sfb_C52var - erythrocyte membrane protein 1, putative	
PFTT_b040500	490.0084746	605.904284	450.8628	292.57075	2306.98	1749.343	191.857667	1989.391339	42.932032518	2.11313455	0.00020774	ADPA/TP transporter on adenylate translocase	
PFTT_b040900	23.98642883	11.57108888	16.134707	15.122541	828.642	4227.579	287.888687	3448.036124	206.4236024	7.689464127	0.000207132	erythrocyte membrane protein 1, PTEMPI	
PFTT_0419800	14813.33312	3859.48406	3849.3775	4052.0343	68547.2	56092.36	26595.15128	6643.557246	50411.58365	7.588040832	0.22724	signal recognition particle RNA	
PFTT_1025700	248.0882067	142.008817	150.4391	386.9354	793381	0	11.555392	12.01645467	0.05182457	-4.270219778	0.00046764	probable protein, unknown function	
PFTT_1025800	250.2697167	323.990485	293.15172	307.89493	1860404	971.8573	98.4642191	288.8387811	1256.119729	2.120619797	0.000987771	conserved Plasmodium protein, unknown function	
PFTT_0610700	167.913839	474.414639	64.98306	125.013	1173.32	675.9908	605.657668	345.8089494	3.29878924	1.719787526	0.00111028	RNA binding protein, putative	
PFTT_0628300	213.6107716	189.345089	53.403608	49.400299	547.433	275.3596	261.690925	113.8141778	361.4944603	1.667293253	0.002646731	hydroxymethylglutathione synthetase	
PFTT_0833400	41.35.945656	2372.0732	5935.2987	6299.4455	649.691	1522.576	1868.602968	4685.691	1346.956749	0.28746171	-1.798585292	tryptophan/threonine-rich antigen	
PFTT_0721900	179.9316001	253.320236	299.0602	829.9202	23.8014	0	49.74290309	54.93997155	24.5147778	0.045437375	-4.462837162	erythrocyte binding like protein 1, pseudogene	
PFTT_0308600	494.6327568	48.3881894	117.0334	174.21167	7.05228	16.19762	19.46461425	133.5698525	142.3817034	0.106599855	0.001912211	conserved Plasmodium protein, unknown function	
PFTT_0600700	111.7082257	219.850686	92.036004	62.900769	998.779	32.3.9524	462.8252723	121.6261713	595.185411	4.893563652	2.290885465	conserved Plasmodium protein, unknown function	
PFTT_0419300	115.802849	328.198154	154.07509	117.35901	12.3415	0	6.48820451	178.8610856	6.276562377	0.035091828	0.00357598	erythrocyte membrane protein 1, PTEMPI	
PFTT_0701400	1759.918549	4412.79249	1593.2485	1538.6681	9155.62	5037.46	4872.641768	2326.156156	6355.239598	2.732077802	1.449998568	Plasmodium exported protein, unknown function	
PFTT_1035200	2653.584355	1073.2435	2003.6579	1776.7977	14492.4	11241.15	11189.99046	4289.32087	12307.85532	8.069418189	1.520758242	probable protein, unknown function	
PFTT_0805400	326.9007586	130.437728	142.71262	301.64428	29.9722	0	30.27828884	225.4238452	0.809092113	-3.488558462	0.00408547	adenylate cyclase beta (ACbeta)	
PFTT_0318500	389.9508	123.074308	104.30747	262.5273	9.69688	0	12.9764095	10.93497088	7.557762838	0.034338938	-4.863170736	conserved Plasmodium protein, unknown function	
PFTT_1137600	1125.991502	560.671846	899.9076	1587.2619	206.279	259.1619	233.575371	1043.4582	233.0054572	0.223301189	-2.162937159	subtilisin-like protease 2	
PFTT_0565600	1007.430011	284.017633	266.56354	681.32086	16.7492	48.59286	45.4174326	559.830105	36.91981709	0.06594791	0.004373912	myosin D	
PFTT_0107000	2014.174695	3660.6172	5703.278	5878.8372	495.422	1133.833	1704.235115	1111.163652	0.257557193	-1.957035262	0.004501212	conserved Plasmodium protein, unknown function	
PFTT_1127600	803.8880289	268.238876	204.29721	443.99779	13.223	0	36.76649359	430.1054749	16.66317014	0.038742055	-4.689855693	conserved Plasmodium protein, unknown function	
PFTT_0914200	133.8795543	103.087882	83.400528	214.94171	9.69688	16.19762	12.9764095	178.8274182	12.95696988	0.072455164	-3.786767677	conserved Plasmodium protein, unknown function	
PFTT_0316600	15247.36198	2419.40947	850.82173	638.77611	3773.58	2607.817	2515.260709	1308.435877	296.5642238	2.266555274	1.189501345	formate-nitrite transporter, putative	
PFTT_0370700	243.2909209	388.157432	179.07252	130.45712	808.367	615.5096	700.761131	235.2444979	798.2009372	3.010488884	1.58997773	HVA22/TP2/DPI family protein, putative	
PFTT_1149400	8029.285718	3744.82509	11552.223	15762.333	770.461	1878.924	128.131158	159.250441	0.162959224	-2.617417078	0.005599325	ring-infected erythrocyte surface antigen, pseudogene	
PFTT_1458300	246.7175536	45.48013	149.5301	130.45712	825.116	0	245.5462253	675.5756719	27.51317684	1.460122732	0.005743263	conserved Plasmodium protein, unknown function	
PFTT_1243900	746.3205997	256.66787	248.61084	643.21206	39.6691	48.59286	56.23110784	473.702802	48.16434057	0.10167628	-3.297944942	ubiquitin conjugating enzyme E2, putative	
PFTT_0556600	49.34351073	212.487266	109.9791	805.3549	524.513	194.3715	309.2710931	112.8910089	342.718159	3.035835354	1.602093355	rflin (RIF)	
PFTT_0919400	167.9050018	51.5439408	55.448852	137.11103	7.05228	48.59286	8.650939668	103.7526907	241.3202622	2.08073466	-2.264835096	thioredoxin, putative	
PFTT_1458400	282.3545336	196.708509	95.671995	69.362053	528.039	340.15	493.1035611	161.0242725	453.7642479	2.17986636	0.007287083	thioredoxin 1	
PFTT_1034800	914.9109281	29.2.43297	204.7517	481.90496	15.8676	32.39524	32.44102376	47.5.5001401	26.9012954	0.058613701	-4.137617302	merozoite surface protein 3	
PFTT_0202300	417.444184	3785.84986	2543.8297	1581.2128	8460.09	8658.542	7868.029628	3007.083562	8298.885791	27.59778908	1.464552694	erythrocyte membrane protein 1, PTEMPI	
PFTT_0722500	167.9050018	57.8554438	51.131114	138.1192	7.93381	0	8.650939668	103.7526907	5.528249923	0.053282955	-4.230182102	phosphoglycerate kinase	
PFTT_0723200	8486.398518	19475.1943	7766.0208	5184.4102	26185.1	29803.62	30241.52335	10228.00594	28743.41489	2.810262761	1.490706569	conserved Plasmodium protein, unknown function	
PFTT_0101000	759.341804	301.900225	434.27359	1432.6087	20.2753	178.1738	67.04478243	732.8030732	88.49796907	0.120893733	-3.048188637	histidine-rich protein III	
PFTT_1013300	292.6344317	92.5687101	68.174818	145.98292	7.05228	48.59286	6.48820451	149.8402211	15.31190754	0.102188234	-3.290699005	erythrocyte binding antigen-181	
PFTT_1134400	22.19.08733	3661.72363	3835.0608	3337.4439	899.165	1635.96	1427.405451	3263.328903	1320.843308	0.404753351	-1.304885971	conserved Plasmodium protein, unknown function	
PFTT_0703000	2101.211165	1721.98839	1421.445	1752.6016	428.426	1150.031	813.188238	1749.311536	797.2150579	0.45573075	-1.133746377	conserved Plasmodium membrane protein, unknown function	
PFTT_1334300	5543.23527	727.926675	793.78213	2336.3317	50.2475	64.79048	77.8545702	1850.466007	64.29880182	0.034747356	-4.84695297	merozoite surface protein 7	
PFTT_0903800.1	362.5377385	93.6206272	127.03241	377.45861	8.81534	0	32.44102376	240.1623471	13.75212277	0.057261777	-4.126283748	conserved Plasmodium protein, unknown function	
PFTT_0214000	403.6573308	114.65897	167.0283	514.16638	14.1046	0	30.27828884	299.8777459	14.79428004	0.049334371	-4.341263068	serine/threonine protein kinase, putative	
PFTT_1121900	1593.384201	1874.51638	2266.5855	3054.3499	571.234	97.18573	1064.065579	869.0523912	395.525595	-1.33815704	0.009668816	conserved Plasmodium protein, unknown function	
PFTT_1011700	364.5937181	69.4265325	73.628804	218.16785	3.52614	16.19762	43.2649834	181.4542264	0.044178688	0.041640952	0.009914754	conserved Plasmodium protein, unknown function	
PFTT_1035100	904.65103	225.8859	511.74677	17.6307	16.6302	32.39524	34.60375867	469.4732857	28.24089668	0.06088396	-4.05676978	diffusible binding-like necrotic surface protein	
PFTT_1011900	527.7014342	539.633503	486.08645	307.29002	2652.54	1150.031	93.64642191	465.1778535	1579.677499	3.985857061	1.763775734	pyoxanthine-guanine phosphoribosyltransferase	
PFTT_1013600	971.7930307	195.656592	268.38153	760.36134	16.7492	32.39524	49.74290309	549.0481236	32.96243333	0.0600356	-4.058037952	conserved Plasmodium protein, unknown function	
PFTT_0201000	0	0	0	4.40767	6.27193126	0	0	27.77868666	#DIV/0!	#DIV/0!	0.010765874	Dual protein, putative	
PFTT_0411100	252.8854925	240.88903	105.67097	108.47902	1218.28	437.3358	404.4314295	176.9811235	3.879976421	1.956047885	0.011173665	Plasmodium RNA of unknown function RUF6-14	
PFTT_0210700	2157.407941	2601.39114	3932.3235	5844.7611	722.858	955.6596	906.1859303	3633.970921	861.5679433	0.237088787	-2.076510398	liver stage antigen 3	
PFTT_1329000	17.40729406	47.3362722	62.039085	142.15188	7.05228	0	17.30180754	206.4000446	8.118051661	0.076297446	0.011448076	conserved Plasmodium protein, unknown function	
PFTT_1469500	416.678535	107.29555	113.1702	294.78872	14.1046	32.39524	19.46461923	13.98813593	0.094376466	-3.40542904	0.011619241	zinc finger protein, putative	
PFTT_1028100	142.439339	276.654213	313.83141	941.83182	24.683	80.98811	95.16033635	668.6891971	66.94380224	0.10011984	-3.32031341	merozoite TRAP-like protein	
PFTT_0808100	174.7582672	37.8690178	74.9923	246.19496	3.52614	16.19762	12.9764095	133.4536362	10.90005615	0.081676727	-3.613931143	conserved Plasmodium protein, unknown function	
PFTT_1326500	575.6742918	247.200633	298.1512	859.16194	52.8921	0	54.06837293	495.0466912	41.05268749	-3.592016913	0.013220897	conserved Plasmodium protein, unknown function	
PFTT_1251400	1309.659014	346.080746	514.94713	1780.2255	39.6691	97.18573	101.6485411	987.7280891	79.50110611	0.080488858	-3.6350671	coronin	
PFTT_1418400	278.2425744	51.5439408	87.036518	211.3123	11.4599	16.19762	2.162734917	157.0383832	9.94010132	0.063239106	-3.981671056	cyclic nucleotide-binding protein, putative	





Supp. Table 5. var genes expression profiles (GLM) of IT4\_Ctrl, IT4\_Wild and IT4\_P-selectin IEs populations

var genes (Nom)	IT4_Ctrl	IT4_Ctrl_2	IT4_Ctrl_3	IT4_Ctrl_4	IT4_Wild	IT4_Wild_2	IT4_Wild_3	IT4_P-select_1	IT4_P-select_2	IT4_P-select_3	IT4_P-select_4	X_Interrupt	chvTRIE	ncvTRIE	deviance	coverage	Pval
IT4_var0	4.262619613	8.677591278	15.43158278	12.19189343	5.657181975	0.102858949	685.6804734	47.22725581	720.7143647	6.475362202	4.76232	TR UE	6.475362202	4.76232	TR UE	0	0
IT4_var1	32.3000012	6.799760526	42.53238149	29.23154283	4282.4868299	3586.4668899	29.23154283	645.8691627	1009.180649	1.327025803	1.421531447	TR UE	1.327025803	1.421531447	TR UE	0	0
IT4_var2	185.4329532	79.8338976	95.15440066	83.78690955	2381.673569	5174.757926	7300.681241	622.3393388	8874.930874	6.071597922	228.9704539	TR UE	6.071597922	228.9704539	TR UE	0.0019578406	0.0019578406
IT4_var3	25891.15153	80992.43198	32.49544817	26099.02375	40099.51942	3773.474468	635.5852391	622.3393388	874.930874	6.071597922	228.9704539	TR UE	6.071597922	228.9704539	TR UE	0.0019578406	0.0019578406
IT4_var4	8.677591278	5.51129755	3.24939106	3.24939106	5.657181975	3.411922383	6.341922383	6.341922383	102.7417227	102.7417227	967.57068165	TR UE	102.7417227	967.57068165	TR UE	0.000604137	0.000604137
IT4_var5	223.612677	5756.714654	30.16207015	2751.505906	311.1450031	3048.490066	2855.301564	992.7790949	1194.443166	11.24536268	11.24536268	TR UE	11.24536268	11.24536268	TR UE	0.000604137	0.000604137
IT4_var6	3.19696471	20.85231907	16.16620006	6.175913062	42.32891324	230.5582515	26.67230682	87.85767868	58.68590274	6.341922383	6.341922383	TR UE	6.341922383	6.341922383	TR UE	0.000604137	0.000604137
IT4_var7	14.9191665	10.43110953	5.14869927	5.14869927	1329.471634	6886.197122	83.60173323	139.6984576	17.14308617	39.84932255	32.31319489	TR UE	32.31319489	32.31319489	TR UE	0.000604137	0.000604137
IT4_var8	51.9114536	16.83742343	125.2897554	89.53177949	83.4434326	333.2029754	487.796536	44.0345264	47.14907234	51.62643778	49.28316028	TR UE	49.28316028	49.28316028	TR UE	0.001125447	0.001125447
IT4_var9	150.2574413	463.383743	272.4963008	192.1819137	35.35738072	17.09761416	112.5883448	108.071389	141.071389	108.2932324	8.940601627	TR UE	8.940601627	8.940601627	TR UE	0.011199357	0.011199357
IT4_var10	41.50654123	32.97484666	27.92381646	21.9320628	30.77506494	30.77506494	118.5906585	190.8537424	190.8537424	190.8537424	190.8537424	TR UE	190.8537424	190.8537424	TR UE	0.002477017	0.002477017
IT4_var11	7.45984432	27.76829209	7.20667597	6.20667597	907.463867	333.2029754	27.01423067	14.1102223	15.01526338	18.40150257	3.861012629	TR UE	3.861012629	3.861012629	TR UE	0.002477017	0.002477017
IT4_var12	7.45984432	5.206554767	9.20303217	12.19189343	967.3781006	333.2029754	17.84147369	148.1126623	199.073361	326.626707	262.1032891	TR UE	262.1032891	262.1032891	TR UE	0.003317845	0.003317845
IT4_var13	6.46632169	36.44583837	39.31379423	87.57082848	257.4017583	1101.55738	13.61480292	87.66315988	159.3461329	154.8681605	159.3461329	TR UE	159.3461329	159.3461329	TR UE	0.00540946	0.00540946
IT4_var14	15.98482355	52.06554767	22.4125369	19.991951	5.657181975	3.41922383	46.23631588	60.9601677	52.68149776	52.68149776	52.68149776	TR UE	52.68149776	52.68149776	TR UE	0.006194148	0.006194148
IT4_var15	9.59089413	24.29725581	25.71930464	14.11866848	152.7431066	742.0107914	12.20176651	56.34406843	70.00922863	70.00922863	70.00922863	TR UE	70.00922863	70.00922863	TR UE	0.015234654	0.015234654
IT4_var16	9.59089413	19.09700881	10.2225911	6.16758387	11.31436375	0.102858949	6.341922383	6.341922383	45.492321762	45.492321762	45.492321762	TR UE	45.492321762	45.492321762	TR UE	0.018151649	0.018151649
IT4_var17	52.21709026	52.06554767	18.73835052	15.72105805	6.341922383	6.341922383	53.52402286	41.12790041	41.12790041	41.12790041	41.12790041	TR UE	41.12790041	41.12790041	TR UE	0.02574104	0.02574104
IT4_var18	5.21274517	5.206554767	62.4611684	5.45244629	5.657181975	0.102858949	6.341922383	6.341922383	45.24405932	44.74029789	36.31293877	TR UE	44.74029789	44.74029789	TR UE	0.01837236	0.01837236
IT4_var19	1.06565093	10.43110953	6.24611684	7.99272445	8.48572812	25.61761349	68.3045663	56.04397608	63.34168305	81.78445589	64.5419755	TR UE	64.5419755	64.5419755	TR UE	0.02272327	0.02272327
IT4_var20	36.2326671	154.611248	95.52884579	86.405403	8.48572812	25.61761349	68.3045663	56.04397608	32.86147466	22.40072537	17.83273288	TR UE	22.40072537	22.40072537	TR UE	0.03114482	0.03114482
IT4_var21	180.0952757	541.4816958	249.109836	186.727691	19.80013656	0.102858949	60.04711723	60.48416351	8.17485328	4.436257031	2.09930335	TR UE	2.09930335	2.09930335	TR UE	0.045349941	0.045349941
IT4_var22	13.8351374	19.09700881	15.43158278	15.43158278	18.3884109	0.102858949	68.3045663	66.03612314	3.900320148	2.15234232	1.8486016	TR UE	1.8486016	1.8486016	TR UE	0.064107028	0.064107028
IT4_var23	88.4453698	65.9493371	76.62758846	116.896822	66.47183703	25.61761349	48.4807079	132.0346579	163.057589	163.057589	135.252699	TR UE	135.252699	135.252699	TR UE	0.085597247	0.085597247
IT4_var24	6.3392462	50.3302941	36.0072459	34.18809159	42.4286406	20.57113699	58.01032622	86.67809259	69.5167675	5.775976648	0.82423435	TR UE	0.82423435	0.82423435	TR UE	0.142396763	0.142396763
IT4_var25	6.3064991	17.51518256	45.7457401	39.14222616	19.80013656	0.102858949	68.3045663	66.03612314	54.6934588	37.61108548	6.362526849	TR UE	6.362526849	6.362526849	TR UE	0.142396763	0.142396763
IT4_var26	30.9089922	192.6425264	115.7366709	88.35126274	12.72863922	25.61761349	23.9582602	40.488152676	40.38107509	35.98969726	7.259400759	TR UE	7.259400759	7.259400759	TR UE	0.142396763	0.142396763
IT4_var27	71.39087852	204.7911542	187.3835052	149.5104704	19.80013656	0.102858949	68.3045663	66.03612314	65.72294933	79.73984449	58.36167124	TR UE	58.36167124	58.36167124	TR UE	0.16065674	0.16065674
IT4_var28	21.3130897	41.62348467	32.12305842	20.5326284	41.04456389	30.77506494	72.56999332	65.92794002	47.57214498	40.0706295	6.229056835	TR UE	6.229056835	6.229056835	TR UE	0.16065674	0.16065674
IT4_var29	105.4998354	118.0152414	41.51830665	37.85887448	83.4434326	128.0880675	34.19523832	34.19523832	1.428759768	4.089222794	6.160398631	TR UE	6.160398631	6.160398631	TR UE	0.25657485	0.25657485
IT4_var30	127.878884	355.7812424	304.222632	196.9944825	96.1729187	25.61761349	54.7123663	51.509392436	4.38759768	4.38759768	4.38759768	TR UE	4.38759768	4.38759768	TR UE	0.29664788	0.29664788
IT4_var31	24.51006278	46.8589929	48.49926017	35.29271718	25.6271844	0.102858949	68.3045663	66.03612314	53.3403467	43.98148074	38.9778083	TR UE	38.9778083	38.9778083	TR UE	0.342482531	0.342482531
IT4_var32	12.7878884	60.74313895	40.4168014	27.91289898	22.6287275	51.2352609	13.67809133	10.02933792	5.277429096	0.66701593	0.66701593	TR UE	0.66701593	0.66701593	TR UE	0.342482531	0.342482531
IT4_var33	620.211157	1530.271101	605.873335	425.751919	79.20054625	0.102858949	68.3045663	66.03612314	89.0535887	90.5697142	73.6005206	TR UE	73.6005206	73.6005206	TR UE	0.380161961	0.380161961
IT4_var34	62.8736933	53.80106592	32.1153832	37.21719864	56.57181975	25.61761349	17.09761416	85.8459393	63.89410616	63.89410616	63.89410616	TR UE	63.89410616	63.89410616	TR UE	0.380161961	0.380161961
IT4_var35	1340.593868	4557.470939	3303.82839	9.945975208	63.64329609	76.88284048	37.61475115	20.51690038	10.95382489	14.31229798	16.53880685	TR UE	16.53880685	16.53880685	TR UE	0.380161961	0.380161961
IT4_var36	152.3886512	231.487454	116.891268	102.248375	49.5004414	51.2352609	34.19523832	57.04476137	27.14643559	42.93683352	5.631583352	TR UE	5.631583352	5.631583352	TR UE	0.380161961	0.380161961
IT4_var37	42.62619612	12.14862779	10.2877185	12.5167885	12.72863922	25.61761349	23.9582602	36.02827034	4.269220549	0.006133864	0.006133864	TR UE	0.006133864	0.006133864	TR UE	0.404257404	0.404257404
IT4_var38	14.9116865	12.14862779	11.8997717	25.762459	24.04302397	25.61761349	23.9582602	36.02827034	32.09891918	42.0560214	42.0560214	TR UE	42.0560214	42.0560214	TR UE	0.404257404	0.404257404
IT4_var39	26.44137258	42.8018134	26.6101937	25.762459	8.80013656	0.102858949	68.3045663	66.03612314	42.4556849	38.8046267	39.88047535	TR UE	39.88047535	39.88047535	TR UE	0.414837375	0.414837375
IT4_var40	9.20924162	19.09700881	23.8821145	13.7964363	19.80013656	0.102858949	68.3045663	66.03612314	32.3249885	32.3249885	15.9666579	TR UE	15.9666579	15.9666579	TR UE	0.414837375	0.414837375
IT4_var41	1.06565093	17.51518256	13.9793188	16.0418997	8.48572812	25.61761349	34.19523832	18.5142781	3.070416862	0.15319155	0.15319155	TR UE	0.15319155	0.15319155	TR UE	0.414837375	0.414837375
IT4_var42	1.06565093	17.51518256	13.9793188	16.0418997	8.48572812	25.61761349	34.19523832	18.514278									

Supp. table 6A. Part of whole transcriptome profiles (GLM) of IT4\_Ctrl, IT4\_Wild and IT4\_P-selectin IEs populations

	IT4_Ctrl	IT4_Ch2	IT4_CtrB	IT4_CtrH	IT4_Wild	IT4_Wild2	IT4_Wild3	IT4_Pack_1	IT4_Pack_2	IT4_Pack_3	IT4_Pack_4	X_Interept.	cbvTRUE	recTRUE	deviance	covered	p	name
PTT1_017600	289.883707	13.884146	38.2115832	122.88923	2.8285904	0	10.2585689	598.399259	417.197823	495.307111	556.381268	6.1364758	-3.78027574	6.59104127	14.506	TRUE	0	unspecified product
PTT1_017610	289.883707	13.884146	38.2115832	122.88923	2.8285904	0	10.2585689	598.399259	417.197823	495.307111	556.381268	6.1364758	-3.78027574	6.59104127	14.506	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_018100	606.35714	203.055636	166.645128	477.30971	15.557502	0	6.83904563	959.753049	835.348211	1348.932369	1280.390221	8.44732108	-1.93871473	4.89926608	10.606	TRUE	0	erythrocyte membrane protein 1 (PEM1P), psudogene
PTT1_040150	31.906947	3.47103651	6.63811028	6.41675839	2.8285904	0	6.83904563	592.4890165	592.4890165	592.4890165	592.4890165	6.83904563	-0.55939828	6.72636737	2.8656	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_040630	669.231793	173.899387	121.6142715	257.623849	63.6432961	128.0880675	12.8.0880675	894.915628	874.9274373	67.49747375	6087.440542	8.20287437	-2.156368953	6.74808858	25.1192	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_042150	395.357961	18.9983215	11.6142715	215.630382	9.9000628	25.61761349	31.29284237	1949.780831	1949.780831	1386.72768	67.49747375	8.20287437	-4.51113497	4.73375545	3.9908	TRUE	0	nucleotide binding protein, putative
PTT1_052200	20.34744316	6.94207302	9.92030217	15.721058	9.9000628	25.61761349	10.50419105	1048.582511	1048.582511	1050.419105	888.3942398	3.74782801	-0.771779801	6.59861041	24.5	TRUE	0	nucleotide binding protein, putative
PTT1_052400	20.34744316	6.94207302	9.92030217	15.721058	9.9000628	25.61761349	10.50419105	1048.582511	1048.582511	1050.419105	888.3942398	3.74782801	-0.771779801	6.59861041	24.5	TRUE	0	nucleotide binding protein, putative
PTT1_053400	409.21160278	8.6759128	8.65062666	12.918409	1.41429547	0	10.70794743	1098.464743	1098.464743	1098.464743	1170.151508	3.721797959	-3.20110322	6.7043989601	34.9975	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_053500	312.248887	85.040345	47.3418676	67.0551251	15.557502	25.61761349	6.83904563	642.0037167	635.614937	59.61042223	6095.158402	5.90358363	0.091788	6.62163599	19.0481	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_070410	197.079208	69.4207302	41.5783063	52.4216084	107.486456	0	16.64433328	1544.332648	1544.332648	1646.433328	1544.332648	7.55687573	-2.3957118	5.59871014	28.5245	TRUE	0	unspecified product
PTT1_072700	330.089289	55.783584	70.9117927	98.4972412	45.257455	0	3.90752952	3063.269244	3063.269244	1740.911428	1948.455262	4.02942686	0.370191494	4.80843812	6.97572	TRUE	0	nuclear pore-associated assembly protein, putative
PTT1_083200	403.8832084	468.580929	300.187027	171.327449	217.801502	281.793784	15.0490046	4294.87006	4294.87006	14030.12341	1748.455262	4.02942686	0.370191494	4.80843812	6.97572	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_099200	34.10095691	3.47103651	6.63811028	6.41675839	2.8285904	0	6.83904563	592.4890165	592.4890165	592.4890165	592.4890165	6.83904563	-0.55939828	6.72636737	2.8656	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_091500	34.10095691	3.47103651	6.63811028	6.41675839	2.8285904	0	6.83904563	592.4890165	592.4890165	592.4890165	592.4890165	6.83904563	-0.55939828	6.72636737	2.8656	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_092600	155.586159	190.397068	152.437954	131.545347	152.743911	128.0880675	164.170959	313.139756	285.102674	387.297392	7.22740828	4.7324065	0.029315404	4.37424065	1.90627	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_093100	802.4381422	55.368133	39.2403105	86.2739711	59.661279	76.8524889	150.590046	4216.39985	4216.39985	1282.33561	6.02321874	0.0149114	-0.739517935	3.093003486	20.6471	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_093500	64.9181504	55.368133	39.2403105	86.2739711	59.661279	76.8524889	150.590046	4216.39985	4216.39985	1282.33561	6.02321874	0.0149114	-0.739517935	3.093003486	20.6471	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_094800	17.0487845	0.29349401	0.92959247	6.09592947	4.24288641	0	10.2585689	479.372975	479.372975	573.752975	588.480422	2.684418618	-1.23289796	7.831927261	14.857	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_100200	451.87679	149.25457	84.7536827	194.788617	7.07147734	0	10.2585689	479.372975	479.372975	573.752975	588.480422	2.684418618	-1.23289796	7.831927261	14.857	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_100300	230.0748936	444.292673	84.7536827	194.788617	7.07147734	0	10.2585689	479.372975	479.372975	573.752975	588.480422	2.684418618	-1.23289796	7.831927261	14.857	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_101300	151.079863	322.806396	43.5921411	1209.87979	26.8716139	102.470454	235.9470524	1729.275573	1729.275573	2173.93368	1940.525669	9.76097493	-4.112586562	5.200498841	14.6669	TRUE	0	membrane skeleton protein (MSC1-related)
PTT1_101600	8.5252327	78.9083215	54.7453701	101.70562	5.65718187	0	6.83904563	6421.038403	6421.038403	7654.255516	574.741589	7.45801537	-5.01839253	4.542821056	3.9063	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_102800	190.085207	149.049015	189.588017	433.72867	35.373867	0	13.6780913	898.2136468	898.2136468	1279.415582	1374.741589	7.45801537	-5.01839253	4.542821056	3.9063	TRUE	0	calcium/calmodulin-dependent protein kinase c, putative
PTT1_105700	587.698768	209.997709	139.145402	351.317522	175.372638	51.23522699	8.02533332	814.348331	802.579806	4917.57806	4225.702229	8.67024904	-0.850025896	5.02518017	6.43879	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_110800	181.1613336	3.47103651	6.63811028	6.41675839	2.8285904	0	6.83904563	592.4890165	592.4890165	592.4890165	592.4890165	6.83904563	-0.55939828	6.72636737	2.8656	TRUE	0	conserved Phasmodium protein, unknown function
PTT1_112400	35.69943926	171.1562485	30.9158642	36.6471876	21.214432	0	34.9522382	1965.542304	1745.493248	1470.30448	1289.662342	7.461208638	-2.856093597	9.364546685	72.7954	TRUE	0	unspecified product
PTT1_113600	39.4292132	24.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_1148700	69.2676872	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701	83.7386969	8.34434327	102.470454	58.13188814	2905.701553	2899.389853	2660.039428	2761.1573141	5.71176181	-1.80360009	5.321348053	7.32225	TRUE	0	unspecified product
PTT1_120350	65.0049491	34.2972556	54.7453701															



Supp. Table 7. *var* genes expression profiles (GLM) of IT4\_Wild and IT4\_E-selectin IEs populations

<i>var</i> genes (Nom ID)	IT4_Ctrl	IT4_Ctrl2	IT4_Ctrl3	IT4_Ctrl4	IT4_Wild	IT4_Wild2	IT4_Wild3	IT4_E-selectin	IT4_E-selectin_1	IT4_E-selectin_2	IT4_E-selectin_3	IT4_E-selectin_4	X-Intercept	abs(T <sub>1</sub> -T <sub>2</sub> )	dev(T <sub>1</sub> -T <sub>2</sub> )	convergence	p		
PFTT_13000100	5.87202245	3.770931	2.2153526	3.96641818	0.0	0	0	55.3009449	90.48877554	66.17072692	88.41260379	62.17072692	1.52474748	0.82926	4.62309	5.16127	TR UE	0.00039	0.02717
PFTT_13011500	36.38037725	11.5564185	85.725832	61.8083377	58.5046681	23.33860284	21.07586418	34.83188679	34.83188679	34.83188679	34.83188679	34.83188679	6.20485622	4.45328	-2.4469	3.98755	TR UE	0.00258	0.17556
PFTT_07110000	131.8768625	54.02260263	40.47466	87.823303	166.948108	36.34229057	7.121983724	5984.32741273	10580.011167	64.523671177	66.24107776	66.24107776	6.14370087	5.61952	1.18635	1.18635	TR UE	0.00894	0.89903
PFTT_12400000	3.03169804	5.87202245	10.558607	5.87202245	3.96641818	0	0	36.86718369	23.29411333	23.29411333	23.29411333	23.29411333	2.815400238	-0.5966	3.18039	2.78231	TR UE	0.05504	1
PFTT_12400000	75.03452388	79.85949953	28.40768	26.1411607	58.5046681	89.95616477	24.04207833	16.38541498	17.0236238	10.24467259	25.70133925	25.70133925	5.70117524	0.03987	-1.633	3.94475	TR UE	0.06053	1
PFTT_09001000	37.13830178	35.23213115	12.821165	10.85522278	7.92834636	4.788145667	4.788145667	23.55403403	18.81448447	28.127384961	92.52489123	92.52489123	4.5746668	-2.9731	3.2228	2.12263	TR UE	0.07834	1
PFTT_06016500	21.97981176	130.35888889	79.1899551	81.4373119	8.92403409	17.99123295	7.12126235	3.072265308	3.072265308	3.072265308	3.072265308	3.072265308	6.19046153	-2.0723	1.88084	6.5205	TR UE	0.09209	1
PFTT_06076000	10.01094336	7.04621243	3.5195356	5.54456417	0	0	0	3.072265308	3.072265308	3.072265308	3.072265308	3.072265308	6.209911224	-3.2989	32.8028	40.0734	TR UE	0.10079	1
PFTT_06011000	25.76943389	104.521692	6.5362804	5.9529851	5.94962727	0	0	10.24088436	17.91855663	5.634509922	21.58912497	21.58912497	5.99361212	-4.3234	1.60982	7.08221	TR UE	0.19453	1
PFTT_14000200	6.821320755	12.91844845	7.541862	4.43070521	7.93283636	9.616831333	9.616831333	16.38541498	20.6034013	17.4139434	34.95381238	34.95381238	4.37499684	-1.1375	1.48178	1.65006	TR UE	0.24029	1
PFTT_04193000	128.0892449	366.4141743	170.44608	128.933522	138.246336	0	0	18.43391748	21.0015788	28.784580996	4.61312095	4.61312095	2.12694	2.85569	TR UE	0.2411	1		
PFTT_11510000	59.52994562	22.31368369	10.10605	14.6213272	7.93283636	7.2126235	7.2126235	18.43391748	21.0015788	28.784580996	4.61312095	4.61312095	1.13563	2.72245	TR UE	0.28978	1		
PFTT_12411000	50.78094325	138.5797198	128.21165	103.235431	138.246336	2.404207833	2.404207833	15.36132654	16.1706181	15.87924251	18.50496426	18.50496426	6.71681468	-3.7469	0.95954	3.78069	TR UE	0.3066	1
PFTT_06097000	22.97269259	460.1316458	2910.11532	2018.40776	3002.527856	25.18772614	27.6243488	19.97995639	19.97974716	14.04432378	32.73325827	32.73325827	11.52930602	-0.4096	-0.3645	7.66411	TR UE	0.31925	1
PFTT_06004000	1588.609987	3895.499411	2063.9566	1899.386639	2181.53	2140.95722	2007.513541	17.59262653	14.29408492	10.91569864	6.168321451	6.168321451	3.68511188	0.54408	-1.4159	3.48383	TR UE	0.32751	1
PFTT_05368000	62.90775566	44.62736739	53.798616	76.42966346	46.6054136	17.99123295	60.10519583	64.51757147	66.29865985	64.02930366	96.63703559	96.63703559	5.89647627	-0.3862	0.67611	1.57554	TR UE	0.36681	1
PFTT_08110900	1841.453429	541.975888	2232.36929	14230.3175	28114.9637	26.50108614	25.63125971	18.90467253	17.950440257	11.35938052	29.952739389	29.952739389	14.7349784	-0.0278	-0.4866	6.28792	TR UE	0.37418	1
PFTT_05001000	45.47547156	34.05772774	24.636749	28.5780486	29.7481363	0	0	27.65038777	30.46154628	36.36858768	38.03798209	38.03798209	5.04936383	-0.8687	0.86998	2.3205	TR UE	0.39599	1
PFTT_04110000	53.05471682	32.88332334	32.430007	38.7686076	49.890272	89.95616477	129.8327233	118.7942586	123.6380408	142.4000498	135.7030713	135.7030713	5.29492532	-0.4299	1.18102	0.54828	TR UE	0.40012	1
PFTT_06091000	9.095094313	41.10415417	27.653494	19.2735694	15.8656727	35.98246591	9.616831333	81.92707488	53.7556049	17.17127081	13.36460941	13.36460941	4.97553447	-0.4299	-1.0804	1.92778	TR UE	0.40288	1
PFTT_07109000	0.757924526	7.04621243	4.2737218	5.09531099	5.94962727	17.99123295	4.808415667	24.57812246	6.271494822	9.732438956	40.09409823	40.09409823	2.09514211	0.8676	1.3673	2.69909	TR UE	0.41071	1
PFTT_06002000	5.305471682	18.79047048	11.815584	4.43070521	35.6977636	233.8660284	189.9324188	153.6132654	60.1358236	180.8184711	180.9374283	180.9374283	3.27717694	3.89253	0.50186	10.2256	TR UE	0.42299	1
PFTT_06001000	5.305471682	3.525213215	6.7876758	8.4183399	678.257389	37.24185222	5051.240658	3062.024424	39.17566933	44.79348038	3749.31157	3749.31157	2.64055583	8.9795	0.53212	6.72456	TR UE	0.42991	1
PFTT_05376000	18.94811315	56.37141144	27.653494	19.7166382	31.7313454	0	0	31.74674152	26.87873495	28.68508345	19.06603566	19.06603566	4.93003443	-0.7623	0.8129	3.29412	TR UE	0.43671	1
PFTT_06098000	0	0	0	0	0	0	0	2.687873495	3.585635405	1.02805357	1.02805357	1.02805357	7.402101	2.89112	32.6832	3.27878	TR UE	0.45172	1
PFTT_06002000	6.063396209	15.26725726	26.899308	31.9319908	8.9244409	17.99123295	36.06631175	46.08397962	15.20226796	17.92817702	62.71126778	62.71126778	1.14400874	0.80593	3.67253	TR UE	0.48233	1	
PFTT_06010000	3.78962263	3.523213215	4.2737218	3.76690493	3.96641818	0	0	5.12044218	6.271494822	3.585635405	11.2160786	11.2160786	1.14400874	-0.0949	1.86606	1.16696	TR UE	0.48736	1
PFTT_04112500	11.36868789	35.23213215	15.335119	13.535119	16.6341818	0	0	8.192707488	3.192707488	6.271494822	3.0758242131	3.0758242131	4.23571388	-2.7304	1.23048	1.09495	TR UE	0.49402	1
PFTT_08100000	6.821320755	16.44166167	17.597678	9.74755146	9.99693491	521.7457557	858.2021965	424.9967009	1209.502573	691.5153995	512.9987315	512.9987315	3.65944161	5.29029	0.52152	0.93775	TR UE	0.51516	1
PFTT_06090000	26.52735841	12.91844845	17.849073	16.6151445	932.108272	46.95711801	58.87828152	394.37528152	664.5096728	4987.106615	4.27356284	4.27356284	2.11455804	7.69302	0.37364	7.27139	TR UE	0.5182	1
PFTT_06042000	1.515849052	8.720220225	5.2793034	4.65224047	1.98320099	0	0	6.144530616	4.479639159	6.65907182	13.36460941	13.36460941	2.11455804	-0.6605	1.48211	0.77673	TR UE	0.5366	1
PFTT_07108000	953.4690538	3083.985967	2260.5474	1475.42483	44.6222045	53.97369886	24.64628617	17.40950341	33.14932977	26.12391509	31.869696067	31.869696067	10.9242467	-5.6001	-0.5617	1.93666	TR UE	0.5438	1
PFTT_06049000	62.14981114	14.09235826	11.061398	19.49680464	36.6893682	161.92109966	187.528211	65.54166599	146.0362366	100.9718624	49.34657186	49.34657186	5.06596647	6.52603	-0.4994	3.8213	TR UE	0.56883	1
PFTT_06083000	2.273773578	313.5659761	165.929969	12.32699621	24.7901136	12.02103919	16.192103919	15.36132654	17.59262653	17.59262653	28.784580996	28.784580996	3.4193402	-3.5012	-0.3798	7.56485	TR UE	0.58664	1
PFTT_06085000	11.36868789	0.7541862	2.2153526	6.94123181	0	0	0	4.096352654	3.433484353	3.585635405	6.252482131	6.252482131	1.83599782	0.10375	1.02479	3.18352	TR UE	0.63412	1
PFTT_06108000	9.853018839	12.91844845	14.589033	10.5636977	12.8908591	4.808415667	4.808415667	5.12044218	9.732438956	16.448818747	3.45639186	3.45639186	3.45639186	-0.5929	0.73037	3.21778	TR UE	0.6549	1
PFTT_06001000	8.853018839	14.09235826	14.589033	10.5636977	32.72295	35.98246591	36.06416872	29.56561845	22.02604606	80.18817874	3.45639186	3.45639186	4.06638	2.17877	TR UE	0.65775	1		
PFTT_06103000	0	0	0.2513954	0.22153526	0	0	0	0	0	0	2.0561074	2.0561074	-3.0292784	-27.339	29.4024	0.48689	TR UE	0.66924	1
PFTT_08335000	44.71754704	36.40653655	26.145122	25.6989092	39.664818	17.99123295	12.02103919	16.38541498	26.87783495	8.70797168	22.61717854	22.61717854	6.595132829	-0.4384	-0.3971	1.88166	TR UE	0.7062	1
PFTT_06107000	32.59075462	30.53451453	46.759545	26.362696	39.664818	17.99123295	9.616831333	61.44850616	27.77376278	10.24467259	27.7544639	27.7544639	5.09150065	-0.5314	-0.3955	3.06329	TR UE	0.70865	1
PFTT_06010000	17.4322641	31.70891893	33.184193	24.3688786	1.98320099	0	0	18.43391748	19.7104123	23.50274695	25.70133925	25.70133925	4.73735928	-0.6852	0.398	1.82437	TR UE	0.72701	1
PFTT_06105000	210.7030182	940.6979283	141.03282	81.0819053	43.6306	17.99123295	26.44428617	14.372218	30.46154628	10.75690621	20.5610714	20.5610714	-36.144796	2.89112	31.8771	0.13616	TR UE	0.72951	1
PFTT_06092000	9.853018839	11.74404405	4.5251172	5.75991677	9.91604545	0	0	4.808415667	5.37556699	10.24467259	5.142026785	5.142026785	8.42316966	-3.5168	-0.6571	1.48632	TR UE	0.73696	1
PFTT_07106000	139.4581128	205.5207709	86.982809	78.4232887	31.7313454	17.99123295	31.25470183	20.48176872	30.46154628	21.0015788	26.72939282	26.72939282	6.99885231	-2.1288	-0.2392	2.0651	TR UE	0.74608	1
PFTT_06003000	10.61094336	8.220830834	7.7932574	8.4183399	16.8572773														

Supp. table 8. Part of whole transcriptome profiles (GLM) of IT4\_Ctrl, IT4\_Wild and IT\_E-selectin IEs populations

IT4_Ctrl	IT4_Ctr2	IT4_Ctr3	IT4_Ctr4	IT4_Wild	IT4_Wild2	IT4_Wild3	IT4_E-select-1	IT4_E-select-2	IT4_E-select-3	IT4_E-select-4	X.Interept.	chrTRIE	recTRIE	deviance	convged	P-value	Padj	name
PTT_111800	2.3488881	205.929536	251.664658	14.8740837	116.220542	1177.11288	6.9324771	6.1196108	57.5124453	TRIE	0	6.1196108	57.5124453	TRIE	0	0	conserved Plasmodium protein, unknown function	
PTT_123000	314.323609	43.7578932	83.3306515	183.320544	264.627833	185.306515	5.9804441	5.9804441	210.696804	6.1091694	0	-0.8150736	1.7113719	TRIE	0	0	osimipalic body protein	
PTT_114870	49.2426199	16.4416617	30.0527115	21.8159953	7.2126235	61.1331066	526.941387	538.357449	5.8329723	6.0181906	TRIE	4.58299723	6.0181906	TRIE	1.85178E-16	5.0377E-12	conserved Plasmodium protein, unknown function	
PTT_113400	253.9047162	48.458306	37.92105139	22.8192993	6.51320243	65.1302453	173.221241	66.832247	1.818559217	4.53191242	TRIE	2.011502472	4.53191242	TRIE	1.08574E-16	5.95279E-11	conserved Plasmodium protein, unknown function	
PTT_103500	61.5460361	67.6456932	68.6768761	295.912994	194.8183159	324.7323759	401.833879	269.111868	8.92918866	2.19716564	TRIE	-2.6693248	3.209746628	TRIE	3.07536E-08	1.20889E-07	ADAP1 transporter on adobe-like translocase	
PTT_133500	248.792444	43.4529638	34.7061907	33.7145542	0.61485785	92.8000346	389.182632	1370.12698	1.6177388	6.92928863	TRIE	5.98723793	10.9947206	TRIE	2.1444E-08	2.9898E-07	conserved Plasmodium protein, unknown function	
PTT_104200	104.503586	92.7794799	57.5991671	33.7145542	0.61485785	453.134829	6.1177388	6.1177388	4.1177388	3.94806655	TRIE	4.6169781	7.56115163	TRIE	3.40E-10	1.04E-06	conserved Plasmodium protein, unknown function	
PTT_103800	60.6390269	1.1744045	13.7538615	11.8992584	0	20.110427	249.968651	27.7283459	3.94806655	3.94806655	TRIE	5.27900694	11.7412983	TRIE	4.03577E-10	2.3689E-06	conserved Plasmodium protein, unknown function, pseudogene	
PTT_093800	142.829242	4.20916948	2.97481864	2.97481864	23.8371732	23.8371732	23.8371732	23.8371732	23.8371732	23.8371732	TRIE	2.87874658	11.7412983	TRIE	4.9308E-06	2.3689E-06	conserved Plasmodium protein, unknown function	
PTT_102000	170.531054	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_102100	191.524051	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251	46.715335	27.740034	21.577729	82.424043	98.5178125	9.8130799	17.1027268	1.3830799	40.6282891	TRIE	-2.56430801	10.8507521	TRIE	1.9797E-09	1.1152E-05	6-systeme protein	
PTT_079400	43.9596251																	

Supp. table 9. Part of whole transcriptome profiles (GLM) of IT4\_Ctrl, IT4\_Wild and IT4\_CD151 IEs populations (First experiment)

IT4_Ctrl	IT4_Ctrl_2	IT4_Ctrl_3	IT4_Wild_1	IT4_Wild_2	IT4_Wild_3	IT4_CD151_1	IT4_CD151_2	IT4_CD151_3	IT4_CD151_4	X.intercept	ch2TRUE	recTRUE	de-variant	converted	P-value	padj	name	
PPT1_027800	957.150102	1090.422902	1838.8859	51.8628785	35.910723	195.115377	945.87793	106.361004	9012.23043	47.011646	10.676979	11.1394745	0.21602806	0.41425E+3		2.468E-09	18S ribosomal RNA	
PPT1_027800	971.532900	1035.51569	1834.051452	51.3018569	36.5012019	1210.02854	1027.02563	100.852150	8921.67602	465.394845	60.0095512	11.1311072	-0.10936942	1.0077E-2		5.98543E-09	18S ribosomal RNA	
PPT1_011100	252.529700	157.112835	139.534808	47.245942	20.8975907	30.9759971	1913.32317	817.674291	8.549005954	2.81961379	0.48067139	0.09271631	2.2651528	TRIE		2.26515E-09	28S ribosomal RNA	
PPT1_016000	465.427204	343.148181	494.818721	146.238098	158.290975	210.09197	2029.58692	232.932892	2.87576457	2.59799495	2.15299459	-2.52494545	3.851212196	TRIE		1.281E-08	17-28E-04 ADP/ATP transporter on atypical translocase	
PPT1_040200	175.282152	198.5181391	154.871436	176.646036	104.3001468	104.3001468	994.5730045	132.742175	1.76463667	1.774714926	2.64685778	TRIE		1.738E-08		9.705991E-06	6-cysteine protein	
PPT1_040200	219.548166	378.650927	161.321458	1406.488985	12.9222137	940.097785	9147.406763	1100.230288	60.0738376	99.5730045	1.07283903	3.35690419	9.24069902	TRIE		2.717E-08	0.00151682 probable protein, unknown function	
PPT1_040200	40.8236837	186.169367	249.1108834	90.4389046	291.4583386	474.083321	307.079184	361.487008	352.3410573	8.16256162	1.71442439	2.80334832	7.36571816	TRIE		3.98319E-07	0.00522157 erythrocyte binding like protein 1, pseudogene	
PPT1_031000	201.296384	316.5743908	144.177291	103.194634	680.049026	514.702765	572.834755	93.6409756	57.4232136	122.7481965	1.58265546	1.64872113	2.03339639	TRIE		4.94762E-07	0.02759783 HVJ-217B2DP1 family protein, putative	
PPT1_114900	6645.347765	362.4213635	9.81107972	12469.16908	648.175274	1571.197901	1739.07384	11252.51188	7146.279262	7409356998	76422.5022	12.9470615	-2.85088813	2.669498121		5.4212E-07	0.0025191 ring-infected erythrocyte surface antigen, pseudogene	
PPT1_062300	134.853621	154.426521	42.9701629	39.0760452	461.545883	230.27613	2139.27299	7.810426297	18.338728	27.02712583	6.53063706	1.71887826	-3.746283089	5.4293E-07		0.00303498 RNA binding protein, putative		
PPT1_041700	303.929883	409.23011	82.5176708	85.3109813	482.1625842	352.1650407	371.2790106	31.2162519	61.8239457	57.43264359	55.4008938	7.77812446	8.840666319	TRIE		5.712E-07	0.00318471 gh1amp4-RNA (Chi) and/oramidease subunit A, putative	
PPT1_041700	74.8715597	244.307474	494.008723	495.764064	459.656452	2072.35352	2008.442648	46.82043778	107.336128	115.914415	124.69005	10.0915942	4.0514883	-4.857082914		7.08732E-07	0.03950474 Phasmodium exported protein (PHSTb), unknown function	
PPT1_040700	1718.62497	336.1350848	2118.014276	1453.178637	2245.63235	1896.273328	2031.426386	998.834066	417.208856	332.2084217	337.516889	11.0783093	-0.16982921	-0.14144171		7.8933E-07	0.004398407 erythrocyte membrane protein 1, PEMP1 (VAR)	
PPT1_040700	18.6316975	386.924254	133.833354	88.38824001	98.7092546	514.7925795	495.8386808	62.4272507	83.6933822	33.46881016	4.4528426	1.76825511	-3.9999581	1.7111E-06		0.00545303 dihydropyrene synthase		
PPT1_062300	1805.997135	47597.90889	1138.17945	90.0424812	637.25996	4334.339136	4122.96213	749.1279045	749.5740639	692.590094	749.5740639	1.91575107	3.62676946	TRIE		4.28909E-06	0.025701824 Phasmodium exported protein, unknown function	
PPT1_071000	198.461237	66.918139	105.0224994	74.6193049	74.6193049	10.14423621	21.0109197	156.2924991	147.523618	375.4150468	8.29492473	-3.40900223	5.92572638	TRIE		4.8326E-06	0.01375498 Phasmodium exported protein, unknown function	
PPT1_114900	416.315765	314.8585404	297.4396281	260.4588444	581.429635	419.8890941	410.1749069	124.854907	115.76710	84.8976822	8.200239784	0.685784735	-2.78477957	1.51939793	TRIE		2.8066E-06	0.01589694 translation machinery-associated protein 7, putative (TMA7)
PPT1_040700	5145.259723	802.160417	570.4800715	506.760772	1247.403671	150.473853	1147.428942	359.2926307	316.4425983	316.4425983	9.255972911	1.063973105	-1.951304935	1.49105485	TRIE		3.15809E-06	0.01758756 conserved Phasmodium membrane protein, unknown function
PPT1_040700	10.5827509	529.6380815	16.9451156	333.919146	1676.801248	900.965133	4655.131895	20976.75557	14611.18974	1704.147084	17665.43835	13.80151666	-2.607122472	9.240670529	TRIE		3.6915E-06	0.02650604 ring-infected erythrocyte surface antigen
PPT1_040700	204.1315444	334.590185	106.8142808	87.4044439	56.2144786	257.351303	298.7912038	31.2162519	41.2459638	50.6758693	44.8195257	1.319426493	4.84239423	TRIE		3.99969E-06	0.02176502 Cfpb protein, putative	
PPT1_114900	457.181742	909.400889	67.0245758	516.7108026	1539.625398	1137.763997	1324.228291	354.103294	354.103294	415.5201696	375.4150468	1.11211479	-1.91575107	1.62676946	TRIE		4.28909E-06	0.025701824 Phasmodium exported protein, unknown function
PPT1_071000	198.461237	66.918139	105.0224994	74.6193049	74.6193049	10.14423621	21.0109197	156.2924991	147.523618	375.4150468	8.29492473	-3.40900223	5.92572638	TRIE		4.8326E-06	0.01375498 Phasmodium exported protein, putative	
PPT1_114900	416.315765	314.8585404	297.4396281	260.4588444	581.429635	419.8890941	410.1749069	124.854907	115.76710	84.8976822	8.200239784	0.685784735	-2.78477957	1.51939793	TRIE		2.8066E-06	0.01589694 translation machinery-associated protein 7, putative (TMA7)
PPT1_040700	5145.259723	802.160417	570.4800715	506.760772	1247.403671	150.473853	1147.428942	359.2926307	316.4425983	316.4425983	9.255972911	1.063973105	-1.951304935	1.49105485	TRIE		3.15809E-06	0.01758756 conserved Phasmodium membrane protein, unknown function
PPT1_040700	10.5827509	529.6380815	16.9451156	333.919146	1676.801248	900.965133	4655.131895	20976.75557	14611.18974	1704.147084	17665.43835	13.80151666	-2.607122472	9.240670529	TRIE		3.6915E-06	0.02650604 ring-infected erythrocyte surface antigen
PPT1_040700	204.1315444	334.590185	106.8142808	87.4044439	56.2144786	257.351303	298.7912038	31.2162519	41.2459638	50.6758693	44.8195257	1.319426493	4.84239423	TRIE		3.99969E-06	0.02176502 Cfpb protein, putative	
PPT1_114900	457.181742	909.400889	67.0245758	516.7108026	1539.625398	1137.763997	1324.228291	354.103294	354.103294	415.5201696	375.4150468	1.11211479	-1.91575107	1.62676946	TRIE		4.28909E-06	0.025701824 Phasmodium exported protein, unknown function
PPT1_071000	198.461237	66.918139	105.0224994	74.6193049	74.6193049	10.14423621	21.0109197	156.2924991	147.523618	375.4150468	8.29492473	-3.40900223	5.92572638	TRIE		4.8326E-06	0.01375498 Phasmodium exported protein, putative	
PPT1_114900	416.315765	314.8585404	297.4396281	260.4588444	581.429635	419.8890941	410.1749069	124.854907	115.76710	84.8976822	8.200239784	0.685784735	-2.78477957	1.51939793	TRIE		2.8066E-06	0.01589694 translation machinery-associated protein 7, putative (TMA7)
PPT1_040700	5145.259723	802.160417	570.4800715	506.760772	1247.403671	150.473853	1147.428942	359.2926307	316.4425983	316.4425983	9.255972911	1.063973105	-1.951304935	1.49105485	TRIE		3.15809E-06	0.01758756 conserved Phasmodium membrane protein, unknown function
PPT1_040700	10.5827509	529.6380815	16.9451156	333.919146	1676.801248	900.965133	4655.131895	20976.75557	14611.18974	1704.147084	17665.43835	13.80151666	-2.607122472	9.240670529	TRIE		3.6915E-06	0.02650604 ring-infected erythrocyte surface antigen
PPT1_040700	204.1315444	334.590185	106.8142808	87.4044439	56.2144786	257.351303	298.7912038	31.2162519	41.2459638	50.6758693	44.8195257	1.319426493	4.84239423	TRIE		3.99969E-06	0.02176502 Cfpb protein, putative	
PPT1_114900	457.181742	909.400889	67.0245758	516.7108026	1539.625398	1137.763997	1324.228291	354.103294	354.103294	415.5201696	375.4150468	1.11211479	-1.91575107	1.62676946	TRIE		4.28909E-06	0.025701824 Phasmodium exported protein, unknown function
PPT1_071000	198.461237	66.918139	105.0224994	74.6193049	74.6193049	10.14423621	21.0109197	156.2924991	147.523618	375.4150468	8.29492473	-3.40900223	5.92572638	TRIE		4.8326E-06	0.01375498 Phasmodium exported protein, putative	
PPT1_114900	416.315765	314.8585404	297.4396281	260.4588444	581.429635	419.8890941	410.1749069	124.854907	115.76710	84.8976822	8.200239784	0.685784735	-2.78477957	1.51939793	TRIE		2.8066E-06	0.01589694 translation machinery-associated protein 7, putative (TMA7)
PPT1_040700	5145.259723	802.160417	570.4800715	506.760772	1247.403671	150.473853	1147.428942	359.2926307	316.4425983	316.4425983	9.255972911	1.063973105	-1.951304935	1.49105485	TRIE		3.15809E-06	0.01758756 conserved Phasmodium membrane protein, unknown function
PPT1_040700	10.5827509	529.6380815	16.9451156	333.919146	1676.801248	900.965133	4655.131895	20976.75557	14611.18974	1704.147084	17665.43835	13.80151666	-2.607122472	9.240670529	TRIE		3.6915E-06	0.02650604 ring-infected erythrocyte surface antigen
PPT1_040700	204.1315444	334.590185	106.8142808	87.4044439	56.2144786	257.351303	298.7912038	31.2162519	41.2459638	50.6758693	44.8195257	1.319426493	4.84239423	TRIE		3.99969E-06	0.02176502 Cfpb protein, putative	
PPT1_114900	457.181742	909.400889	67.0245758	516.7108026	1539.625398	1137.763997	1324.228291	354.103294	354.103294	415.5201696	375.4150468	1.11211479	-1.91575107	1.62676946	TRIE		4.28909E-06	0.025701824 Phasmodium exported protein, unknown function
PPT1_071000	198.461237	66.918139	105.0224994	74.6193049	74.6193049	10.14423621	21.0109197	156.2924991	147.523618	375.4150468	8.29492473	-3.40900223	5.92572638	TRIE		4.8326E-06	0.01375498 Phasmodium exported protein, putative	
PPT1_114900	416.315765	314.8585404	297.4396281	260.4588444	581.429635	419.8890941	410.1749069	124.854907	115.76710	84.8976822	8.200239784	0.685784735	-2.78477957	1.51939793	TRIE		2.8066E-06	0.01589694 translation machinery-associated protein 7, putative (TMA7)
PPT1_040700	5145.259723	802.160417	570.4800715	506.760772	1247.403671	150.473853	1147.428942	359.2926307	316.4425983	316.4425983	9.255972911	1.063973105	-1.951304935	1.49105485	TRIE		3.15809E-06	0.01758756 conserved Phasmodium membrane protein, unknown function
PPT1_040700	10.5827509	529.6380815	16.9451156	333.919146	1676.801248	900.965133	4655.131895	20976.75557	14611.18974	1704.147084	17665.43835	13.80151666	-2.607122472	9.240670529	TRIE		3.6915E-06	0.02650604 ring-infected erythrocyte surface antigen
PPT1_040700	204.1315444	334.590185	106.8142808	87.4044439	56.2144786	257.351303	298.7912038	31.2162519	41.2459638									

Supp. table 10. Part of whole transcriptome profiles (GLM) of IT4\_Ctrl, IT4\_Wild and IT\_CD151 IEs populations (second experiment)

PTI	IT4_Ctrl.1	IT4_Ctrl.2	IT4_Ctrl.3	IT4_Ctrl.4	IT4_Wild.1	IT4_Wild.2	IT4_Wild.3	IT4_CD151.1	IT4_CD151.2	IT4_CD151.3	IT4_CD151.4	IT4_CD151.5	Mutectp.	chrTRUE	reTRUE	deviance	converged	P-value	Padj	name	
PTI_020100	0	0	0	0	0	0	0	0	0	0	0	0	-40.0289743	2.94379084	93.9194397	0.00458812	TRUE	7.81277E-47	0.00458812	alpha-1b thioester, putative	
PTI_020130	0	0	0.329131691	11.5457849	40.7635667	176.489532	497.6463823	39.3697575	21.2165797	210.9683193	210.9683193	210.9683193	0.0000000	-3.37656846	9.83493084	84.39927075	FALSE	4.32327E-46	0.024167107	knob-associated histidine-rich protein	
PTI_0621800	1701.248626	1386.11863	1901.327702	2562.619344	845.272475	665.801437	3015.49309	396.171256	2584.42512	836.927444	1.22691918	15.83890133	8.635660716	0.97275E-45	-1.32697544	-1.32697544	0.409540316	0.409540316	transcription factor with AP2 domains, putative		
PTI_1134400	1869.224851	3172.74301	2723.888783	736.033788	1372.632631	1188.609771	2553.673328	269.7694901	2572.72432	11.36880995	1.378472859	5.738472859	3.738472859	0.000153646	0.000153646	0.000153646	TRUE	0.000153646	0.851420974	conserved Pflam domain protein, unknown function	
PTI_1035800	412.754526	492.22579	367.723434	1888.45394	1467.81193	1592.07615	802.573604	581.087255	555.0792645	8.55966087	2.13620853	-1.177931643	6.26735073	TRUE	0.000736225	TRUE	0.000736225	TRUE	ADP-ATP transporter on atropine tartrase		
PTI_1001000	86.5916068	31.61929269	125.06673	176.641287	221.5217429	162.2839372	68.62567827	68.62567827	48.3011148	48.3011148	48.3011148	48.3011148	0.0000000	-4.54164294	11.05191083	-1.925653965	TRUE	0.000778264	0.000778264	conserved Pflam domain protein, unknown function	
PTI_1006000	17.3183534	10.2548499	8.15158605	6.08893627	26.6993176	12.2290094	44.40712602	9.91832869	9.91832869	9.91832869	9.91832869	9.91832869	0.0000000	-2.70959885	3.862757809	-3.504937199	TRUE	0.000874766	0.000874766	B_TFvar19 - erythrocyte membrane protein 1, PHEM1, putative	
PTI_1013000	32.882638	158.930718	43.8016463	288.8136586	44.81209149	43.8016463	43.8016463	43.8016463	43.8016463	43.8016463	43.8016463	43.8016463	0.0000000	0.967992016	0.967992016	0.967992016	TRUE	0.001015813	0.001015813	nuclear transport factor 2, putative	
PTI_1013200	144.100877	301.137491	2470.556478	3.929361575	1.89734148	1888.702646	1773.898028	2991.863024	3306.14324	4191.34691	348.640665	99.58354643	1.485159015	0.0001081875	0.0001081875	0.0001081875	TRUE	0.0001081875	0.0001081875	conserved Pflam domain protein, unknown function	
PTI_0732300	23.6683779	37.601159	25.7688172	71.3719833	20.8511236	14.40749874	140.6601171	64.5885767	109.781054	115.9751566	51.6477312	4.83313951	-0.954100397	2.722160304	3.970898414	TRUE	0.0021396155	0.0021396155	Pflam domain exported protein (Pflam), unknown function		
PTI_0219400	504.876062	2103.93368	516.559458	438.481421	391.7927974	94.4848758	331.9478961	680.2579444	955.7280531	928.1659162	1.25593682	7.529939015	7.529939015	0.0001421534	0.0001421534	0.0001421534	TRUE	0.0001421534	0.0001421534	pre-mRNA splicing factor, putative	
PTI_0219800	187.260602	100.632633	379.127016	591.1271477	81.07695495	754.582795	194.891663	239.544349	102.2944533	102.2944533	102.2944533	102.2944533	0.0000000	-1.153987274	-2.06106867	1.699957387	TRUE	0.000151885	0.000151885	liver stage antigen 3	
PTI_1352500	1522.88975	2074.63273	1601.923249	1790.845546	376.1795742	923.960979	794.2929324	1340.21919	1786.201422	1785.456638	1639.1872486	10.134017529	2.1425901	0.000157199	0.000157199	0.000157199	TRUE	0.000157199	0.000157199	trypanin-rich antigen, pseudogene	
PTI_1452400	481.449444	503.671462	640.365137	584.867045	144.222311	217.4046211	236.5169388	635.6199485	464.2322925	464.1088698	456.9724486	32.15215265	9.76746886	-1.564472784	1.27668732	2.926606962	TRUE	0.005260054	0.005260054	Y1L nuclear protein, putative	
PTI_1118500	17133.9031	993.865859	1290.31266	1105.55349	2535.741601	2439.389777	2512.27446	1529.108769	1695.457934	1172.674501	825.540474	1108.624262	10.3028033	0.90313304	0.90313304	0.90313304	TRUE	0.002973138	0.002973138	GTP-binding nuclear protein in yeast	
PTI_1112800	45.0276487	6.8562627	32.9913167	80.0898328	95.1162175	91.58459319	22.86321697	13.6279931	4.89613378	19.7594266	4.88429254	1.58002344	-0.39532344	4.70093397	4.70093397	4.70093397	TRUE	0.00363312	0.00363312	conserved Pflam domain protein, unknown function	
PTI_0201100	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	FALSE	0.004571245	0.004571245	knob-associated heat shock protein 40 (Knob40)	
PTI_0197000	5530.27878	9990.37469	5944.27473	9827.70772	2594.356938	524.129896	5447.62616	8886.362336	1312.03398	1145.57495	1082.162728	3338.060754	17.7854197	-1.93444351	15.1648755	10.9792622	TRUE	0.002906067	0.002906067	conserved Pflam domain protein, unknown function	
PTI_1325000	1080.6510	196.20649	1397.164072	1397.164072	649.4390401	638.026746	815.813872	572.729923	1081.8634	1481.127933	1268.09854	1360.400044	3.860001553	0.049257095	-1.03317113	1.00893343	3.860001553	TRUE	0.002376707	0.002376707	line expressed repeat antigen
PTI_1023400	1091.04957	167.12293	1757.147336	1820.817199	7114.023254	9937.024784	197.523101	4528.45977	1747.10025	1950.14268	32.15215265	9.74316096	0.006943909	-1.927196485	0.911662499	2.74316096	TRUE	0.005418608	0.005418608	conserved Pflam domain protein, unknown function	
PTI_0219600	359.982768	339.0097441	799.764003	392.5671091	824.851181	756.837806	567.5079021	390.6345777	609.59231	370.653077	0.468467655	-1.027224656	9.751764651	0.005418608	0.005418608	0.005418608	TRUE	0.005418608	0.005418608	Pflam domain exported protein, unknown function	
PTI_1409100	1239.21014	1316.02966	1199.13473	920.252074	302.439365	288.01123	3104.77933	1673.33032	1784.25765	1573.276071	1309.49971	101.9423661	-1.08737891	-0.90742002	1.10218439	1.0284668	TRUE	0.00854668	0.00854668	histo-septic peptase	
PTI_0617000	136.788434	59.653887	1705.16915	158.400039	350.247057	2174.6621	486.246698	1045.925641	1768.124017	79.8772234	164.836494	333.710253	7.41020067	1.23576017	-1.19228295	9.1279396	TRUE	0.00623991	0.00623991	6S ribosomal protein L36, putative	
PTI_0707900	136.788434	59.653887	1705.16915	158.400039	350.247057	2174.6621	486.246698	1045.925641	1768.124017	79.8772234	164.836494	333.710253	7.41020067	1.23576017	-1.19228295	9.1279396	TRUE	0.00643691	0.00643691	chloramphenicol resistance transporter	
PTI_1458400	69.423934	140.01467	1589.23865	1954.05488	632.430823	285.345563	140.60829	134.899292	210.477084	7.04913785	15.24436892	0.911662499	2.74316096	0.006943909	-1.927196485	0.911662499	TRUE	0.006943909	0.006943909	thioredoxin 1	
PTI_0705400	69.423934	140.01467	1589.23865	1954.05488	632.430823	285.345563	140.60829	134.899292	210.477084	7.04913785	15.24436892	0.911662499	2.74316096	0.006943909	-1.927196485	0.911662499	TRUE	0.007360808	0.007360808	conserved Pflam domain protein, unknown function	
PTI_0609000	20.9471239	9.4002795	13.159457	12.342484	678.514864	35.461283	403.23372	353.953396	331.079776	340.040974	36.6232469	724.900457	7.74660558	-2.88496158	5.5714651	4.07958724	TRUE	0.00758174	0.00758174	erythrocyte membrane protein 1, PHEM1	
PTI_0606200	4.0494219	13.6733365	1705.16915	3.29136098	42.74727378	720.01928	3.29136098	87.37302393	80.7602019	63.8972925	80.1622226	339.248833	2.875319438	3.948497894	-2.875319438	3.948497894	5.091044629	TRUE	0.007841628	0.007841628	B_TFvar54 - erythrocyte membrane protein 1, PHEM1, putative
PTI_1327100	146.152103	53.87964	158.840213	217.556674	155.680096	2033.06823	282.734339	39.7728393	80.7399218	63.8972925	79.970228	101.0063243	1.73533283	0.86874326	-1.353199142	5.14721496	TRUE	0.007841628	0.007841628	conserved Pflam domain protein, unknown function	
PTI_0833400	3483.86978	1927.05719	4840.26942	5141.366141	531.8274218	1277.52149	1555.98943	4458.877765	1594.537819	2888.123651	2412.16711	2114.155698	11.9100297	-1.783791743	1.263143538	9.448246111	TRUE	0.007963838	0.007963838	trypanin-thionein-rich antigen	
PTI_1464000	4975.55869	5147.08024	5567.77775	327.328811	105.5614193	9022.291778	9768.15232	623.632626	106.692754	150.481902	159.641266	45.852839	12.2111618	1.043824669	-0.830295635	4.81949415	TRUE	0.008113996	0.008113996	glyceraledehyde-3-phosphate dehydrogenase	
PTI_1064800	1415.484457	2242.49321	1420.63584	1135.339707	927.919462	1413.33087	1348.88438	1889.529242	1596.50104	2911.8487819	3072.69643	312.085719	10.6063933	-0.343698537	0.21015762	6.83398424	TRUE	0.008312093	0.008312093	RNA binding protein, putative	
PTI_1461900	1261.559075	1035.39629	1650.65965	1790.282927	665.596746	2238.397367	2731.98919	1493.01353	1683.34752	1515.735656	186.792502	1197.506393	0.475138072	0.935842423	-0.36678326	2.448338239	TRUE	0.008625834	0.008625834	6S ribosomal protein L27, putative	
PTI_0625600	71.1597409	438.976653	620.347956	488.292037	3067.949819	964.739064	923.868672	931.796973	941.2299162	312.591429	390.647412	390.647412	390.647412	0.0000000	-0.94945434	1.145819142	6.74543229	TRUE	0.012405765	0.012405765	co-chaperone p25
PTI_0802900	50.8609044	258.934957	122.8813855	147.7801292	55.5648599	106.7032016	64.83280931	145.9408723	141.2298163	258.9318643	19.3171573	188.697968	7.2948909	-1.149257821	1.388016953	4.44018088	TRUE	0.012559694	0.012559694	micro-fibrillar-associated protein, putative	
PTI_0935600	29125.95955	66147.19943	4796.93154	38986.65823	19648.02855	31632.37238	3984.249872	5883.78151	41.1624237	4952.149427	487.403668	6381.26717	15.4706203	-0.69919164	0.811814208	7.68435536	TRUE	0.012735047	0.012735047	ring-exported protein 1 (REX1)	
PTI_1420300	144.319378	50.8195711	135.116107	105.818396	271.257945	244.500198	208.531252	134.589872	100.695754	92.034347	50.48016215	6.800751214	1.23339281	-1.227474418	9.545457867	0.012751442	TRUE	0.012751442	0.012751442	glutathione S-transferase (GST)	
PTI_1001000	44.0904259	5.8371248	5.00300767	6.25250125	493.840385	621.472286	393.740124	827.5406154	180.949615	102.044459	687.024713	9.02890747	0.28904857	2.890714957	4.84304601	0.013543694	TRUE	0.013543694	0.013543694	sbf, CS3 - yeast cytochrome membrane protein 1, putative	
PTI_1021000	755.8884165	508.55691	1068.53691	1667.209507	995.1071837	1521.823248	1674.849641	884.2866499	1069.75532	616.28924	59.5230653	68.2673249	9.7833337	0.6							

Supp. table 11. var genes profiles (GLM) of IT4\_Ctrl, IT4\_Wild and IT4\_CD9 IEs populations

var_genes (Nom1)	IT4_Ctrl.1	IT4_Ctrl.2	IT4_Ctrl.3	IT4_Ctrl.4	IT4_Wild	IT4_Wild2	IT4_Wild3	IT4_CD9.1	IT4_CD9.2	IT4_CD9.1+3	IT4_CD9.4	X_Interes	coefIT4_U	recIT4_U	deviance	convergence	P-value	Postj	
PTT.010000	1.827374196	3.559570947	6.48551063	5.2312868	3.27654447	0.420110657	0.097258214	1.218801	-0.84	8.25792	1.218801	0.58	0.64	0.58	0.58	0.58	0.58	0.58	
PTT.011000	0.456843549	4.271485137	1.62508766	3.1663052	3.5661567	11.0487924	634.1854208	1025.029769	183.62036	131.52493	1.3852	0.8678	1.1712	0.90451	1.1712	0.90451	0.90451	0.90451	
PTT.011500	17.81698841	11.5263696	11.7556859	9.0859192	4.754868894	0	13.26199732	64.1490472	150.84887	106.45147	19.77249533	3.69088	-0.7766	4.35708	4.39725	TRUE	6.1549E-09	4.12378E-07	
PTT.041100	3.197904842	19.93397931	19.937826	24.091452	29.71793959	55.243962	79.57196395	40.723037469	41.920264	60.935712684	58.439299747	1.85516	-1.16934	3.09558	3.9779	TRUE	1.162664E-08	1.751888E-06	
PTT.041600	4.11159194	7.83105084	4.63250759	2.7539888	4.754868894	0	83.05186726	117.326899	90.99883727	87.52503156	87.52503156	0.80196	0.13337	5.60118	5.72066	TRUE	3.8437E-08	2.48997E-06	
PTT.042000	22.3853339	21.35742568	7.87526291	6.7456067	4.754868894	0	1.473555258	82.04260192	67.0439242	109.8853884	74.52203156	0.80196	0.13337	5.60118	5.72066	TRUE	8.0481E-08	4.07033E-06	
PTT.043000	64.4149404	190.0810886	101.915167	82.4616	14.83896829	0	7.367110517	156.3232458	67.0439242	118.4710184	101.4310838	3.83549	-1.9453	4.34313	5.71127	TRUE	8.1311E-08	5.07033E-06	
PTT.044000	15.53268066	63.36036287	40.1483991	37.032004	3.5661567	0	2.967176592	16.27224148	58.6344946	123.6210619	100.4834173	6.77348	-0.9042	3.27831	4.31127	TRUE	1.4403E-06	6.97861E-05	
PTT.044500	77.20658976	222.1172271	104.694672	80.121287	8.321026564	0	4.420667575	69.41835867	30.7284756	73.82924533	68.00024621	9.16231	-0.3416	4.55325	8.43318	TRUE	3.61207E-06	0.000216724	
PTT.045000	27.86745648	71.19141895	23.008121	16.795184	8.321026564	0	10.31486881	120.1836981	55.869952	142.5076131	68.06924621	4.99872	-2.0674	3.98157	7.068	TRUE	9.62761E-06	0.000558401	
PTT.045500	114.2108872	304.6992731	111.79785	112.748	34.47279948	33.1463772	2.947110517	104.9309424	181.577344	163.1111234	58.345069338	7.34199	-2.3635	5.25671	4.73743	TRUE	1.36164E-05	0.000767132	
PTT.046000	9.136870978	17.08594055	13.1254382	8.8108385	17.23639974	0	2.947110517	104.9309424	181.577344	163.1111234	58.345069338	7.34199	-2.3635	5.25671	4.73743	TRUE	1.36164E-05	0.000767132	
PTT.046500	27.41061293	30.6455115	15.1328581	17.758842	17.83075835	0	2.947110517	104.9309424	181.577344	163.1111234	58.345069338	7.34199	-2.3635	5.25671	4.73743	TRUE	1.36164E-05	0.000767132	
PTT.047000	5.93896156	8.542970274	8.95618135	9.361501	19.61383419	22.0975848	10.31486881	146.5610361	75.62309376	152.8309682	126.4143144	3.04513	0.9854	3.97461	2.21459	TRUE	3.45149E-05	0.001917325	
PTT.047500	13.24846292	79.02247503	48.6413297	37.995662	5.349227506	11.0487924	4.420667575	83.05186726	53.0716454	4.420667575	4.420667575	4.77488	-0.9979	3.76402	6.28225	TRUE	4.2446E-05	0.002392103	
PTT.048000	6.85265324	21.35742568	9.41943211	8.535274	2.377434447	0	1.473555258	61.5558927	30.7284756	48.07485742	32.41392677	3.0734	-2.5942	4.53536	2.63517	TRUE	5.69574E-05	0.003018742	
PTT.048500	2.284217745	2.135742568	2.62508766	3.49033125	2.377434447	0	48.3588331	25.18418784	60.99883727	48.3588331	48.3588331	48.3588331	4.3413	-0.4354	2.13137	2.94674	TRUE	0.002307403	0.095757951
PTT.049000	30.60851778	84.00587436	78.7526291	8.105283	17.23639974	0	13.26199732	127.9971737	111.739904	34.33918387	22.68974874	4.00838	-3.7018	2.5839	6.86657	TRUE	0.002300023	0.101201081	
PTT.049500	9.136870978	17.08594055	13.1254382	8.8108385	17.23639974	0	13.26199732	127.9971737	111.739904	34.33918387	22.68974874	4.00838	-3.7018	2.5839	6.86657	TRUE	0.002300023	0.101201081	
PTT.050000	17.81698841	11.5263696	11.7556859	9.0859192	4.754868894	0	8.84133155	119.5263971	64.2504448	97.86607404	77.79342424	4.33198	-0.7576	2.91922	3.87984	TRUE	8.68262E-05	0.004341311	
PTT.050500	265.8829455	627.9083151	254.633501	182.68204	33.28408226	0	14.7355258	129.953258	129.953258	143.1902685	35.31696611	10.8166	-0.1007	1.2347	3.70889	TRUE	0.002972865	0.133778924	
PTT.051000	13.24846292	79.02247503	48.6413297	37.995662	5.349227506	11.0487924	5.894215033	68.3954633	41.902464	113.190368	38.89671212	4.8317	-0.4958	1.26403	4.74882	TRUE	0.004405755	0.193853233	
PTT.051500	10.50740162	19.22168312	20.3830334	15.143199	10.69845501	0	1.473555258	67.18913429	47.4894592	56.65965339	35.65531944	4.02861	-0.6045	3.22375	3.22375	TRUE	0.004407296	0.193853233	
PTT.052000	15.9852421	3.559570947	6.48551063	5.2312868	3.27654447	0	4.420667575	43.96490474	30.7284756	60.9357178	68.89671212	3.7631	-2.1126	2.08597	3.32303	TRUE	0.001710485	0.082103291	
PTT.052500	1.370530647	3.559570947	6.48551063	5.2312868	3.27654447	0	8.84133155	29.31232986	55.869952	60.9357178	32.41392677	1.99756	0.77935	2.62421	3.2173	TRUE	0.004489652	0.193853233	
PTT.053000	11.09947086	32.85412793	13.6587942	8.8423997	16.85147922	16.2748712	1.473555258	46.8997277	111.739904	34.33918387	22.68974874	4.00838	-3.7018	2.5839	6.86657	TRUE	0.005442612	0.223147087	
PTT.053500	72.63812428	253.4414515	80.4512152	47.08181	11.88717223	0	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.054000	18.27374196	18.50976893	4.32367375	5.2689152	5.349227506	11.0487924	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.054500	3.985896136	7.83105084	4.68551063	6.6799142	7.726661953	0	2.947110517	26.38109687	19.5544832	29.18830629	25.93114141	-0.5803	1.69882	3.54433	4.97574	TRUE	0.014066358	0.534521608	
PTT.055000	39.28854521	14.95019798	16.5226104	24.7978	10.81723673	47.5098074	543.7418903	111.3686535	64.2504448	92.71579646	81.89671212	3.06147	-2.5164	1.90621	1.90621	TRUE	0.001964087	0.707057534	
PTT.055500	10.9852421	3.559570947	6.48551063	5.2312868	3.27654447	0	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.056000	15.9852421	3.559570947	6.48551063	5.2312868	3.27654447	0	8.84133155	29.31232986	55.869952	60.9357178	32.41392677	1.99756	0.77935	2.62421	3.2173	TRUE	0.005442612	0.223147087	
PTT.056500	11.09947086	32.85412793	13.6587942	8.8423997	16.85147922	16.2748712	1.473555258	46.8997277	111.739904	34.33918387	22.68974874	4.00838	-3.7018	2.5839	6.86657	TRUE	0.005442612	0.223147087	
PTT.057000	72.63812428	253.4414515	80.4512152	47.08181	11.88717223	0	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.057500	18.27374196	18.50976893	4.32367375	5.2689152	5.349227506	11.0487924	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.058000	3.985896136	7.83105084	4.68551063	6.6799142	7.726661953	0	2.947110517	26.38109687	19.5544832	29.18830629	25.93114141	-0.5803	1.69882	3.54433	4.97574	TRUE	0.014066358	0.534521608	
PTT.058500	39.28854521	14.95019798	16.5226104	24.7978	10.81723673	47.5098074	543.7418903	111.3686535	64.2504448	92.71579646	81.89671212	3.06147	-2.5164	1.90621	1.90621	TRUE	0.001964087	0.707057534	
PTT.059000	10.9852421	3.559570947	6.48551063	5.2312868	3.27654447	0	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.059500	15.9852421	3.559570947	6.48551063	5.2312868	3.27654447	0	8.84133155	29.31232986	55.869952	60.9357178	32.41392677	1.99756	0.77935	2.62421	3.2173	TRUE	0.005442612	0.223147087	
PTT.060000	11.09947086	32.85412793	13.6587942	8.8423997	16.85147922	16.2748712	1.473555258	46.8997277	111.739904	34.33918387	22.68974874	4.00838	-3.7018	2.5839	6.86657	TRUE	0.005442612	0.223147087	
PTT.060500	72.63812428	253.4414515	80.4512152	47.08181	11.88717223	0	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.061000	18.27374196	18.50976893	4.32367375	5.2689152	5.349227506	11.0487924	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.061500	3.985896136	7.83105084	4.68551063	6.6799142	7.726661953	0	2.947110517	26.38109687	19.5544832	29.18830629	25.93114141	-0.5803	1.69882	3.54433	4.97574	TRUE	0.014066358	0.534521608	
PTT.062000	39.28854521	14.95019798	16.5226104	24.7978	10.81723673	47.5098074	543.7418903	111.3686535	64.2504448	92.71579646	81.89671212	3.06147	-2.5164	1.90621	1.90621	TRUE	0.001964087	0.707057534	
PTT.062500	10.9852421	3.559570947	6.48551063	5.2312868	3.27654447	0	5.894215033	61.5558927	37.5588331	50.2829568	47.34333008	3.99652	-1.70753	2.6343	1.92337	TRUE	0.021257222	0.761890335	
PTT.063000	15.9852421	3.559570947	6.48551063</																



Supp. table 13. Part of *rifin* genes expression profiles of 3D7\_Ctrl and 3D7\_Wild IE populations

	3D7_Ctrl	3D7_Ch2	3D7_Ch3	3D7_Ch4	3D7_Ch5	3D7_Wild_1	3D7_Wild_2	3D7_Wild_3	3D7_Wild_4	baseMeanA	baseMeanB	foldChange	log2FCChange	P-value	Padj	name
PE3D7_0617500	1.70673491	2.62526003	4.37521149	5.71320174	3.959382638	1202.737782	928.5815327	2008.236763	1916.86001	3.685869491	15.14110522	0.00243613	-3.681193387	2.905767-22	8.198391E-19	rifin_pseudogene
PE3D7_0400500	32.73895959	46.97959339	38.85061338	38.85061338	56.75115191	1029.583777	1202.809441	1189.494083	1555.257119	37.56916232	375.6916232	0.0523340494	-4.959114495	1.056748E-10	3.13831E-08	rifin (RF)
PE3D7_0804000	33.62379633	38.50381338	70.0037838	107.4104193	73.90837878	1045.962777	1421.213709	1783.837878	1525.257119	64.66997666	199.816835	0.063196454	-4.213128177	1.144558E-09	2.02011E-07	rifin (RF)
PE3D7_0804000	15.04222468	15.31401669	25.13860877	25.13860877	32.08485136	220.851573	401.4473526	401.4473526	443.397645	19.9614815	386.3139645	0.051671654	-2.63878E-09	3.914247E-07	5.60188E-07	rifin (RF)
PE3D7_0937500	6.193857219	17.93927669	17.93927669	17.93927669	31.6750611	30.931976117	28.85112938	40.1921501	435.8091764	18.33333333	356.657558	4.2283179433	-4.2283179433	4.169398E-09	5.60188E-07	rifin (RF)
PE3D7_0937500	10.61840993	8.75086678	13.97691319	13.97691319	17.63724871	132.740121	264.932773	247.839838	10.80409065	204.8506563	0.052471328	-4.204922302	2.57302E-08	2.74043E-06	3.20516E-06	rifin (RF)
PE3D7_0302300	15.04222468	24.06488336	8.750422298	18.28262456	18.28262456	18.28262456	18.28262456	39.71029601	383.457936	256.1634461	307.9919163	0.054947724	-4.185796469	5.18934E-08	4.57554E-06	rifin (RF)
PE3D7_0617500	0.884836746	1.750173336	17.93927669	15.99229649	21.1167074	303.1991303	299.51059369	262.0155767	285.7026902	18.93408664	287.775835	0.065744077	-3.926272466	1.77018E-07	9.62272E-06	rifin (RF)
PE3D7_0900200	53.97504148	57.31817674	48.12732264	47.9065614	69.1496886	373.1102861	691.496886	977.084251	1010.706388	75.23373333	888.5949663	0.084653985	-3.562308414	2.08374E-07	1.3363E-05	rifin (RF)
PE3D7_0200200	1.70673491	0.437543334	0	116.1418907	0	13.65112658	30.03072191	38.61993775	35.53795207	0.44144365	35.53795207	0.012421745	-6.330988377	2.88957E-07	1.61444E-05	rifin (RF)
PE3D7_0102000	35.9346982	51.6301134	4.37521149	23.99594473	27.71567846	399.4756314	357.2075343	463.9206465	561.4098869	28.62208351	463.9206465	0.061696076	-4.018674453	3.19077E-07	1.76524E-05	rifin (RF)
PE3D7_0701200	28.31477586	61.69361008	35.00168919	49.1345535	58.07094535	455.5171768	377.548703	508.8236763	618.6914027	16.03308932	16.03308932	0.038496228	#DIV/0!	1.08449E-06	5.32153E-05	rifin_pseudogene (RF)
PE3D7_1040800	7.078093965	16.18910335	8.750422298	15.99229649	27.71567846	35.92406757	30.82100466	40.93713401	46.3627635	0.087508667	50.28263579	0.002875897	-8.441772386	2.20178E-06	9.55743E-05	rifin (RF)
PE3D7_0729000	1.70673491	2.62526003	4.37521149	2.28532807	3.959382638	12.93266433	162.798124	147.5281622	151.7324199	15.14623891	162.798124	0.025288638	-3.435896402	2.48686E-06	0.00058609	rifin (RF)
PE3D7_0833400	3.59346982	4.375433339	13.12563345	12.56930438	22.43650161	94.12105705	81.3996201	86.50997804	94.79997804	11.82409977	118.7479381	0.0523340494	-3.800197751	4.23524E-06	0.000161483	rifin (RF)
PE3D7_0324800	18.58157166	18.37469202	35.00168919	37.70791315	34.31464953	212.588998	171.4725236	215.7971938	28.79652871	23.6082428	223.6082428	0.128781159	-2.879570654	5.78784E-06	0.000210715	rifin (RF)
PE3D7_0937500	25.60926562	31.94066337	4.37521149	30.85192894	32.94845531	216.9813682	212.516534	263.3879534	320.3298598	25.16458488	252.7270891	0.099572171	-3.228113607	8.95283E-06	0.000301558	rifin (RF)
PE3D7_1302300	14.15738793	15.31401669	43.5211149	35.42258898	52.7917685	48.20721108	40.93713401	48.93713401	103.8002913	1.489080177	305.0667316	0.038496228	-4.699139112	1.44064E-05	0.00136123	rifin (RF)
PE3D7_1479400	1.70673491	3.062803337	4.37521149	2.28532807	2.69588425	23.7098846	23.7098846	51.73071668	151.900156	32.28573794	205.0667316	0.038496228	-2.66650336	1.5145E-05	0.00452186	rifin (RF)
PE3D7_0701200	46.01151077	63.44378341	87.50422298	138.2623482	116.1418907	655.9734739	499.4583222	595.1544001	592.599325	90.27275122	585.9876403	0.154078607	-2.698261534	2.27801E-05	0.0006142	rifin (RF)
PE3D7_1301000	0	0	0	0	1.319794213	17.24352444	11.85432333	9.26878506	16.01619797	0.263958843	13.59569195	0.019414888	-5.686697772	2.30409E-05	0.000616208	rifin_pseudogene (RF)
PE3D7_0632600	14.15738793	17.93927669	21.87665574	61.70387788	47.51259165	92.68409434	329.5476638	220.1336452	249.5159053	32.67783398	22.9702259	0.146377544	-2.772233854	2.53358E-05	0.000660885	rifin_pseudogene (RF)
PE3D7_1400200	8.848367456	5.250520007	4.37521149	11.42664035	10.5583537	54.0458271	96.41442297	70.2882867	59.0074617	8.091818352	70.07858439	0.11546778	-3.114437761	3.13444E-05	0.000796665	rifin (RF)
PE3D7_0632300	2.65451037	3.937890005	4.37521149	7.998648244	5.27917685	113.5205315	39.51410777	47.11632405	47.20536313	4.849087297	61.83004037	0.078414072	-3.78723567	3.79188E-05	0.000922309	rifin (RF)
PE3D7_1255100	4.424183728	4.375433339	8.750422298	14.85463245	11.87814791	86.9364353	61.6240813	43.2433028	52.2633826	8.856563946	61.0239912	0.145132403	-6.4407E-05	6.4407E-05	0.00136123	rifin (RF)
PE3D7_1040000	0	0	0	0	1.319794213	12.21418298	7.902821555	12.3583808	14.308284	0.263958843	11.70141675	0.022557853	-5.470224408	7.79294E-05	0.001578556	rifin (RF)
PE3D7_0101600	10.61840993	10.50104001	30.62647804	28.56660087	29.05347268	131.4820873	149.3635274	84.90366305	107.8989699	2.150952651	18.4240987	0.184666776	-2.47903766	8.42859E-05	0.001663017	rifin (RF)
PE3D7_1200200	18.58157166	28.87876004	52.0253379	79.98648254	85.78662382	296.0143168	365.1103538	199.278988	244.4577585	53.17014435	276.2132375	0.192411532	-2.37773926	0.000102349	0.001925189	rifin (RF)
PE3D7_0832800	1.70673491	0.97508668	0	2.28532807	0	9.34257659	11.06395108	23.9443614	30.34648037	0.986017	18.67376238	0.052802302	-4.243255355	0.000135407	0.002410417	rifin (RF)
PE3D7_0832800	0.848367456	0	0	0	0	12.93266433	8.69310771	13.1307883	16.85915576	1.280152386	12.90392586	0.099236652	-3.33298312	0.000242157	0.0083838461	rifin (RF)
PE3D7_1200500	7.255661314	83.57077677	39.37690034	33.13725701	23.75629583	274.4598763	234.7138002	380.0201875	474.5852347	50.47506862	340.9447746	0.148057904	-3.755766587	0.000259947	0.00409744	rifin (RF)
PE3D7_0609000	0	0	0	0	0	17.13996452	17.93927669	8.69310771	16.9927261	16.9927261	18.07711873	0.069010406	-8.875069744	0.000277292	0.004264172	rifin_pseudogene (RF)
PE3D7_0937500	4.424183728	13.56384335	0	17.13996452	18.4771898	92.68409434	80.687986	54.803116	80.0808987	10.72102132	77.05334392	0.13913729	-2.845418964	0.000275305	0.00429157	rifin (RF)
PE3D7_0937500	7.96353071	16.18910335	4.37521149	7.998648244	5.27917685	99.1502651	75.86708692	40.93713401	57.3212959	8.561134062	68.31894426	0.12238383	-3.036515141	0.000314935	0.004739148	rifin (RF)
PE3D7_1373400	69.0021029	84.00832011	105.0058676	170.2569412	133.2992125	487.1303563	531.0690693	546.8583185	580.797916	112.4943294	536.4640498	0.209659333	-2.255029214	0.000338282	0.005046063	rifin (RF)
PE3D7_0533000	45.12667403	48.50731006	48.12732264	48.12732264	48.12732264	48.12732264	48.12732264	48.12732264	48.12732264	170.2873614	170.2873614	0.201883903	-2.308042214	0.00065982	0.006546667	rifin_pseudogene (RF)
PE3D7_0632400	0.848367456	4.812976673	4.37521149	12.56930438	10.5583537	52.4913866	53.73918657	28.57875393	28.57875393	6.64013653	39.3817486	0.168609548	-2.568241856	0.000611837	0.007447875	rifin (RF)
PE3D7_1101400	0	2.187116669	0	4.57065614	1.319794213	15.08810838	7.902821555	19.3996887	35.402271	1.615633404	19.4263148	0.083167404	-3.587837986	0.000626262	0.007567449	rifin_pseudogene (RF)
PE3D7_1254500	0	0.87508668	0	4.57065614	6.598971063	11.49570162	20.5473604	26.24515767	21.079447	2.408432774	19.84463301	0.121390128	-3.042277001	0.000703886	0.008240724	rifin (RF)
PE3D7_1040100	0	2.187116669	8.750422298	2.28532807	1.319794213	17.96203379	17.96203379	13.90317759	25.28873364	2.90865225	18.64503811	0.15608125	-2.67989842	0.000716761	0.008356784	rifin (RF)
PE3D7_0732300	0.848367456	0.437543334	0	0	0	7.902821555	7.902821555	10.0418381	13.48732461	0.264476016	9.835656211	0.026894983	-5.216519117	0.000845252	0.009322526	rifin (RF)
PE3D7_0101000	0	1.312630002	4.37521149	1.42664035	3.959382638	64.0632163	14.229788	35.21314446	45.5192956	2.43506095	2.43506095	0.068681838	-3.52846449	0.001078204	0.01112023	rifin (RF)
PE3D7_0115300	0	0.437543334	0	3.427992105	3.959382638	15.01536095	15.01536095	10.0418381	21.91609249	1.564983615	14.79690756	0.105764235	-3.241076237	0.001110148	0.01130878	rifin (RF)

Supp. table 14. var genes expression profiles of 3D7\_Ctrl and 3D7\_Wild IEs populations

	3D7_Ctrl_1	3D7_Ctrl_2	3D7_Ctrl_3	3D7_Ctrl_4	3D7_Ctrl_5	3D7_Wild_2	3D7_Wild_3	3D7_Wild_4	baseMean	baseMeanB	foldChange	log2FoldChange	P-value	Padj
PF3D7_0809100	4989.59441	4841.416989	10487.38112	14738.08072	14476.82272	11.85423233	11.58598132	17.70211355	9906.65919	11.9021648	832.34095	9.701030807	2.67091E-21	1.84292E-19
PF3D7_0712900	322.08557	310.2182237	686.9081504	875.9026736	830.5572735	6.322275244	5.406791285	5.406791285	4.371833415	4.4973072	107.95853	6.734333415	2.16845E-16	1.47455E-14
PF3D7_1200400	897.22446	802.8920177	1281.9536867	2118.499123	1919.592206	12.811824298	12.811824298	24.49874682	95.96497	23.174782	40.466154	5.338643825	4.62423E-13	3.09831E-11
PF3D7_0421200	839.710072	896.5262911	2103.948058	2118.499123	172.41614689	3.089595052	4.214788949	4.214788949	3.76796E-11	0.90835548	10.9835548	9.0835548	3.76796E-11	2.48685E-09
PF3D7_0421300	54.8598782	75.69499676	416.261402	207.994843	271.8930418	2.3740846646	0.842957788	0.842957788	149.534835	0.20655075	548.02524	7.229322608	6.42281E-11	4.17483E-09
PF3D7_0722500	229.186165	116.3865268	271.2630912	157.6876368	177.8107782	0.718481351	10.27366802	6.433492523	3.371833152	4.79059226	38.2950742	5.181701858	4.85148E-08	1.485148E-08
PF3D7_0722600	226.518210	246.336978	866.2918075	588.6282188	598.6282188	0.718481351	3.951410777	10.27366802	5.29356694	4.79059226	38.2950742	5.181701858	4.85148E-08	1.485148E-08
PF3D7_0632800	245.984615	223.1471003	415.6450594	445.6389736	403.8570291	6.436623163	12.64451449	10.34118381	23.60281807	2.72553669	88.3150424	6.464503482	1.63473E-10	1.02988E-08
PF3D7_0712000	385.788821	397.2893472	1437.070946	541.6297525	712.6888748	1.436962303	13.164128632	701.4924558	701.4924558	2.36104024	297.11148	8.211860517	1.97051E-08	1.221771E-08
PF3D7_0617400	611.422191	574.0668541	2253.233742	1993.948741	1726.29083	18.56196572	18974.67455	34424.62338	1431.79047	24191.3254	0.0591861	4.078597526	4.54659E-10	2.72795E-08
PF3D7_0200100	189.358064	168.4541835	183.7588683	417.0725727	378.780939	15.806858973	15.806858973	15.806858973	267.4842882	42.3880659	22.0552523	4.46175922	1.30022E-09	9.58797E-08
PF3D7_0712800	742.374003	693.9437275	1146.305321	1117.686809	1096.748991	51.3683430	51.3683430	10.75011658	959.63521401	4.9880659	19.4299774	4.280212058	1.65310E-08	1.22901E-06
PF3D7_0412900	206.166992	216.5839503	388.7639583	444.4260316	390.6598869	4.310888109	21.3376482	77.24974771	309.334049	2.06337575	14.991649	3.904807173	3.58808E-08	1.99253E-06
PF3D7_0733500	230.953898	241.3865268	56.25464528	116.0097433	116.0097433	6.466332163	17.466332163	17.466332163	396.43405	4.65011911	15.546798	3.985854585	4.14274E-08	2.7875E-06
PF3D7_0733600	233.569901	241.3865268	56.25464528	116.0097433	116.0097433	6.466332163	17.466332163	17.466332163	396.43405	4.65011911	15.546798	3.985854585	4.14274E-08	2.7875E-06
PF3D7_0733700	233.569901	241.3865268	56.25464528	116.0097433	116.0097433	6.466332163	17.466332163	17.466332163	396.43405	4.65011911	15.546798	3.985854585	4.14274E-08	2.7875E-06
PF3D7_0413100	342.431681	336.4708238	1264.436022	507.3428315	427.6133249	1.436962303	13.164128632	701.4924558	1.80140903	575.6519999	1.47113277	128.5776	7.06495463	5.15263E-06
PF3D7_0413200	21.2360819	23.1897962	65.62816723	73.13049823	58.07094335	0.718481351	3.951410777	10.27366802	0.842957788	4.8259975	30.707519	4.940520039	9.92011E-08	5.18965E-06
PF3D7_0500100	276.953901	234.0856836	301.8895693	461.6362701	380.1007332	10.77722027	8.74326131	28.57875393	330.93232	25.4469923	13.004047	9.009731119	1.17468E-07	5.99085E-06
PF3D7_1255200	135.380022	136.5135202	459.3971706	521.0546799	459.2888386	10.05873892	36.91301371	13.13077883	10.1149346	10.4995287	32.604014	5.026977677	2.14657E-07	1.02829E-05
PF3D7_0733000	90.2533481	95.82199012	240.6366132	221.6768228	196.6493377	10.77722027	11.06395018	14.67557634	169.007622	11.2365812	15.4084	3.910881326	2.59124E-07	1.26971E-05
PF3D7_0800100	150.422247	117.6991568	218.7605574	291.3793289	253.4004888	14.36962703	24.49874682	11.095845125	206.3323567	11.2634976	11.550502	3.529883359	1.14757E-06	5.0832E-05
PF3D7_0632500	1058.26475	1019.475968	6326.55323	4190.149016	4247.097776	23.7098846	75.86708692	84.19146429	3368.30857	69.7565665	48.287239	5.95370081	7.30386E-06	0.00343281
PF3D7_1000100	181.391533	179.8303102	214.3853462	246.8154315	203.2483087	12.21418298	52.15862226	16.22037385	20.1341159	25.4167800	80.708169	3.01271471	9.56543E-06	0.0044001
PF3D7_0712600	741.493193	640.1258975	3023.270904	844.4282718	902.7392414	10.05873892	90.89344768	23.2323402	252.0437386	170.636011	15.430713	9.947732776	2.2857E-05	0.001438561
PF3D7_0103300	1459.09579	1340.63275	1623.203336	4240.426233	4501.818059	11.63939789	90.89344768	23.2323402	252.0437386	170.636011	15.430713	9.947732776	2.2857E-05	0.001438561
PF3D7_0115700	30.084494	35.4410104	70.00337838	63.98918595	46.1970744	4.310888109	7.112539399	6.95158879	0.82957788	49.1421642	4.80449352	10.228376	4.12204E-05	0.001772477
PF3D7_0711700	1199.58863	1064.105588	8194.770482	2251.0481449	322.0297879	5.02396946	8.95140371	10.95845125	3017.04343	8.6052706	346.85387	8.438121414	4.82113E-05	0.002024874
PF3D7_0712800	105.295573	102.626835	826.9149071	450.7844311	322.0297879	2.155444054	3.951410777	10.08959502	4.214788949	3.5268997	106.6477	7.873620104	4.91978E-05	0.002024874
PF3D7_0937600	320.130972	316.348304	538.1509713	761.0142472	680.0508266	106.335254	48.99749364	18.53717012	559.2605825	79.5284566	70.272104	2.812925099	5.49518E-05	0.002198074
PF3D7_1240900	729.105478	761.325401	8072.26457	3318.296357	3017.049592	2.873925406	19.75709368	18.53717012	3179.60828	14.9284505	17.24645178	6.89222E-05	0.002686794	0.000698719
PF3D7_0420900	145.998063	166.7040102	1400.0607568	585.0439859	599.7958329	5.02396946	2.370846466	3.089595018	567.52196	3.67194947	154.37942	4.720336611	7.10189E-05	0.002686794
PF3D7_0420900	231.827227	223.1471003	2804.150346	1103.813458	992.4852479	2.873925406	6.743662305	10.7115668	1071.15668	3.67194947	154.37942	4.720336611	7.10189E-05	0.002686794
PF3D7_1206000	14445.2267	13780.86484	18472.14147	3569.682345	3180.704052	330.5014217	487.6040899	1293.767915	1420.383873	10687.97239	883.064325	12.102999	3.597292340	0.00145539
PF3D7_1240400	422.061628	414.353572	3469.542441	1279.987319	1240.569942	11.85423233	16.22037385	20.23098691	1366.06255	14.591083	93.623109	6.548792768	0.000287957	0.001007881
PF3D7_0806000	468.963475	463.7959339	5407.76098	2443.0157017	239.690779	11.49570162	6.322275244	46.33029253	47.20563613	22.604537	77.818809	6.307146166	0.000468356	0.015924115
PF3D7_1219300	454.806087	372.3493771	1833.213471	495.9169111	481.7248876	12.21418298	9.483385865	7.746517136	72.042003	15.771219	45.50222	5.9906942	0.000474563	0.015924115
PF3D7_0200100	95.5623685	108.9482901	345.6416808	345.0845858	264.6693763	26.58381001	33.98213268	35.30534273	44.6767277	6.9326126	2.793399138	0.000596206	0.0019078582	0.000596206
PF3D7_0200300	51.5873627	52.06765673	418.7232664	13.71196842	15.83753055	2.155444054	23.70846466	10.95845125	34.29632611	6.57458108	5.21822115	0.000816663	0.025316558	0.025316558
PF3D7_1100200	82.2899172	80.0704301	892.5430744	369.084833	471.058779	4.310888109	23.70846466	9.62878506	5.90074517	35.4217937	54.6280074	64.841756	0.01125541	0.037662218
PF3D7_0800700	335.453573	328.1408362	226.2474789	188.7308724	188.7308724	8.621776218	7.112539399	22.3995628	17.5330313	14.3804772	12.06726	3.592056849	0.001158917	0.037662218
PF3D7_0600400	74.3528266	74.8199109	883.7926521	447.923017	903.8472516	2.02107376	82.18934417	80.4118381	63.1270028	116.257536	5.4299278	5.478941538	0.001183949	0.037662218
PF3D7_0700100	125.646818	122.9496768	135.6315456	133.619921	102.9439486	17.24355244	37.14326131	38.61993775	36.8497353	38.2838598	44.475581	5.478941538	0.001649873	0.04454657
PF3D7_0700100	11.502877	10.5014001	8.750422298	38.85057719	19.79691319	1.436962303	3.161128622	3.54479751	17.8803661	1.95720111	91.356816	1.91512386	0.002413792	0.0608344797
PF3D7_0425800	148.652573	149.6398202	936.2951859	410.2163888	422.534148	12.21418298	11.06395018	3.75919004	38.7706825	413.4276231	2.3961335	16.94644	0.00279529	0.0608344797
PF3D7_0400400	50.4356945	43.3167005	485.6484375	189.6822298	170.2534534	1.436962303	4.741692933	6.17919004	4.214788949	187.867321	4.41318965	45.343984	5.028292529	0.003173988
PF3D7_0421300	475.157332	442.3563106	5355.258446	1064.962881	1059.794753	17.184813515	7.112539399	5.406791285	9.27255669	1679.50594	7.24416997	231.84243	7.857000791	0.003324137
PF3D7_0432400	101.756226	99.32233679	183.7588683	221.6768228	203.2483087	41.67191839	45.04608286	20.08236769	43.8380498	161.925212	4.0998363	1.943684321	0.006790621	0.142603042
PF3D7_0812400	549.483619	506.6751806	10220.49324	2449.871691	2262.12728	12.93266433	15.01536095	20.08236769	31.8943816	31.977302	19.8094578	161.4611	7.335042787	0.006790621
PF3D7_0800300	42.4721638	39.81644338	166.2580237	67.41717806	56.75115114	17.184813515	7.902821555	6.17919004	16.01619797	74.542992	9.32075577	7.9975266	2.999553882	0.008653462
PF3D7_1240600	1199.83863	1020.788598	17973.3674	2025.943334	2									

Supp. table 15. var genes expression profiles of 3D7\_Ctrl, 3D7\_Wild and 3D7\_ICAM-1 IEs populations

Gene	3D7_Ctrl_1	3D7_Ctrl_2	3D7_Ctrl_3	3D7_Ctrl_4	3D7_Ctrl_5	3D7_Wild_1	3D7_Wild_2	3D7_Wild_3	3D7_Wild_4	3D7_ICAM-1_1	3D7_ICAM-1_2	3D7_ICAM-1_3	3D7_ICAM-1_4	Xtme.rcpt	cbtTRUE	recTRUE	deviance	coverage	padj		
PF3D7_1200400	1085.4216	961.4036537	1503.488753	988.977339	930.867661	14.6523238	28.84406756	28.87614336	36.4489313	69735.92459	69240.5181	115757.3999	70123.93446	10309.4153	-5.33022363	1.1545515	2.4308628	TRUE	0		
PF3D7_0623200	1280.24246	1220.746874	7419.947905	4691.5562933	4823.7289868	28.44455048	89.23534605	98.35564035	111.37106654	84728.73738	8602.646673	1363.598768	6409.1566887	11.922394	-5.56931486	6.2466236	9.57799877	TRUE	1.49227E-07		
PF3D7_0412100	66.36698039	90.63194555	277.0934902	232.8510682	196.3668412	2.791364337	9.020279498	9.020279498	0.985106262	17.569918246	24.74915878	23.24915878	163.65969894	6.3097667	-7.21303062	6.8999407	7.7288235	TRUE	3.60705E-09		
PF3D7_0803000	51.38808005	4.677478341	194.9917153	75.48466894	64.45629137	8.619560753	9.304539722	9.304539722	0	16.23721049	13.8783185	18.6316224	163.65969894	9.82679971	-2.87905783	3.1669227	7.998831	TRUE	7.21190E-09		
PF3D7_0600600	0	3.6674979878	5.13136093	12.79401474	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
PF3D7_0600400	405.6949285	379.3220485	677.3396428	1012.006586	1161.712228	26.63444273	97.86719439	99.26174279	88.65956263	16.29807651	11.96670797	13.2553856	87.04443046	8.70144405	-2.9957139	4.4795703	2.188676E-05	TRUE	0.002199466		
PF3D7_0700100	15.91629410	1.257421672	10.26272186	43.8996301	22.8487528	17.29291151	3.672871316	3.672871316	19.70212801	16.76555856	17.267307136	16.76555856	27.027097657	4.3900680	-3.18928774	3.2801842	3.8483826	TRUE	4.44477E-05		
PF3D7_0576000	58.2424408	378.7928788	631.1579944	852.3801384	977.372553	9.768813319	13.62603663	9.768813319	97.25518859	9.236616063	88.26170482	38.11470038	59.82232154	7.4468526	-3.27495669	2.7395854	1.9978069	TRUE	0.00179634		
PF3D7_1373500	139.3642384	139.3642384	11.28894945	27.9466471	230.508775	7.757604891	13.62603663	9.768813319	97.25518859	9.236616063	88.26170482	38.11470038	59.82232154	7.4468526	-3.27495669	2.7395854	1.9978069	TRUE	0.00179634		
PF3D7_0228300	200.25835345	1.5719436584	318.1443777	11.62574053	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063	9.236616063
PF3D7_0712300	8.56431341	9.95482824	5.13136093	29.42631309	28.48068369	8.149546053	8.149546053	8.149546053	15.7410002	15.7410002	15.7410002	15.7410002	15.7410002	15.7410002	15.7410002	15.7410002	15.7410002	15.7410002	15.7410002	15.7410002	
PF3D7_0429000	17.6518027	199.6158095	1642.035498	655.055544	613.0842598	63.0362572	2.91134371	6.90517019	18.49431257	18.49431257	18.49431257	18.49431257	18.49431257	18.49431257	18.49431257	18.49431257	18.49431257	18.49431257	18.49431257	18.49431257	
PF3D7_0100300	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
PF3D7_1300800	7.49067130	17.289548	63.97007868	4.496590561	5.17136452	5.827272375	10.0718161	9.26174279	294.554702	294.554702	294.554702	294.554702	294.554702	294.554702	294.554702	294.554702	294.554702	294.554702	294.554702	294.554702	
PF3D7_0712300	27.40312409	294.9701673	1016.009464	806.029283	668.5466591	0.861956075	15.81771447	11.74093324	10.83616877	58.10618437	60.74979355	60.74979355	60.74979355	60.74979355	60.74979355	60.74979355	60.74979355	60.74979355	60.74979355	60.74979355	
PF3D7_0500100	33.05402075	280.4002478	354.0639042	51.66781953	431.7072519	12.92934113	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	
PF3D7_0712900	389.638401	371.633191	805.632666	644.8183426	602.5913752	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	
PF3D7_0800100	43.25485817	542.630962	309.457987	611.7605894	437.7031879	86.1954073	186.0907584	63.16656589	216.7323753	216.7323753	216.7323753	216.7323753	216.7323753	216.7323753	216.7323753	216.7323753	216.7323753	216.7323753	216.7323753	216.7323753	
PF3D7_0420700	181.9779785	140.9360124	256.5688465	326.2473757	287.8048359	17.23912151	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
PF3D7_0200300	50.31043288	62.4710154	32.89202356	135.35281768	117.235607	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	3.44782493	
PF3D7_0223300	19.26703282	18.37719939	56.44497023	15.35281768	2.585868226	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
PF3D7_0712000	46.6709731	475.245327	172.4173723	606.4362984	809.451101	1.72391251	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	
PF3D7_0115700	36.3947957	42.4379814	82.10177488	171.6468252	52.4642321	5.17136452	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	
PF3D7_0712400	10.5842974	1075.523753	483.1216553	277.0210332	2515.294347	3.499786694	4.652268961	3.69591319	22.97518754	22.97518754	22.97518754	22.97518754	22.97518754	22.97518754	22.97518754	22.97518754	22.97518754	22.97518754	22.97518754	22.97518754	
PF3D7_1300100	24.9113941	259.3432199	338.6698214	497.6871732	443.699122	5.17136452	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	
PF3D7_0617400	4.28740671	5.763182665	15.36480279	23.2525571	1.969670445	6.29566382	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	
PF3D7_0809100	60.3618319	5797.237835	1239.87125	1630.172021	1642.15023	17.257604762	1.49898352	6.958646602	12.06738505	12.06738505	12.06738505	12.06738505	12.06738505	12.06738505	12.06738505	12.06738505	12.06738505	12.06738505	12.06738505	12.06738505	
PF3D7_0413100	25.9944402	177.0686193	76.9704139	583.929797	583.4070719	1.64262651	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	4.652268961	
PF3D7_0632500	282.5890665	286.63059	0.16631972	242.248512	248.238859	223.2485448	14.92934113	13.02635039	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	
PF3D7_0632800	282.5890665	286.63059	0.16631972	242.248512	248.238859	223.2485448	14.92934113	13.02635039	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	
PF3D7_0100100	103.2162632	158.84652	23.9111239	33.17170965	353.1450284	13.02635039	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	9.87102513	
PF3D7_1206600	43.8714937	1680.15675	21.6614685	308.850093	363.259284	396.4097946	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	3.72181516	
PF3D7_0900100	45.8874187	48.2016411	61.57633116	59.9384881	84.19560753	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	23.2401492	
PF3D7_0400100	15.2007938	147.2311208	159.0721888	149.6899724	116.9207146	20.68849581	22.41085796	9.304539722	30.5385938	65.0230442	36.5487774	3.93439726	37.19299657	11.20284	-3.38820803	1.968259	5.770154	TRUE	0.18781201		
PF3D7_0300100	115.6069981	130.4574985	405.3775135	386.279245	368.749946	31.8923478	40.09251307	41.50945607	52.21063132	20.19393586	87.81935924	6.76755856	23.92868854	8.1317986	-1.46338855	-1.243207	3.675996	TRUE	0.12129966		
PF3D7_0712800	95.26872992	101.641852	92.36449674	131.7783518	128.915827	56.88910097	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	6.652268961	
PF3D7_0412900	36.3947957	27.8006193	41.05088744	78.0438988	73.450122	0	2.791364337	9.0													











Supp. table 21. var gene expression profiles of 3D7\_Ctrl, 3D7\_Wild and 3D7\_CD9 IE's populations

	3D7_Ctrl_3D7_Ctrl_3	3D7_Ctrl_4	3D7_Ctrl_5	3D7_Wild_3	3D7_Wild_4	3D7_Wild_5	3D7_Wild_3	3D7_Wild_4	3D7_Wild_5	3D7_CD9_1	3D7_CD9_2	3D7_CD9_3	X_Intere	rho	TRU	recTRU	deviance	converge	P-value	Padj
PF3D7_0421100	34.220633	48.10303	111.155184	91.616215	0	1.4067511	0.47186487	0.80892581	385.7706511	35.482359	15.7177409	15.7177409	6.40105	-7.1677	5.70556	6.67118	TRUE	1.2E-08	7.9E-07	
PF3D7_0200400	55.972670	51.022578	763.331331	472.10416	434.30266	1.633086	14.536428	15.09966625	18.830022	197.7706511	138.11473	15.7177409	9.09625	-5.3018	3.13332	6.48745	TRUE	2.8E-07	1.9E-05	
PF3D7_0617400	381.390434	364.80448	1341.69159	1065.74684	915.76349	10886.122	11258.698	16985.70894	18972.2453	66088.5703	69860.941	79264.5676	9.66616	-4.16015	3.30413	4.84767	TRUE	3.4E-05	0.00099	
PF3D7_0937600	199.380434	201.30979	320.442845	406.754684	455.92833	130.20367	29.072856	46.6025074	50.3836552	3.31475426	8.3705896	0	8.06674	-2.7333	3.4283	4.84767	TRUE	0.00015	0.00297	
PF3D7_0600400	209.187442	201.30979	343.889883	483.09753	542.00733	130.20367	48.767371	51.90512077	45.80033259	9.94426277	9.94426277	0	8.78887047	-3.5497	3.5497	4.84767	TRUE	0.0002	0.01305	
PF3D7_0800100	93.830769	74.796041	130.261319	185.279406	134.27742	8.4274121	14.536428	13.21210279	6.530636681	53.03606681	54.408833	31.4354819	6.87702	-3.4573	1.24405	2.77371	TRUE	0.00128	0.08187	
PF3D7_0320500	81.687963	70.931884	75.3149805	113.598155	102.80598	3.9293399	6.648384	5.662374843	1.25446295	9.94426277	25.111769	55.0120933	8.34652	-7.211	4.7897	2.21435	TRUE	0.00274	0.17292	
PF3D7_0420700	144.60977	141.80662	166.95011	589.977515	525.91904	1.4067511	1.887458281	4.07140648	129.275416	154.84591	23.57666114	9.26236	-8.0124	5.43682	5.21677	TRUE	0.00383	0.23368		
PF3D7_0712300	141.2981	156.54339	515.834824	384.767944	311.91475	0.4213711	2.3448851	6.134239414	5.59818392	16.5737713	37.667653	15.7177409	3.2627	-0.3874	7.1413	3.9617	TRUE	0.0066	0.36383	
PF3D7_0712400	523.79647	569.72895	2443.70235	1132.31709	1175.5268	1.0685855	1.387458281	1.887458281	2.3448851	25.2032798	41.852948	78.8887047	10.119	-9.0399	3.48701	2.38247	TRUE	0.00068	0.46291	
PF3D7_0100300	910.15846	851.19513	966.538989	2266.46642	2385.51484	68.262117	53.925458	136.3068606	152.168817	9.94426277	12.555884	16.741179	10.12377	-6.9945	2.20736	3.13377	TRUE	0.01042	0.59389	
PF3D7_0712900	200.90800	197.13912	307.814355	281.14284	1.6854844	3.7513362	3.303051992	2.3448851	6.629598851	62.92950851	16.741179	15.7177409	8.12126	-6.9945	2.20736	3.13377	TRUE	0.01184	0.00528	
PF3D7_0809100	3112.4218	3076.6475	6244.72764	7877.36023	7671.2845	7.077968554	10.6874427	29.8327883	25.111769	39.2943524	14.24503	-9.6169	2.1738	2.98551	1.55059	4.01526	TRUE	0.03238	0	
PF3D7_0632800	153.44000	141.80662	247.496507	338.18968	214.00240	3.7923359	7.5026725	6.134239414	14.24503	29.8327883	16.741179	31.4354819	7.63345	-6.6557	1.55059	4.01526	TRUE	0.03238	0	
PF3D7_0100100	1.1038914	1.6683132	0	2.44297108	4.8955249	10.956468	4.6891703	7.077968554	2.03570324	23.2032798	16.741179	31.4354819	1.11303	1.51995	-29.6627	3.0034	TRUE	0.0369	1	
PF3D7_0100100	99.902171	94.815799	117.235187	207.041798	165.04296	63.6276035	62.369565	96.2607324	1.16066665	16.5737713	33.4823259	47.1532228	7.09704	-1.3475	3.4496	3.4496	TRUE	0.04862	1	
PF3D7_0420600	43.60371	46.990821	112.024735	168.565004	134.97656	0.61074277	0.3379668	4.6891703	36.33317191	43.767192	104.63237	78.5887047	6.65703	-1.2509	1.16143	6.12971	TRUE	0.0583	1	
PF3D7_0225500	80.584072	73.961884	161.524036	84.2825021	99.30918	0.4213711	6.0959214	2.831187422	2.03570324	9.94426277	4.1852948	15.7177409	6.63091	-5.1304	1.70706	7.65814	TRUE	0.06878	1	
PF3D7_0500100	240.64852	252.47139	875.356065	289.492072	377.65346	0.8427422	1.87596681	1.415593711	15.2677743	9.94426277	0	15.7177409	8.6677	-8.1709	2.36774	3.89681	TRUE	0.07415	1	
PF3D7_0100100	49.123167	53.942126	46.8940749	62.9065052	60.144996	27.810492	15.943179	23.59322851	38.1694358	82.8688564	50.223358	31.4354819	5.77487	-1.0542	1.09812	2.50156	TRUE	0.09344	1	
PF3D7_1240400	2.63.2781	263.31543	2065.94452	684.031901	659.49688	5.8991953	7.0337554	9.90155976	12.2142195	29.8327883	62.779422	70.7298343	9.61999	-6.489	2.62969	1.82891	TRUE	0.10989	1	
PF3D7_0300100	59.610135	69.234997	205.812884	184.444316	172.04266	15.590731	20.1634321	21.70577023	26.973068	13.259017	0	7.85887047	7.1055	-2.7066	1.5435	6.18122	TRUE	0.11741	1	
PF3D7_0700100	7.1752994	6.6732527	2.01405277	20.7652541	10.490406	0.8427422	1.8756681	0.943729141	1.01785162	8.3705896	0	8.3705896	3.35575	-3.1316	2.05783	4.50499	TRUE	0.13351	1	
PF3D7_0412900	18.766154	14.736766	127.851819	37.2553089	34.268661	1.4067511	4.07186487	1.01785162	3.31475426	3.31475426	0	8.3705896	4.65244	-5.1518	2.13286	6.45798	TRUE	0.1588	1	
PF3D7_0100100	113.14887	114.27945	127.656093	131.920438	107.7015	7.1633086	30.948524	9.90155976	12.7231163	16.5737713	20.926474	55.0120933	6.69288	-2.6995	9.1014	5.9759	TRUE	0.19838	1	
PF3D7_1240900	454.80326	483.81082	4806.64268	1773.507	1598.7379	1.6854844	11.722926	11.32474969	11.1963678	46.4065986	62.79422	15.7177409	8.0323	-7.666	2.2432	7.6417	TRUE	0.19809	1	
PF3D7_0835500	145.71366	153.20676	310.02194	251.626021	273.34992	10.534277	17.4859891	17.4859891	20.8695852	29.8327883	20.926474	39.2943524	7.82128	-3.8265	0.8758	2.90398	TRUE	0.19902	1	
PF3D7_0219300	283.70000	236.62242	1091.58985	265.062362	255.24655	7.1633086	5.6270043	10.82588512	10.785092	8.021697665	5.0892595	62.8709638	8.73495	-5.4728	2.7859	1.60087	TRUE	0.24652	1	
PF3D7_0808700	184.90181	184.07055	2425.46576	321.861439	377.65346	5.0564251	4.2202533	3.684047254	11.7052936	6.629598851	0	6.62159	6.62159	-5.5077	1.53106	3.25144	TRUE	0.25144	1	
PF3D7_0600800	40.843982	37.537046	195.391979	120.927068	100.00854	4.213711	5.1580873	3.774916562	8.14281297	4.1852948	0	4.1852948	3.20503	-0.6155	1.8229	4.38713	TRUE	0.25375	1	
PF3D7_0732800	4.4155656	5.2829917	2.60522638	14.0470837	13.287948	4.213711	5.1580873	3.774916562	8.65173878	13.259017	13.259017	4.88734	4.88734	-1.7618	9.4981	4.86552	TRUE	0.2611	1	
PF3D7_0900100	22.629174	25.580802	31.2627166	40.197655	27.74417	4.213711	5.1580873	3.774916562	6.6695904	23.2032798	12.555884	0	5.41781	-2.9386	1.20277	4.95495	TRUE	0.2748	1	
PF3D7_0800300	26.493394	25.30275	98.9986260	36.0338234	30.072498	4.213711	6.6891703	3.774916562	5.0892595	13.259017	14.852948	23.57666114	6.5839	-8.3315	0.89627	4.255	TRUE	0.28311	1	
PF3D7_0733000	5.6298461	60.893431	118.484097	104.20474	6.3205664	4.6548384	8.965426835	8.965426835	9.8472576	11.79661426	39.2943524	39.2943524	11.4898	-8.5657	2.69774	1.42246	TRUE	0.32344	1	
PF3D7_0420600	748.43837	648.69577	10702.27	1082.84693	1198.7038	2.1068555	9.8472576	11.79661426	5.98183592	16.5737713	20.926474	23.57666114	9.93541	-7.8201	2.22809	1.23304	TRUE	0.32703	1	
PF3D7_1240100	296.39484	281.11077	3188.79709	569.212621	561.58466	4.213711	4.2202533	3.303051992	5.98183592	46.4065986	70.926474	15.7177409	8.12126	-6.713	1.60897	1.87195	TRUE	0.34063	1	
PF3D7_0712800	65.68158	65.342266	492.38778	230.250024	170.64394	1.2641133	2.3448851	1.887458281	2.54462905	6.629598851	4.1852948	7.85887047	7.67608	-6.713	1.60897	1.87195	TRUE	0.34063	1	
PF3D7_0211000	748.43837	676.22294	4879.58902	1203.16525	1242.0641	1.2641133	2.3448851	1.887458281	6.616003554	13.259017	8.3705896	31.4354819	10.7728	-8.911	1.73383	1.61488	TRUE	0.34704	1	
PF3D7_0711000	25.941448	33.088211	28.6574902	7.32891323	8.392325	1.2641133	1.4067511	6.616003554	6.616003554	0	4.35981	-2.3787	1.3947	8.89186	3.06181	1.73383	1.61488	TRUE	0.36181	1
PF3D7_1150400	57.402353	62.283692	778.962689	215.592197	198.61836	4.213711	7.5026725	6.616003554	6.616003554	9.94426277	12.555884	31.4354819	8.03499	-5.1015	1.50195	1.90858	TRUE	0.44088	1	
PF3D7_0632500	660.12706	647.86161	3767.15735	2239.59373	2250.5418	13.905246	45.016035	51.43323816	57.5086166	66.2959851	104.63237	62.8709638	10.9014	-5.1015	0.89636	3.16033	TRUE	0.44291	1	
PF3D7_0400400	31.460905	27.527167	289.180129	101.3833	90.217494	0.8427422	2.8135022	3.774916562	2.54462905	6.629598851	4.1852948	4.1852948	6.74992	-5.4335	1.3006	2.16397	TRUE	0.47775	1	
PF3D7_1300800	3.8636199	9.1757224	0	3.05371384	2.0980813	2.5282266	2.8135022	5.190510273	2.54462905	3.31475426	0	1.98024	1.98024	-0.2716	1.2216	4.46159	TRUE	0.5005	1	
PF3D7_0800200	16.006425	55.87923	36.4731694	10.9933698	12.588488	2.9495977	4.6891703	7.549833124	5.0892581	6.629598851	0	4.14243	-1.8032	1.8452	4.96603	TRUE	0.50635	1		
PF3D7_1200100	51.33095	50.583552	531.466182	197.269914	183.93179	2.5282266	1.4067511	5.662374843	3.56248067	9.94426277	4.1852948	7.85887047	7.66293	-5.9456	1.5882	2.26951	TRUE	0.5179	1	
PF3D7_0400100	222.98606	287.78402	187.5763	244.297108	204.21324	4.213711														

Supp. table 22. Part of whole transcriptome profiles of 3D7\_Ctr1, 3D7\_Wild and 3D7\_CD9 IEs populations

3D7_Ctr1	3D7_Ctr1_3D7_Ctr1	3D7_Ctr1_3D7_Wild	3D7_Ctr1_3D7_CD9	3D7_Wild_3D7_Wild	3D7_Wild_3D7_CD9	3D7_CD9_3D7_CD9	3D7_Wild_3D7_CD9	3D7_CD9_3D7_Wild	3D7_Wild_3D7_Wild	3D7_Wild_3D7_CD9	3D7_CD9_3D7_Wild	3D7_CD9_3D7_CD9	name	ref	coverage	P-value	Padj	function			
PF3D7_0625600	966.4569184	1134.731	700.8058974	142.57442	1296.61422	8.00668081	5.62700435	16.0433954	12.2412045	908.242666	1079.80606	1076.665255	10.1147872	-6.7285012	6.60975	3.32857	TRUE	0	0	poly(A) polymerase PAP, putative	
PF3D7_1109800	2.207782795	2.693011	2.065226384	54.96684919	9.791045895	0	0	0	0	142.534433	83.7058965	133.600798	4.32907034	-31.833484	34.4081	17.0206	TRUE	0	0	small nuclear RNA snR12	
PF3D7_0603900	47.46733069	346.73109	78.1567913	137.705827	144.767072	0.84274219	0	0.47186457	0.50692581	1557.9345	1996.38553	629.907121	7.21319303	-8.3224958	12.0683	11.2279	TRUE	0	0	CD small nuclear RNA	
PF3D7_0725800	47.58475814	64942.162	5134.901203	147.1645776	10419.07155	2.94979767	3.2824192	13.684077	7.30347756	737.3007891	6238.8783	137781.711	14.01006678	-10.7873	13.269	11.1529	TRUE	0	0	5.8S ribosomal RNA	
PF3D7_0510900	121.979994	425.41986	93.78814983	330.698925	98.6981937	1.94959767	0.93758406	1.88745828	3.05354863	991.1115224	1820.6025	1964.77618	7.4510592	-6.3081935	9.49275	8.12117	TRUE	0	0	small nuclear RNA snR04	
PF3D7_0340800	44.15658966	64.230057	62.5243322	90.38992979	34.96802105	0	0	0	0	215.4590266	159.041203	306.4950485	10.866482	-34.621728	36.7645	13.8222	TRUE	0	0	U4 Splecosomal RNA	
PF3D7_0531800	47.1913574	46521.91	5325.082729	15019.99691	10689.02468	2.94959767	0.6891703	16.0433954	13.7409688	72.98473488	64231.7196	136783.6406	14.0062902	-10.957333	13.4263	15.1791	TRUE	0	0	5.8S ribosomal RNA	
PF3D7_0410000	20.42547258	116.405424	119.8405	53.3426289	1051.838073	0.42313711	0	0	0	86.18361064	2887.8543	5249.725476	10.9964986	-11.258729	11.8121	6.3543	TRUE	0	0	unspecified product	
PF3D7_1341000	1496.876755	838.88347	31.26271661	776.864802	8.32238901	0	0	0	0	49.7213383	5895.941275	78.58870474	6.21882709	-36.608631	36.3229	4.91166	TRUE	1.1E-16	6.3E-13	molecular chaperone (Ch (C2))	
PF3D7_0113900	445.4201798	973.73879	630.464785	373.7745745	876.2986076	81.246213	70.3735543	67.004709	65.65142955	1133.645955	1284.88551	2121.895028	9.36653353	-33.149719	4.40998	3.65977	TRUE	7.8E-16	4.4E-12	unspecified product (LSUA)	
PF3D7_0400300	20.42547258	116.405424	119.8405	53.3426289	1051.838073	0.42313711	0	0	0	86.18361064	2887.8543	5249.725476	10.9964986	-11.258729	11.8121	6.3543	TRUE	1.1E-16	6.3E-13	unspecified product (LSUA)	
PF3D7_1341000	1496.876755	838.88347	31.26271661	776.864802	8.32238901	0	0	0	0	49.7213383	5895.941275	78.58870474	6.21882709	-36.608631	36.3229	4.91166	TRUE	1.1E-16	6.3E-13	molecular chaperone (Ch (C2))	
PF3D7_0113900	445.4201798	973.73879	630.464785	373.7745745	876.2986076	81.246213	70.3735543	67.004709	65.65142955	1133.645955	1284.88551	2121.895028	9.36653353	-33.149719	4.40998	3.65977	TRUE	7.8E-16	4.4E-12	unspecified product (LSUA)	
PF3D7_0214300	84.9996376	86.752285	33.867943	55.57759196	39.863544	798.076854	71.81605	1173.99905	1145.083074	40.72131383	37.6676534	31.45348189	1.88834674	-4.9893323	8.27784	10.0584	TRUE	6.8E-12	3.8E-08	small nuclear RNA snR14a	
PF3D7_1103100	96.61085517	163.49469	52.10452768	47.4834246	23.77825432	3.7906876	0.6891703	3.77491656	5.998183915	109.3680994	146.483519	165.062799	6.9264037	-4.4745892	5.4078	11.4873	TRUE	2.2E-12	1.2E-08	conserved Plasmodium protein, unknown function	
PF3D7_0193000	96.61085517	163.49469	52.10452768	47.4834246	23.77825432	3.7906876	0.6891703	3.77491656	5.998183915	109.3680994	146.483519	165.062799	6.9264037	-4.4745892	5.4078	11.4873	TRUE	2.2E-12	1.2E-08	conserved Plasmodium protein, unknown function	
PF3D7_0114000	11.59085967	15.848975	20.84181107	25.65119629	22.37953347	476.570709	350.25261	413.33364	498.283685	6.629508511	8.3705965	15.7174095	4.2559882	-4.5142784	-5.5456	2.02249	TRUE	2E-11	1.1E-07	RESA-like protein with DnaJ domain, putative (GEXPM6)	
PF3D7_0910500	761.1331185	578.07051	416.8362215	107.0783122	543.4030472	52.6713869	60.1451121	20.997765	24.4284389	586.7115032	527.437498	660.1451121	9.1842764	-3.9777901	4.00018	3.2988	TRUE	3.5E-11	2E-07	DNA repair protein REV1, putative	
PF3D7_0814200	376.842100	376.842100	376.842100	1129.26338	1018.268773	18072.1849	18229.6184	29997.804	28729.87985	798.855253	958.432514	1848.843561	11.1550252	-3.31902058	-4.272	7.4288	TRUE	1.1E-10	6.4E-07	DNA/RNA-binding protein, Alpha 1 (ALBA1)	
PF3D7_0914200	740.711276	1094.4134	578.3602573	699.3004703	799.368613	53.092758	67.992692	70.779685	87.02631359	855.5213522	975.173604	1029.512032	9.61275677	-3.896214	3.77458	1.42179	TRUE	2E-10	1.1E-06	phosphatidyl or glycerol acyltransferase, putative	
PF3D7_0918500	205.8757456	306.13547	338.6794	54.96684919	9.791045895	0	0	0	0	157.1074019	91.467797	1210.266053	1.5782819	-2.526474	4.7098	10.4648	TRUE	2.7E-10	1.5E-06	telomerase RNA	
PF3D7_1343000	143.300493	351.2629	216.2337899	254.0689918	298.6268998	6.74193752	5.7834058	25.008222	11.70529384	195.5705011	184.152972	227.972437	8.12076653	-3.993508	3.9572	3.8376	TRUE	3.2E-10	1.8E-06	zinc finger protein, putative	
PF3D7_1210400	20.97793655	24.468293	10.636215	19.705827	82.5425296	458.40685	173.171059	342.015935	366.103673	34.2015905	23.0788389	45.9433959	868.23089	9.3869681	20.8237	1.89	TRUE	5.9E-10	3.3E-06	conserved Plasmodium membrane protein, unknown function	
PF3D7_0218800	475.252466	642.3057	482.8884853	632.7259085	725.2367566	48.457659	49.32288	54.2642556	50.83865524	62.6488554	514.791263	809.463658	9.18994781	-3.5293514	3.68164	1.10911	TRUE	6.2E-10	3.5E-06	ribonuclease P	
PF3D7_0402300	20.97793655	24.468293	10.636215	19.705827	82.5425296	458.40685	173.171059	342.015935	366.103673	34.2015905	23.0788389	45.9433959	868.23089	9.3869681	20.8237	1.89	TRUE	5.9E-10	3.3E-06	conserved Plasmodium membrane protein, unknown function	
PF3D7_0335000	121.428037	94.259694	143.287415	47.0271932	51.0531074	3.79233986	6.09592137	14.519371	5.981890711	808.248266	778.43267	950.923373	10.395624	-4.0489842	3.48884	2.77818	TRUE	2.6E-09	1.6E-05	gamma-tubulin complex component, putative	
PF3D7_1428500	78.3033448	689.29139	270.943544	456.8355911	480.4606093	50.1431603	70.804713	43.4115405	49.58508362	50.6360809	565.014801	605.1130265	9.04455639	-3.3049145	3.40617	2.84913	TRUE	8E-09	4.5E-05	protein kinase, putative	
PF3D7_1011000	38.8619891	11.40014	10.424906554	9.771884301	16.78468014	178.239973	168.34213	246.313306	263.114644	6.629508511	12.5558845	3.38057645	4.360801	-4.8815	5.53168	1.91E-09	5.1E-05	TRUE	9.1E-09	5.1E-05	ribin (RIF)
PF3D7_0924000	145.273025	115.11361	130.26313192	191.595666	147.603873	121.354875	119.104925	136.40725	140.463527	1909.327931	1305.81198	1249.50405	6.4033806	-3.417813	3.31528	0.32225	TRUE	1.1E-08	6.1E-05	putain-like phosphatase, putative	
PF3D7_0421100	34.2206332	48.10303	140.6822247	111.1551839	91.61621516	0	0	1.40675109	0.47186457	39.77705107	33.4823586	15.7174095	6.40104966	-7.1677436	5.70556	6.67118	TRUE	1.2E-08	6.5E-05	erythrocyte membrane protein 1, PREAMP1 (VAR)	
PF3D7_1020000	85.5518329	82.025398	247.4965065	182.6120879	200.7164048	16.433727	12.6670598	16.51526	16.6874202	155.79345	184.152972	165.9362799	7.31057842	-3.8941523	3.57871	4.03142	TRUE	1.2E-08	6.8E-05	cysteine protein (P12p)	
PF3D7_0560800	1346.747673	199.64148	690.33849918	45.80570662	41.96162526	1641.24042	731.041648	1183.98021	1262.13601	39.77705107	138.114729	39.29435237	7.92203145	-2.321007	4.03557	22.167	TRUE	1.7E-08	9.5E-05	native parasite-infected erythrocyte surface antigen, erythrocyte membrane protein, unknown function	
PF3D7_1345900	20.97793655	24.468293	10.636215	19.705827	82.5425296	458.40685	173.171059	342.015935	366.103673	34.2015905	23.0788389	45.9433959	868.23089	9.3869681	20.8237	1.89	TRUE	5.9E-10	3.3E-06	conserved Plasmodium membrane protein, unknown function	
PF3D7_0340400	71.75294083	6.0732527	33.867943	12.82559815	9.09168574	47.935626	120.511676	74.0827275	94.6020075	0	0	0	3.65065683	2.7438796	-3.1985	6.23555	TRUE	1.9E-08	0.0001	circumsporozoite (CS) protein (CSP)	
PF3D7_1020000	85.5518329	82.025398	247.4965065	182.6120879	200.7164048	16.433727	12.6670598	16.51526	16.6874202	155.79345	184.152972	165.9362799	7.31057842	-3.8941523	3.57871	4.03142	TRUE	1.2E-08	6.8E-05	cysteine protein (P12p)	
PF3D7_0924000	145.273025	115.11361	130.26313192	191.595666	147.603873	121.354875	119.104925	136.40725	140.463527	1909.327931	1305.81198	1249.50405	6.4033806	-3.417813	3.31528	0.32225	TRUE	1.1E-08	6.1E-05	putain-like phosphatase, putative	
PF3D7_0421100	34.2206332	48.10303	140.6822247	111.1551839	91.61621516	0	0	1.40675109	0.47186457	39.77705107	33.4823586	15.7174095	6.40104966	-7.1677436	5.70556	6.67118	TRUE	1.2E-08	6.5E-05	erythrocyte membrane protein 1, PREAMP1 (VAR)	
PF3D7_1020000	85.5518329	82.025398	247.4965065	182.6120879	200.7164048	16.433727	12.6670598	16.51526	16.6874202	155.79345	184.152972	165.9362799	7.31057842	-3.8941523	3.57871	4.03142	TRUE	1.2E-08	6.8E-05	cysteine protein (P12p)	
PF3D7_0560800	1346.747673	199.64148	690.33849918	45.80570662	41.96162526	1641.24042	731.041648	1183.9802													



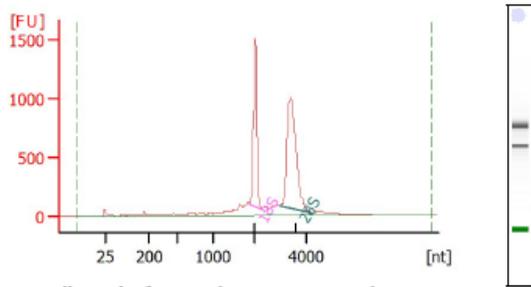


**Supp. table 26. var gene nomenclature of IT4 *P. falciparum* isolate (according to PlasmoDB 28)**

IT4_var08	PFIT_1150900
IT4_var35	PFIT_0536800 (Pseudo)
IT4_var03	PFIT_bin02700
IT4_var02	PFIT_bin08900
IT4_var07	PFIT_1300100
IT4_var18	PFIT_bin10900
IT4_var64	PFIT_0710900
IT4_var22	PFIT_bin01000
IT4_var60	PFIT_bin06900
IT4_var09	PFIT_1400200
IT4_var14	PFIT_bin07000
IT4_var46	PFIT_bin11000
IT4_var10	PFIT_0411300
IT4_var67	PFIT_1240500
IT4_var13	PFIT_0411400
IT4_var19	PFIT_bin09600
IT4_var32A	PFIT_bin00100
IT4_var32B	PFIT_bin00900
IT4_var06	PFIT_bin06100
IT4_var41	PFIT_0900100
IT4_var16	PFIT_bin09100
IT4_var31	PFIT_bin02200
IT4_var44	PFIT_1151000
IT4_var11	PFIT_0500100
IT4_var12	PFIT_bin00300
IT4_var17	PFIT_bin07600
IT4_var29	PFIT_bin02100
IT4_var24	PFIT_bin07650
IT4_var45	PFIT_bin04300
IT4_var33	PFIT_bin04200
IT4_var25	PFIT_0537600
IT4_var61	PFIT_0835600
IT4_var54	PFIT_bin06200
IT4_var63	PFIT_1219000
IT4_var26	PFIT_1400100
IT4_var40	PFIT_0731700
IT4_var20	PFIT_bin10800
IT4_var27	PFIT_0411500
IT4_var58	PFIT_0411350
IT4_var59	PFIT_bin10500
IT4_var21/IT4_var59	PFIT_bin10500
IT4_var15	PFIT_bin10700
IT4_var36	PFIT_0100100
IT4_var39	PFIT_1240000
IT4_var01	PFIT_0616500
IT4_var51	PFIT_bin08300
IT4_var66	PFIT_0710800
IT4_var23	PFIT_0411000
IT4_var05	PFIT_1240400
IT4_var34	PFIT_0811900
IT4_var47	PFIT_1241100
IT4_var62	PFIT_0419300
IT4_var68	PFIT_0710600
IT4_var28	PFIT_0711000
IT4_var30	PFIT_bin11100
IT4_var04	PFIT_1200200
IT4_var65	PFIT_0811500

**Supp. table 27. var gene nomenclature of 3D7 *P. falciparum* isolate (according to PlasmoDB 28)**

PFE1640w**	PF3D7_0533100
PFD1235w	PF3D7_0425800
PF11_0521	PF3D7_1150400
PF13_0003	PF3D7_1300300
PF08_0141	PF3D7_0800200
PF11_0008	PF3D7_1100200
PFD0020c	PF3D7_0400400
PFA0015c	PF3D7_0100300
MAL6P1.314	PF3D7_0600400
PF11820w	PF3D7_0937600
PF08_0140	PF3D7_0800300
MAL6P1.316	PF3D7_0600200
PFL0020w	PF3D7_1200400
MAL6P1.4	PF3D7_0632500
PF11_0007	PF3D7_1100100
PF08_0142	PF3D7_0800100
PFE0005w	PF3D7_0500100
PFA0005w	PF3D7_0100100
PFA0765c	PF3D7_0100100
PFC1120c	PF3D7_0324900
PFD0005w	PF3D7_0400100
PFI0005w	PF3D7_0900100
PF13_0364	PF3D7_1373500
PF07_0139	PF3D7_0733000
PFB1055c	PF3D7_0223500
PF10_0406	PF3D7_1041300
PFL0005w	PF3D7_1200100
PFB0010w	PF3D7_0200100
PFC0005w	PF3D7_0300100
PFL2665c	PF3D7_1255200
PF13_0001	PF3D7_1300100
MAL6P1.1	PF3D7_0632800
PFD1245c	PF3D7_0426000
PFI1830c	PF3D7_0937800
PF10_0001	PF3D7_1000100
MAL7P1.212	PF3D7_0833500
MAL8P1.220	PPF3D7_0700100
PFL0935c	PF3D7_1219300
PFD0635c	PF3D7_0413100
PFL1955w	PF3D7_1240400
PF08_0106	PF3D7_0808700
MAL7P1.50	PF3D7_0712300
PF08_0103	PF3D7_0809100
MAL7P1.55	PF3D7_0712800
PF07_0050	PF3D7_0712400
PFD1005c	PF3D7_0421100
PFL1950w	PF3D7_1240300
MAL6P1.252	PF3D7_0617400
MAL7P1.56	PF3D7_0712900
PF08_0107	PF3D7_0808600
PFD0995c	PF3D7_0420700
PF07_0049	PF3D7_0712000
PFD0630c	PF3D7_0412900
PFD1000c	PF3D7_0420900
PFD1015c	PF3D7_0421300
PFD0615c	PF3D7_0412400
PF07_0051	PF3D7_0712600
PF07_0048	PF3D7_0711700
PFL1960w	PF3D7_1240600
PFD0625c	PF3D7_0412700
PFL0030c	PF3D7_1200600

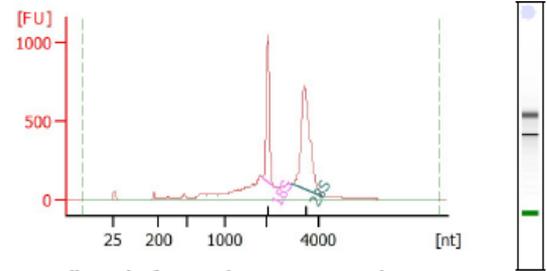


**Overall Results for sample 1 :** Sample 1

RNA Area: 6,641.9  
 RNA Concentration: 11,555 pg/ $\mu$ l  
 rRNA Ratio [28s / 18s]: 1.7  
 RNA Integrity Number (RIN): 9.4 (B.02.08)  
 Result Flagging Color:    
 Result Flagging Label: RIN: 9.40

**Fragment table for sample 1 :** Sample 1

Name	Start Size [nt]	End Size [nt]	Area	% of total Area
18S	1,894	2,313	1,805.1	24.2
28S	2,965	4,180	2,740.9	41.3

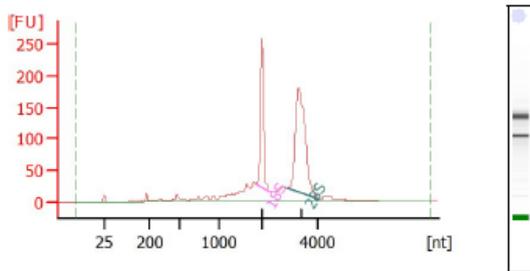


**Overall Results for sample 2 :** Sample 2

RNA Area: 6,351.5  
 RNA Concentration: 11,050 pg/ $\mu$ l  
 rRNA Ratio [28s / 18s]: 1.6  
 RNA Integrity Number (RIN): 8 (B.02.08)  
 Result Flagging Color:    
 Result Flagging Label: RIN:8

**Fragment table for sample 2 :** Sample 2

Name	Start Size [nt]	End Size [nt]	Area	% of total Area
18S	1,928	2,331	1,143.0	18.0
28S	2,989	4,131	1,864.1	29.3

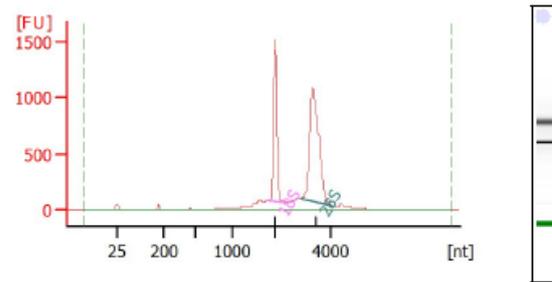


**Overall Results for sample 5 :** Sample 5

RNA Area: 1,545.5  
 RNA Concentration: 3,090 pg/ $\mu$ l  
 rRNA Ratio [28s / 18s]: 1.7  
 RNA Integrity Number (RIN): 8.4 (B.02.08)  
 Result Flagging Color:    
 Result Flagging Label: RIN: 8.40

**Fragment table for sample 5 :** Sample 5

Name	Start Size [nt]	End Size [nt]	Area	% of total Area
18S	1,859	2,248	294.2	19.0
28S	2,908	4,005	507.2	32.8



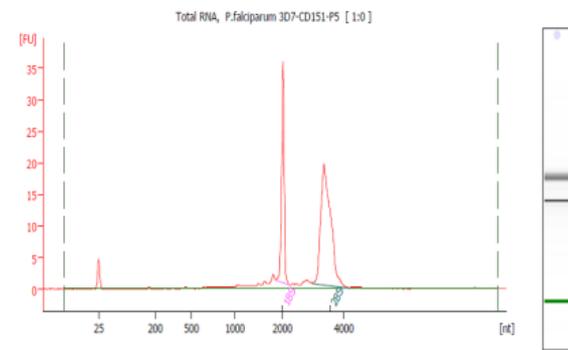
**Overall Results for sample 1 :** Sample 1

RNA Area: 7,137.1  
 RNA Concentration: 15,038 pg/ $\mu$ l  
 rRNA Ratio [28s / 18s]: 1.7  
 RNA Integrity Number (RIN): 9.6 (B.02.08)  
 Result Flagging Color:    
 Result Flagging Label: RIN: 9.60

**Fragment table for sample 1 :** Sample 1

Name	Start Size [nt]	End Size [nt]	Area	% of total Area
18S	1,813	2,278	1,815.5	25.4
28S	2,914	4,091	2,998.2	42.0

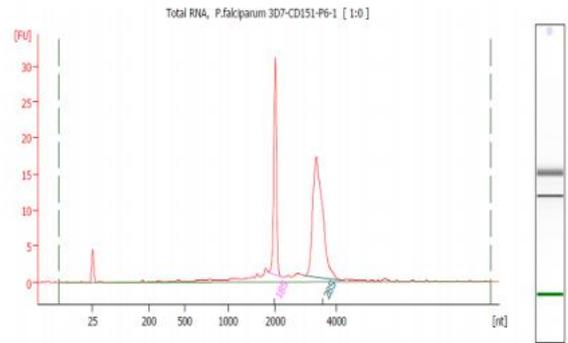
**Supp. fig. 5. Bioanalyzer electropherogram summary.** Examples from total RNA that were isolated from IT4 *P. falciparum* isolate form different IEs populations used in this study.



Overall Results for sample 2 : Total RNA, P.falciparum 3D7-CD151-P5  
 RNA Area: 100.5 RNA Integrity Number (RIN): 9.7 (B.02.07)  
 RNA Concentration: 56 ng/ul Result Flagging Color:    
 rRNA Ratio (28s / 18s): 1.9 Result Flagging Label: RIN: 9.70

Fragment table for sample 2 : Total RNA, P.falciparum 3D7-CD151-P5

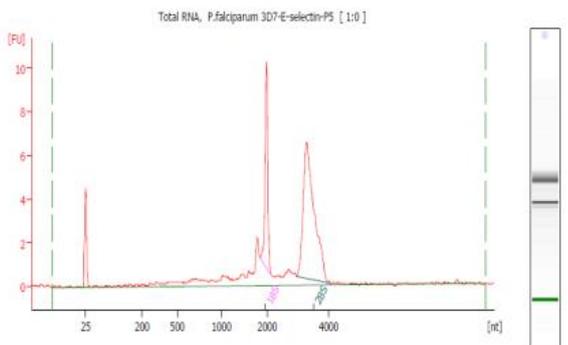
Name	Start Size [nt]	End Size [nt]	Area	% of total Area
18S	1,861	2,213	27.4	27.2
28S	2,963	4,152	50.7	50.5



Overall Results for sample 3 : Total RNA, P.falciparum 3D7-CD151-P6-1  
 RNA Area: 94.0 RNA Integrity Number (RIN): 9.6 (B.02.07)  
 RNA Concentration: 52 ng/ul Result Flagging Color:    
 rRNA Ratio (28s / 18s): 1.9 Result Flagging Label: RIN: 9.60

Fragment table for sample 3 : Total RNA, P.falciparum 3D7-CD151-P6-1

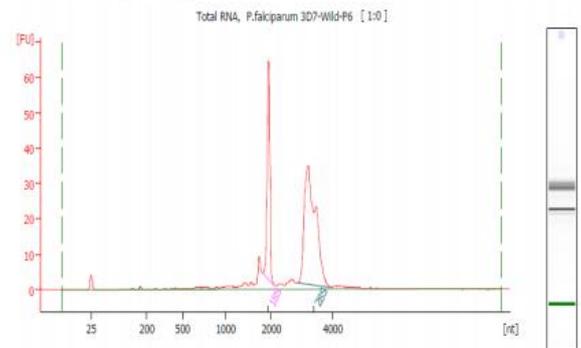
Name	Start Size [nt]	End Size [nt]	Area	% of total Area
18S	1,859	2,212	23.7	25.2
28S	2,978	4,127	44.2	47.0



Overall Results for sample 6 : Total RNA, P.falciparum 3D7-E-selectin-P5  
 RNA Area: 48.8 RNA Integrity Number (RIN): 8.6 (B.02.07)  
 RNA Concentration: 27 ng/ul Result Flagging Color:    
 rRNA Ratio (28s / 18s): 2.4 Result Flagging Label: RIN: 8.60

Fragment table for sample 6 : Total RNA, P.falciparum 3D7-E-selectin-P5

Name	Start Size [nt]	End Size [nt]	Area	% of total Area
18S	1,838	2,139	7.9	16.2
28S	2,951	4,062	18.6	38.2

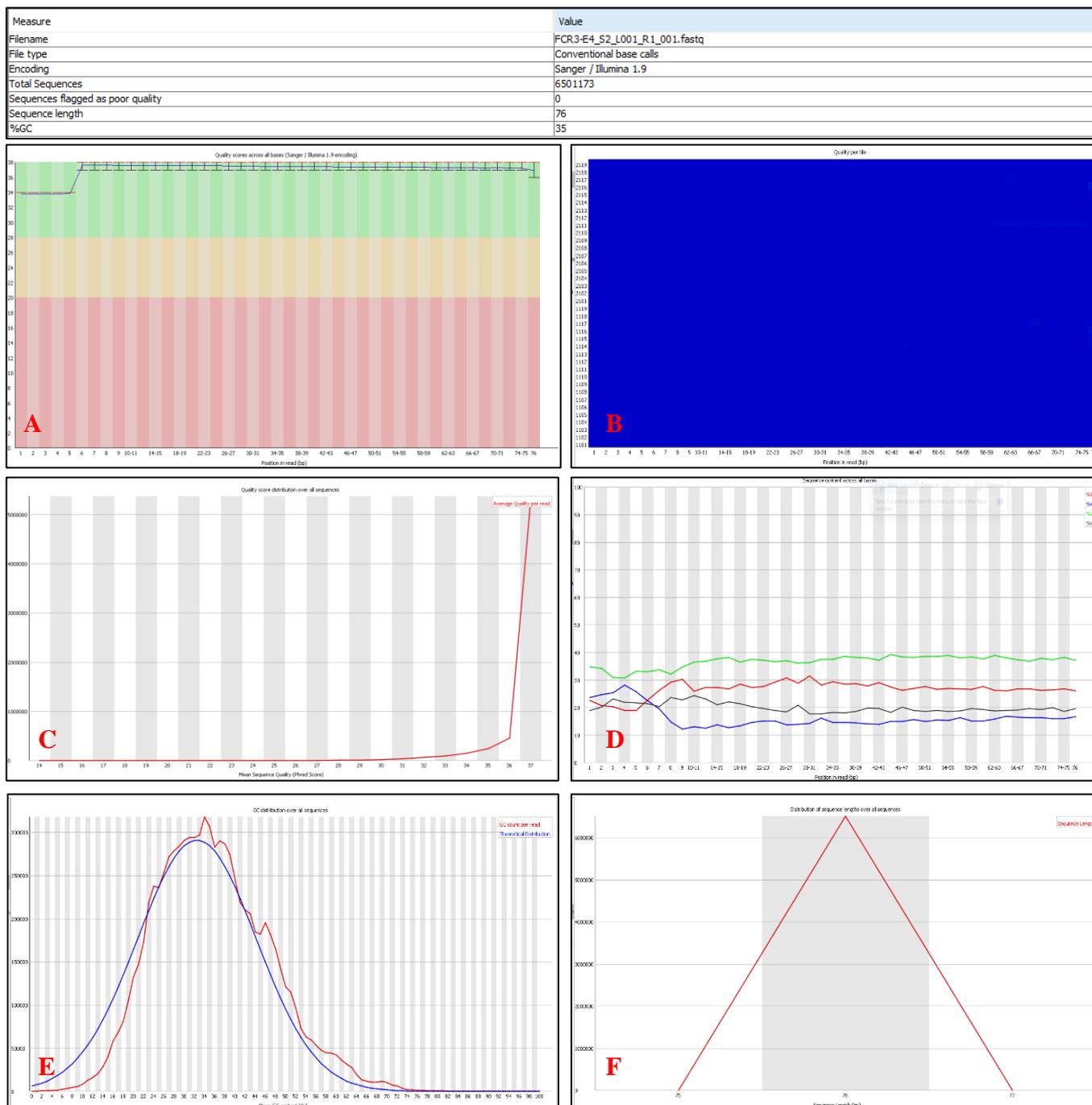


Overall Results for sample 12 : Total RNA, P.falciparum 3D7-Wild-P6  
 RNA Area: 221.6 RNA Integrity Number (RIN): 9.5 (B.02.07)  
 RNA Concentration: 123 ng/ul Result Flagging Color:    
 rRNA Ratio (28s / 18s): 2.1 Result Flagging Label: RIN: 9.50

Fragment table for sample 12 : Total RNA, P.falciparum 3D7-Wild-P6

Name	Start Size [nt]	End Size [nt]	Area	% of total Area
18S	1,796	2,069	50.0	22.6
28S	2,826	3,947	106.0	47.8

**Supp. fig. 6. Bioanalyzer electropherogram summary.** Examples from total RNA that were isolated from 3D7 *P. falciparum* isolate form different IEs populations used in this study.



**Supp. fig. 7. Example of FastQC quality report used in this study to evaluate the NGS runs.**

(A) Showed a plot of the Qscore of the raw sequence reads, as a box-plot for each cycle. The background of the graph is divided according to phred score of 30 into very good quality (green), reasonable quality (orange) and poor quality (red). This run showed very good quality profile with no decay at the end. (B) The graph shows the quality scores from each tile across all the bases and reports any loss in quality associated with only one part of the flow cell. The plot of that run showed a good plot which was blue all over. (C) Shows the per sequence quality score report which showed very high quality in that run. (D) Shows the Sequence content and plots out the proportion of each base position. (E) GC content across the whole length of each sequence compared to a modelled normal distribution of GC content. (F) Shows the read distribution all over the sample. Project website: <http://www.bioinformatics.bbsrc.ac.uk/projects/fastqc>

## 8. Acknowledgements

This thesis is the end of a long and hard journey that would not have been completed without the help of many people (my supervisors, my colleagues, my family and my friends) who were always there when I needed them. I would like to take this opportunity to thank them for helping me over the last three years.

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My beloved daughter **Nadeen Fareed** ♥♥♥ for being with me through my hard times, your laugh, your drawing your jokes as well as your achievements softened my hard days ☺

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Best friends and volley ball team ☺ **Maha, Rahma, Amira, Sara, Wael, Hefny, Ali** and **Ahmed** those good days I will never forget.

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My school friends who form the backbone and origin of my happiness **Raghda, Sara** and **Salama** thanks for the happy time we spent together, your great words and support throughout my life.

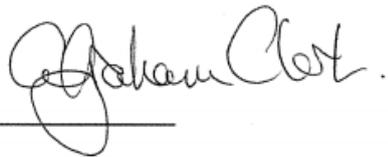
Last but not the least, I would like to thank my **Parents** ♥♥ who always supported, encouraged and believed in me, my brothers **Mostafa** ♥♥ and **Hisham**♥♥ for standing behind me with their love and support.

الحمد لله

Thanks God

## Language Certificate

I am a native speaker, have read the present PhD thesis and hereby confirm that it complies with the rules of the English language.



A handwritten signature in black ink, appearing to read 'Graham Clark', is written above a horizontal line.

Dr. Graham Clark  
Faculty of Infectious and Tropical Diseases  
London School of Hygiene and Tropical Medicine,  
July, 2016